

**INCREASED LYMPHOCYTE CASPASE-3 ACTIVITY
IN PATIENTS WITH SCHIZOPHRENIA**

POVEĆANA AKTIVNOST KASPAZE-3 U LIMFOCITIMA OBOLELIH OD SHIZOFRENIJE

Dušan Lazarević³, Vladimir V. Đorđević¹, Vladan Ćosić², Predrag Vlahović²,
Suzana Tošić-Golubović³, Tatjana Ristić², Vidosava B. Đorđević⁴

¹Clinic for Mental Health Protection, Clinical Centre Niš, Niš, Serbia

²Centre for Medical Biochemistry, Clinical Centre Niš, Niš, Serbia

³Clinic of Psychiatry, Clinical Centre Niš, Niš, Serbia

⁴Institute of Biochemistry, Faculty of Medicine, Niš, Serbia

Summary: A growing body of evidence indicates that cortical brain cells of schizophrenic patients are vulnerable to apoptosis. As apoptosis is an important mechanism in organism modeling during development, active since the early phase of intrauterine life, it could be involved in the pathogenesis of schizophrenia. To test this hypothesis, caspase-3 activity was determined in peripheral blood mononuclear cells from 30 patients with schizophrenia and from 30 age and gender matched healthy subjects by a colorimetric commercially available kit. Consistent with increased susceptibility to apoptosis, caspase-3 activity in lymphocytes of patients with schizophrenia was significantly increased (0.111 ± 0.055 $\mu\text{mol}/\text{mg}$ protein, $p < 0.05$) in comparison with those in the matched control group (0.086 ± 0.030 $\mu\text{mol}/\text{mg}$ protein). The highest activity was obtained in the group showing almost equally positive and negative symptoms (0.159 ± 0.096 $\mu\text{mol}/\text{mg}$ protein) and it was significantly higher ($p < 0.05$) compared to the group with a relative predomination of positive symptoms (0.100 ± 0.029 $\mu\text{mol}/\text{mg}$ protein). Caspase-3 activity in patients receiving typical antipsychotic drugs (0.124 ± 0.071 $\mu\text{mol}/\text{mg}$ protein) was not significantly different from that in patients treated with atypical antipsychotics (0.104 ± 0.039 $\mu\text{mol}/\text{mg}$ protein). To our knowledge to date, this has been the first demonstration that there is a significant increase in caspase-3 activity, determined in native cells, in patients with schizophrenia, indicating a dysregulated apoptotic mechanism in this disease.

Keywords: apoptosis, caspase-3 activity, lymphocytes, schizophrenia

Kratak sadržaj: Sve je više dokaza da su kortikalne ćelije mozga obolelih od shizofrenije osetljive na apoptozu. Kako je apoptoza aktivna od rane faze intrauterinog života, važan mehanizam u modelovanju organizma tokom razvoja, mogla bi biti uključena u patogenezu shizofrenije. U cilju testiranja ove hipoteze određivana je aktivnost kaspaze-3, kolorimetrijskom metodom, u mononuklearnim ćelijama periferne krvi kod 30 obolelih od shizofrenije i 30 zdravih osoba slične starosti i pola. Shodno povećanoj osetljivosti na apoptozu, aktivnost kaspaze-3 u limfocitima obolelih od shizofrenije je značajno veća ($0,111 \pm 0,055$ $\mu\text{mol}/\text{mg}$ proteina, $p < 0,05$) u poređenju sa vrednostima kontrolne grupe ($0,086 \pm 0,030$ $\mu\text{mol}/\text{mg}$ proteina). Najviša aktivnost je dobijena u grupi bolesnika sa skoro podjednako izraženom pozitivnom i negativnom simptomatologijom ($0,159 \pm 0,096$ $\mu\text{mol}/\text{mg}$ proteina) i bila je značajno viša ($p < 0,05$) od vrednosti grupe sa relativnom predominacijom pozitivnih simptoma ($0,100 \pm 0,029$ $\mu\text{mol}/\text{mg}$ proteina). Nije nađena značajna razlika u aktivnosti kaspaze-3 između bolesnika tretiranih tipičnim ($0,124 \pm 0,071$ $\mu\text{mol}/\text{mg}$ proteina) odnosno atipičnim ($0,104 \pm 0,039$ $\mu\text{mol}/\text{mg}$ proteina) anti-psihoticima. Prema našim saznanjima ovo su prvi rezultati koji pokazuju da je aktivnost kaspaze-3 značajno povećana u nativnim ćelijama obolelih od shizofrenije što ukazuje na disregulaciju apoptotičnog mehanizma u ovoj bolesti.

Ključne reči: apoptoza, aktivnost kaspaze-3, limfociti, shizofrenija

Introduction

Schizophrenia is a major chronic debilitating psychiatric disorder associated with social, occupational, behavioral, and cognitive impairment. Besides a strong genetic component (1, 2), the mechanisms playing a role in the pathophysiology of this disease are not

Address for correspondence:

Prof. Vidosava B. Đorđević, PhD
Institute of Biochemistry, Faculty of Medicine
Bul. dr Zorana Đinđića 81, 18000 Niš, Serbia
Tel: ++381 18 4535 666,
e-mail: vidadjordjevic@yahoo.com

known yet. Many individuals manifest the disorder in the absence of genetic loading. A number of human genes, expressed in the brain, are located in chromosomal loci associated with schizophrenia and are potential candidate genes (3, 4). However, none of these gene polymorphisms approached an acceptable genome-wide significance (5).

According to the neurodevelopmental hypothesis, schizophrenia seems to be caused by a combination of factors including genetic vulnerability and environmental factors that occur during individual development (6, 7). However, neurodevelopment does not include important characteristics of schizophrenia such as: protracted period of symptomatic dormancy between the putative insult and the emergence of symptoms, the progressive clinical deterioration that affects at least a subgroup of patients, and progressive neurostructural changes in certain cortical and ventricular brain structures. In order to identify a pathophysiological mechanism that could explain the progressive elements of schizophrenia and its relationship with neurodevelopment, oxidant stress, glutamate excitotoxicity, neurochemical sensitisation and a dysregulation of apoptosis were considered.

Indications of apoptotic mechanism dysregulation include: reduced neuronal densities in the prefrontal cortex (8), reduced incidence rates of cancer (9), negative disease association between schizophrenia and rheumatoid arthritis (10), and absence of evidence of gliosis in the brain tissue of schizophrenics (11). Although much less DNA fragmentation in individuals with schizophrenia than in healthy controls was found (12) the results of Coulson (13) based on the studies of apoptotic nuclei, caspase-3 activity, and protein levels of Bcl-2, Bax and p53^{P392Ser} in dermal fibroblast cell lines suggested significant abnormalities in the regulation of apoptosis in schizophrenia that did not occur in non-schizophrenic psychiatric disorders, and healthy subjects. The exact role of apoptosis in schizophrenia remains uncertain, but the potential involvement of non-lethal localized apoptosis appears plausible (14).

Considering the evidence, the aim of this study was to assess the activity of caspase-3 in peripheral blood mononuclear cells of patients with schizophrenia.

Patients and Methods

Patients were recruited at the Clinic of Psychiatry and the Clinic for Mental Health Protection of the Clinical Centre Niš. The subjects were assessed with the Diagnostic Interview for Psychosis by two independent psychiatrists, and diagnosed according to DSM-IV and ICD-10. The patients with coincidental immune, inflammatory and liver diseases, as well as epilepsy, neurological disorders and mental retar-

ation were excluded from the study. For the disease evaluation and clinical management of the patients the Positive and Negative Syndrome Scales (PANSS) were used. This scale includes three subscales: the positive scale has 7 items, negative also 7 items and the general psychopathology scale includes 16 items. Each patient is tested according to PANSS and each item is scored by two independent psychiatrists. Patients are selected on the basis of total score. The values of total score higher than 3 indicated predominance of the positive syndrome, those lower than -8 indicated predominance of the negative syndrome, and the score values between -8 and 3 indicated a mixed type, i.e. almost equal expression of positive and negative syndromes.

The study included 30 schizophrenic patients (19 males, 11 females; mean age 31.1 ± 8.1) and 30 healthy controls (19 males, 11 females; mean age 29.1 ± 6.9), without any somatic or mental disorder, recruited at the Department for Blood Transfusion, who were matched according to age, gender, marital status, education, living conditions and settings, and habits.

All the subjects provided written informed consent, and the study was approved by the Clinical Centre Niš Ethics Committee.

Venous blood was collected in vacutainer tubes containing potassium EDTA as anticoagulant. Peripheral blood mononuclear cells were isolated immediately from peripheral blood using lymphocyte separation medium (PAA, Pashing, Austria). In brief, the blood was mixed with equal amounts of physiological saline and layered over lymphocyte separation medium. Samples were centrifuged on $400 \times g$ for 30 minutes at room temperature. Separated lymphocytes were collected, centrifuged at $200 \times g$ and washed two times with physiological saline. Lymphocyte suspension was resuspended in $50 \mu\text{L}$ of chilled cell lysing buffer and frozen until caspase-3 activity determination. Before caspase-3 activity analysis resuspended frozen cells were thawed and centrifuged for one minute in a microcentrifuge ($10000 \times g$). Then, the cell supernatant was transferred to a fresh tube, and protein concentration was determined in each sample on an autoanalyzer AU-400 (Olympus, Japan). Further, the assay was performed according to the instructions of the manufacturer. Samples were assayed for caspase-3 activity using a colorimetric commercially available kit (Bio Source Europe S.A, Novellas, Belgium). The caspase-3 colorimetric protease assay is based on the caspase ability to recognize the amino acid tetrapeptide sequence Asp-Glu-Val-Asp (DEVD). The kit includes optimized buffers and substrate, DEVD-pNa, composed of the chromophore, p-nitroanilide (pNa), and a synthetic tetrapeptide, DEVD. After the cleavage of the substrate by caspase-3 during 2 hour incubation at 37°C , the absorbance of pNa is quantified using a

microplate reader at 405 nm. Caspase-3 activity was calculated using the molar extinction coefficient for pNa, and expressed per mg protein.

Data analysis was performed using the SigmaStat computer program. The difference between groups was tested by the Mann-Whitney Rank Sum Test (ANOVA). P values less than 0.05 are presented as significant. Correlations between caspase-3 activity and demographic and clinical characteristics of patients were assessed using Pearson's coefficients.

Results

The demographic characteristics of patients with that schizophrenia and healthy controls are presented in *Table I*. Both groups consisted of the same number of males and females. The age of patients at the first onset of the disease was between 18 and 34 years, and the duration of psychiatric disease was between 6 months and 15 years.

PANSS scores in patients with schizophrenia included PANSS positive score, PANSS negative score, PANSS total score as well as PANSS general psychopathology score for each individual patient involved in this study were presented (*Table II*). It was observed that in 15 patients positive symptoms showed relative predominance, negative ones in 9 patients, and in 6 patients both types of symptoms were almost equally expressed.

Patients with schizophrenia had significantly higher lymphocyte caspase-3 activity (0.111 ± 0.055 $\mu\text{mol}/\text{mg}$ protein; $p < 0.05$) in comparison with the control group (0.086 ± 0.030 $\mu\text{mol}/\text{mg}$ protein) (*Figure 1*). When the patients were divided into groups according to the PANSS scores, the highest activity was observed in the group showing almost equally positive and negative symptoms ($0.159 \pm$

0.096 $\mu\text{mol}/\text{mg}$ protein) and this enzyme activity was significantly higher compared to the group with the relative predominance of positive symptoms (0.100 ± 0.029 mmol/mg protein). Caspase-3 activity in patients receiving typical antipsychotic drugs (TAD) (0.124 ± 0.071 mmol/mg protein) was not significantly different from that of patients treated with atypical antipsychotics (AAD) (0.104 ± 0.039 mmol/mg protein) (*Figure 2*). There was not any significant correlation between lymphocyte caspase-3 activity and heredity ($r = 0.123$), the first onset of the disease ($r = -0.211$) and the duration of psychiatric disease ($r = -0.206$). Also, the values of enzyme activity in male and female patients were not statistically different.

Discussion

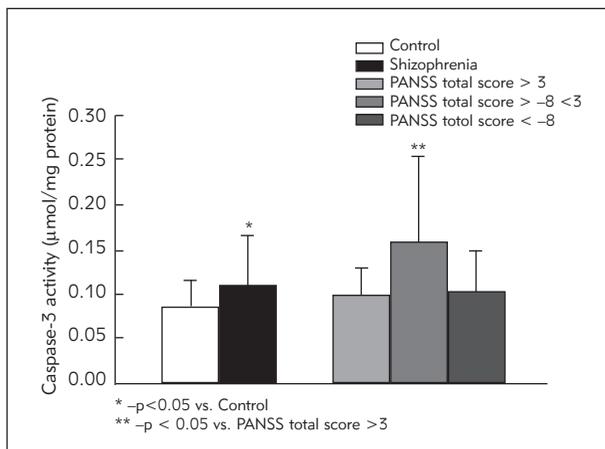
As far as we know, this is the first study undertaken to evaluate caspase-3 activity *in vivo*, in lymphocytes of patients with schizophrenia. Caspase-3 is the main executor caspase capable of degrading a number of cell substrates during activation. This terminal protease can be activated by both apoptotic pathways. In the extrinsic pathway, it is activated by caspase-8, the caspase that is recruited to an activated Fas complex, i.e. through FasL ligation to the Fas receptor that sets in motion a series of events resulting in cell death. In the intrinsic, mitochondrial pathway, which is most frequently implicated in central nervous system apoptosis (15), caspase-3 is activated by the cleavage of caspase-9 (16). The inhibition of caspase-3 activity blocks DNA fragmentation in response to Fas (17, 18), and a variety of other apoptotic stimuli, placing this caspase at a central control point. Mice deficient in two effectors, caspase-3 and -7, died immediately after birth with defects in cardiac development (19). The engagement and activity of apoptotic pathways may favor either cell death or differentiation. During neuronal

Table I Demographic and clinical characteristics of patients with schizophrenia.

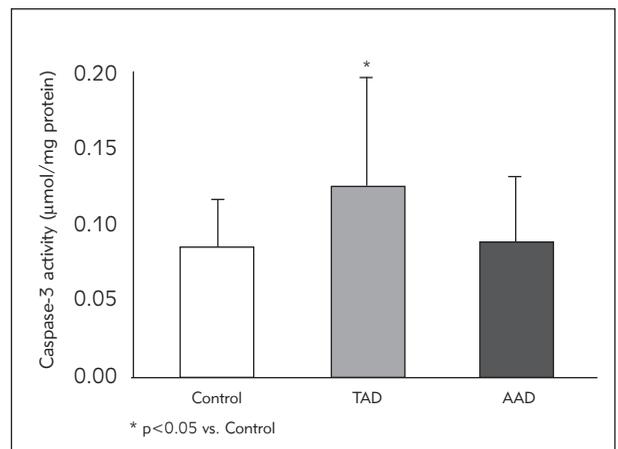
| | | Schizophrenia | Control |
|---|--------------------------------|-------------------|------------------|
| Male/female (N) | | 19/11 | 19/11 |
| Age (years) | | 31.1 ± 8.1 | 29.1 ± 6.9 |
| Heredity (+/-) | | 12/18 | - |
| Patient age at the disease onset | <20 20-24 25-29 30-34 | 10 4 9 7 | - - - - |
| Duration of psychiatric disease (years) | <1 1-3 3-5 >5 | 5 5 8 12 | - - - - |
| PANSS total score | | 91 ± 14 | - |
| PANSS positive score | | 24 ± 7 | - |
| PANSS negative score | | 23 ± 10 | - |
| PANSS general psychopathology | | 48 ± 9 | - |

Table II Individual PANSS scores in patients with schizophrenia.

| Patient | PANSS positive score | PANSS negative score | PANSS total score | PANSS general psychopathology |
|---------|----------------------|----------------------|-------------------|-------------------------------|
| p1 | 13 | 29 | -16 | 50 |
| p2 | 18 | 27 | -9 | 49 |
| p3 | 26 | 22 | +4 | 47 |
| p4 | 17 | 23 | -6 | 46 |
| p5 | 23 | 39 | -16 | 75 |
| p6 | 25 | 11 | +14 | 41 |
| p7 | 35 | 18 | +17 | 48 |
| p8 | 23 | 40 | -17 | 68 |
| p9 | 20 | 16 | +4 | 43 |
| p10 | 31 | 43 | -12 | 51 |
| p11 | 34 | 38 | -4 | 51 |
| p12 | 34 | 34 | 0 | 55 |
| p13 | 21 | 30 | -9 | 42 |
| p14 | 36 | 25 | +11 | 49 |
| p15 | 19 | 40 | -21 | 57 |
| p16 | 25 | 7 | +18 | 49 |
| p17 | 11 | 12 | -1 | 34 |
| p18 | 25 | 13 | +12 | 43 |
| p19 | 15 | 24 | -9 | 56 |
| p20 | 20 | 12 | +8 | 40 |
| p21 | 24 | 15 | +9 | 50 |
| p22 | 31 | 14 | +17 | 54 |
| p23 | 39 | 18 | +21 | 54 |
| p24 | 27 | 16 | +11 | 44 |
| p25 | 19 | 14 | +5 | 31 |
| p26 | 28 | 24 | +4 | 33 |
| p27 | 18 | 23 | -5 | 42 |
| p28 | 31 | 30 | +1 | 52 |
| p29 | 22 | 18 | +4 | 39 |
| p30 | 19 | 30 | -11 | 47 |

**Figure 1** PBMC caspase-3 activity in patients with schizophrenia.

stem cell differentiation into neurons and glia a large percentage of these cells undergoes apoptosis. However, caspase-3 activity is elevated in nonapoptotic differentiating cell populations (20). This finding suggests that the targeted inhibition of this protease in neuronal cells may have unintended consequences. Caspase-3 processing, p53 phosphorylation, and p53 transcriptional activation increased at 3 days of differentiation, with no evidence of

**Figure 2** PBMC caspase-3 activity in patients with schizophrenia treated with typical or atypical antipsychotics.

apoptosis (21) suggesting that apoptosis-associated factors such as caspases and p53 temporally modulate cell signaling and the differentiation of neuronal stem cells. Also, high caspase-3 activity may be involved in the maintenance of synaptic plasticity (22).

As caspase-3 is the end point of both apoptotic pathways we have chosen this enzyme activity to evaluate apoptotic ability *in vivo*, in patients with schi-

zophrenia, and we have shown that caspase-3 activity is significantly increased in the lymphocytes of those patients. Another *in vitro* study based on cultured dermal fibroblast cells showed anomalous apoptotic mechanisms in schizophrenic patients (23) suggesting that altered apoptosis may be observable in all somatic cell types in schizophrenia. Contrary to this finding, Jarskog et al. (24) did not find any significant difference in the level of caspase-3 in a postmortem tissue study of temporal cortex in patients with schizophrenia compared with healthy controls.

In two case-control studies and in one family study the gene of the key glutathione-synthesizing enzyme, glutamate cysteine ligase modifier (GCLM) subunit, is shown to be strongly associated with schizophrenia. GCLM gene expression was decreased in patients' fibroblasts (25). Since oxidative stress is a strong inducer of caspase-3 (26) leading to cellular damage via apoptosis (27) a dysregulation of GSH metabolism may be one of the vulnerability factors contributing to the development of the disease. S-nitrosoglutathione is shown to protect brain dopamine neurons from oxidative stress (28). S-nitrosylation of caspase-3 itself is an important regulator of caspase function (29) in physiological conditions, but the highly significant increase in nitric oxide concentration in schizophrenic patients (30) may act in two different ways: either by increasing caspase-3 S-nitrosylation thereby lowering its activity and cell apoptosis (31), or by initiating apoptosis and elevating the enzyme activity which we showed in this study.

A probable explanation for the decreased cortical volume observed in patients with schizophrenia is reduced neuropil and neuronal size, rather than a loss of neurons. A developmental concept of synaptic apoptosis suggests that the activation of this process may be localized in terminal neurites without neuronal death (14). Synaptic apoptosis represents a potential mechanism of synaptic remodeling and elimination of synapses, in physiological as well as pathological conditions, which may have a direct influence on synaptic plasticity (32). It is thought that in Alzheimer's disease this form of synaptic loss has a direct effect on early cognitive depletion that ends in neuronal death (14). A few studies showed that synaptosome apoptosis might be activated by various pro-apoptotic stimuli (33). The focal application of glutamate to distal dendrites *in vitro*, can cause a localized increase in caspase-3 activity in synaptic terminals without its propagation to the neuronal soma (14). Similarly, β -amiloid induces the localized degeneration of distal hippocampal neurites with apoptotic morphology without the induction of cell death in compartmentalized cell cultures (34).

Previous studies have suggested that the high level of caspase-3 activity in brain tissue postmortem, which frequently accompanies neurodegenerative disorders, is not due to schizophrenia in which it is unchanged or lowered in the temporal cortex of chronic patients (24). However, this does not exclude apoptosis in the early stages of the disease. Jarskog et al. (14) hypothesized that apoptotic activity might contribute to the progressive neurostructural changes in the prodromal stage and the first episodes of psychosis, and cortical neuropil is the main substrate which is damaged by this process. It is supposed that caspase-3 and/or other effector caspases could be sensitive to an activation i.e. to a time limited increase in pro-apoptotic stimuli (oxidative stress, glutamate excitotoxicity) at the beginning of the schizophrenic process. As a consequence, clinical and neurostructural deteriorations could be expressed followed by pro-apoptotic stress depletion and caspase-3 activity normalisation.

On the basis of all known evidence our results may be interpreted in several ways: 1) Elevated caspase-3 activity may be a consequence of increased pro-apoptotic stress. Alternatively, high Bax/Bcl-2 ratio may induce the release of cytochrome c from mitochondria followed by caspase activation. 2) There is a possibility that this finding is a result of antipsychotic drugs acting (35). It is shown that typical antipsychotic haloperidol may induce neuronal apoptosis (36). Others (37) noted that a one-month application of haloperidol, quetapine and clozapine increases caspase-3 activity by 40–50%. Qing et al. (38) found that atypical antipsychotics prevent apoptosis through the modulation of superoxide dismutase 1 expression. 3. In favor of our results is the finding of Miyaoka et al. (39) who showed that the treatment of schizophrenics with minocycline, a caspase inhibitor, led to a significant improvement of the clinical status of patients with schizophrenia, which points to the important role of caspase-3 in the pathogenesis of schizophrenia.

In conclusion, although limited data is available about apoptotic markers and potential inducers of apoptosis *in vivo* obtained in small patient groups with schizophrenia, all of them, including our own results, indicate a dysregulated apoptotic process in schizophrenia.

Acknowledgements. This work was financially supported by the Ministry of Science and Environmental Protection of Serbia.

Conflict of interest statement

The authors stated that there are no conflicts of interest regarding the publication of this article.

References

1. Tsuang MT. Schizophrenia: genes and environment. *Biol Psychiatry* 2000; 47: 210–20.
2. Harrison PJ, Owen MJ. Genes or schizophrenia? Recent findings and their pathophysiological implications. *Lancet* 2003; 361: 417–19.
3. Levinson DF. Molecular genetics of schizophrenia: a review of the recent literature. *Curr Opin Psychiatry* 2003; 16: 157–70.
4. Gregorio SP, Sallet PC, Do KA, Lin E, Gattaz WF, Dias-Neto E. Polymorphisms in genes involved in neurodevelopment may be associated with altered brain morphology in schizophrenia: Preliminary evidence. *Psychiatry Res* 2009; 165: 1–9.
5. Lewis CM, Levinson DF, Wise LH, DeLisi LE, Straub RE, Hovatta I, et al. Genome scan meta-analysis of schizophrenia and bipolar disorder, part II: Schizophrenia. *Am J Hum Genet* 2003; 73 (1): 34–48.
6. Rehn AE, Rees SM. Investigating the neurodevelopmental hypothesis of schizophrenia. *Clin Exp Pharmacol Physiol* 2005; 32: 687–98.
7. Vankatasubramanian G. Schizophrenia is a disorder of aberrant neurodevelopment: A synthesis of evidence from clinical and structural, functional and neurochemical brain imaging studies. *Indian J Psychiatry* 2007; 49: 244–9.
8. Benes FM, Davidson J, Bird ED. Quantitative cytoarchitectural studies of the cerebral cortex of schizophrenics. *Arch Gen Psychiatry* 1986; 43: 31–5.
9. Catts VS, Catts SV, O'Toole BI, Frost ADJ. Cancer incidence in patients with schizophrenia and their first-degree relatives – a meta-analysis. *Acta Psychiatr Scand* 2008; 117: 323–36.
10. Oken RJ, Schulzer M. Schizophrenia and rheumatoid arthritis: the negative association revisited. *Schizophr Bull* 1999; 25: 625–38.
11. Jones EG. Cortical development and thalamic pathology in schizophrenia. *Schizophr Bull* 1997; 23: 483–501.
12. Benes FM, Walsh J, Bhattacharyya S, Sheth A, Berretta S. DNA fragmentation decreased in schizophrenia but not bipolar disorder. *Arch Gen Psychiatry* 2003; 60: 359–64.
13. Coulson E. Cell death and proliferation in mental disorder. PhD Thesis, University of Queensland, 2006.
14. Jarskog LF, Glantz LA, Gilmore JH, Lieberman JA. Apoptotic mechanisms in the pathophysiology of schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* 2005; 29: 846–58.
15. Yuan J, Yankner BA. Apoptosis in the nervous system. *Nature* 2000; 407: 802–9.
16. Sastry PS, Rao KS. Apoptosis and the nervous system. *J Neurochem* 2000; 74: 1–20.
17. Schlegel J, Peters I, Orrenius S, Miller DK, Thornberry NA, Yamin TT, Nicholson DV. CPP32/apopain is a key interleukin 1 beta converting enzyme-like protease involved in Fas-mediated apoptosis. *J Biol Chem* 1996; 271: 1841–4.
18. Hasegawa J, Kamada S, Kamiike W, Shimizu S, Imazu T, Matsuda H, Tsujimoto Y. Involvement of CPP32/Yama(-like) proteases in Fas-mediated apoptosis. *Cancer Res* 1996; 56: 1713–18.
19. Lakhani SA, Masud A, Kuida K, Porter GA, Booth CJ, Mehal WZ, Inayat I, Flavell RA. Caspases 3 and 7: key mediators of mitochondrial events of apoptosis. *Science* 2006; 311: 847–51.
20. Fernando P, Brunette S, Megeney LA. Neural stem cell differentiation is dependent upon endogenous caspase-3 activity. *FASEB J* 2005; 19 (12): 1671–3.
21. Aranha MM, Sola S, Low CW, Steer CJ, Rodrigues CMP. Caspases and p53 modulate FOXO3A/Id1 signaling during mouse neural stem cell differentiation. *J Cell Biochem* 2009; 107: 748–58.
22. Kudriashova IV, Onufriev MV, Kudriashov IE, Guliaeva NV. Caspase-3 activity in the rat hippocampal slices reflects changes in synaptic plasticity. *Russ Fiziol Zh I M Sechenova* 2008; 94 (1): 3–13.
23. Catts VS, Catts SV, McGrath JJ, Feron F, McLean D, Coulson EJ, Lutze-Mann LH. Apoptosis and schizophrenia: a pilot study based on dermal fibroblast cell lines. *Schizophr Res* 2006; 84: 20–8.
24. Jarskog LF, Selinger ES, Lieberman JA, Gilmore JH. Apoptotic proteins in the temporal cortex in schizophrenia: high Bax/Bcl-2 ratio without caspase-3 activation. *Am J Psychiatry* 2004; 161: 109–15.
25. Tošić M, Ott J, Barral S, Bovet P, Deppen P, Gheorghita F, Matthey ML, Parnas J, Preisig M, Saraga M, Solida A, Timm S, Wang AG, Werge T, Cuenod M, Do KQ. Schizophrenia and oxidative stress: glutamate cysteine ligase modifier as a susceptibility gene. *Am J Hum Genet* 2006; 79: 586–92.
26. Labbe D, Teranishi MA, Hess A, Bloch W, Michel O. Activation of caspase-3 is associated with oxidative stress in the hydropic guinea pig cochlea. *Hearing Res* 2005; 202: 21–7.
27. Ristić T, Ćosić V, Vlahović P, Deljanin-Ilić M, Đorđević BV. Could lymphocyte caspase-3 activity predict atherosclerotic plaque vulnerability? *Journal of Medical Biochemistry* 2010; 29 (2): 73–7.
28. Rauhala P, Lin AM-Y, Chieh CC. Neuroprotection by S-nitrosoglutathione of brain dopamine neurons from oxidative stress. *FASEB J* 1998; 12: 165–73.
29. Mannick JB, Schonhoff C, Papeta N, Ghafourifar P, Szibor M, Fang K, Gaston B. S-nitrosylation of mitochondrial caspases. *J Cell Biol* 2001; 154: 1111–16.
30. Djordjević VV, Stojanović I, Stanković-Ferlež D, Ristić T, Lazarević D, Ćosić V, Đorđević BV. Plasma nitrite/nitrate concentrations in patients with schizophrenia. *Clin Chem Lab Med* 2010; 48 (1): 89–94.
31. Saed GM, Jiang ZL, Fletcher NM, Galijasevic S, Diamond MP, Abu-Soud HM. S-nitrosylation of caspase-3 is the mechanism by which adhesion fibroblasts ma-

- nifest lower apoptosis. *Fertility Sterility* 2007; 88: S208–S209.
32. Gilman CP, Mattson MP. Do apoptotic mechanisms regulate synaptic plasticity and growth-cone motility? *Neuromol Med* 2002; 2: 197–214.
 33. Gylys KH, Fein JA, Cole GM. Caspase inhibition protects nerve terminals from in vitro degradation. *Neurochem Res* 2002; 27: 465–72.
 34. Ivins KJ, Bui ET, Cotman CW. Beta-amyloid induces local neurite degeneration in cultured hippocampal neurons: evidence for neuritic apoptosis. *Neurobiol* 1998; 5: 365–78.
 35. Jarskog LF, Gilmore JH, Glantz LA, Gable KL, German TT, Tong RI, Lieberman JA. Caspase-3 activation in rat frontal cortex following treatment with typical and atypical antipsychotics. *Neuropsychopharmacology* 2007; 32: 95–102.
 36. Noh JS, Kang HJ, Kim EY, Sohn S, Chung YK, Kim SU. Haloperidol-induced neuronal apoptosis: a role of p38 and c-Jun-NH2-terminal protein kinase. *J Neurochem* 2000; 75: 2327–34.
 37. German TT, Tong RI, Gilmore JH, Lieberman JA, Jarskog LF. Regulation of apoptosis by typical and atypical antipsychotics in rat frontal cortex. *Biol Psychiatry* 2004; 55: 214S.
 38. Qing H, Xu H, Wei Z, Gipson K, Li XM. The ability of atypical antipsychotic drugs vs. haloperidol to protect PC12 cells against MPP+-induced apoptosis. *Eur J Neurosci* 2003; 17: 1563–70.
 39. Miyaoka T, Yasukawa R, Yasuda H, Hayashida M, Inagaki T, Horiguchi J. Minocycline as adjunctive therapy for schizophrenia: an open-label study. *Clin Neuropharmacol* 2008; 31: 287–92.

Received: July 26, 2010

Accepted: August 27, 2010