

## AGE-DEPENDENT CHANGES IN PLASMA AMINO ACIDS CONTRIBUTE TO ALTERATIONS IN GLYCOXIDATION PRODUCTS

### PROMENE AMINO KISELINA U PLAZMI POVEZANE SA STAROŠĆU DOPRINOSE ALTERACIJAMA U PRODUKTIMA GLIKOKSIDACIJE

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#### Summary

**Background:** Glycative stress is involved in the pathogenesis of various degenerative disorders. This study sought to determine the effect of age-related changes in amino acids on serum levels of pentosidine and carboxymethyl-lysine (CML) in healthy individuals.

**Methods:** The subjects were 78 healthy individuals categorized into three age groups. The ages of the groups were as follows: 26 young adults (20–30 y, 25.2±3.03), 26 middle-aged adults (35–50 y, 39.46±6.97) and 26 older adults (60 y or older, 69.80±10.01). Serum levels of pentosidine and CML were measured by ELISA and levels of plasma amino acids were determined using HPLC.

**Results:** Serum levels of pentosidine and CML in the youngest group were higher than in the oldest group ( $p=0.026$ ,  $0.029$ , respectively). There was a positive correlation between the serum levels of pentosidine and CML and the levels of plasma Tyrosine ( $p=0.032$ ,  $r=0.211$  and  $p=0.037$ ,  $r=0.224$ ), Valine ( $p=0.037$ ,  $r=0.224$  and  $p=0.021$ ,  $r=0.247$ ) and Isoleucine ( $p=0.041$ ,  $r=0.203$  and  $p=0.021$ ,  $r=0.247$ ), respectively. Serum levels of pentosidine and CML may be modulated by the plasma levels of selected amino acids.

**Conclusions:** Better understanding of the role of these selective amino acids might provide new perception of how glycation pathways may be altered and pave the way for new therapeutic principles.

**Keywords:** advanced glycation end product, amino acid, aging, glycation, healthy

#### Kratak sadržaj

**Uvod:** Stres izazvan glikacijom učestvuje u patogenezi raznih degenerativnih bolesti. U ovoj studiji namjera je bila da se utvrdi uticaj promena u amino kiselinama koje su povezane sa starošću na serumске nivoe pentozidina i karboksimetil-lizina (CML) kod zdravih osoba.

**Metode:** Ispitanike je činilo 78 zdravih osoba podeljenih prema starosti u tri kategorije. Starosne grupe su izgledale ovako: 26 mladih odraslih osoba (20–30 god, 25,2±3,03), 26 sredovečnih osoba (35–50 god, 39,46±6,97) i 26 starijih osoba (60 god. i više, 69,80±10,01). Nivoi pentozidina i CML u serumu mereni su tehnikom ELISA a nivoi amino kiselina u plazmi određeni su metodom HPLC.

**Rezultati:** Nivoi pentozidina i CML u serumu u najmlađoj grupi bili su viši nego u najstarijoj grupi ( $p=0,026$ , odnosno  $0,029$ ). Postojala je pozitivna korelacija između serumskih nivoa pentozidina i CML i nivoa tirozina ( $p=0,032$ ,  $r=0,211$  i  $p=0,037$ ,  $r=0,224$ ), valina ( $p=0,037$ ,  $r=0,224$  i  $p=0,021$ ,  $r=0,247$ ) i izoleucina ( $p=0,041$ ,  $r=0,203$  i  $p=0,021$ ,  $r=0,247$ ) u plazmi. Serumski nivoi pentozidina i CML možda modulišu nivoe izabranih amino kiselina u plazmi.

**Zaključak:** Bolje razumevanje uloge ovih izabranih amino kiselina može omogućiti novo shvatanje toga kako se mogu menjati putevi glikacije i otvoriti prostor za nove terapijske principe.

**Ključne reči:** krajnji produkt uznapredovale glikacije, amino kiselina, starenje, glikacija, zdrav

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## Introduction

Aging is an inevitable complex reality that happens to every living organism (1). There are more than 300 theories of aging (2). Over the years, gerontologists have found that aging has no universal cause, phenotype or consequences (3). Aging is a multifactorial process in which genetic, endogenous and environmental factors play a role (4) and is frequently characterized by accumulated damage at the molecular, cellular and organ levels which results in many functional deficits in different body organs, including musculoskeletal, vascular, neural, and endocrine systems (5).

Muscle wasting is an important life-long process with a complex and multifactorial etiology (6). An imbalance between the rate of protein synthesis and protein breakdown can lead to the loss of muscle proteins. The mechanisms involved in this action are not clearly understood (7). Amino acid transportation to the muscular cells is a major determinant in the regulation of protein anabolism (8, 9). The availability of blood amino acids is a potent stimulus for muscle protein synthesis (6, 10). Non-enzymatic modification of ketones or aldehydes and the free amino groups of various proteins lead to the formation of advanced glycation end products (AGEs) (11, 12). AGEs are a group of complex and heterogeneous compounds (13) and accumulation of these products may be amplified by a continuing cycle of metabolic stress, tissue damage, and cell death leading to increased free radical production (14). Recent epidemiological studies demonstrate that elevated circulating AGEs are linked to chronic disease processes through the induction of inflammatory reactions (15, 16). The symptoms of Alzheimer's disease, such as neuronal cell death, amyloid plaque formation, and neuronal tangles may therefore be caused by the glycation of proteins (17).

It has been shown that the amount of AGEs is dependent on the inherent reactivity of specific amino groups, as determined by the glucose concentration and the half-life of the protein (18). We speculated that plasma amino acids can affect serum AGEs, due to alterations in plasma amino acids during the aging process.

Many previous studies have pointed to an increase in the accumulation of AGEs with aging. However, most of these studies have been limited to measuring the amount of AGEs correlated to chronological aging, and did not measure the formation of AGEs with regards to the quantity of their precursors such as amino acids. Since many of these processes in the body are not just dependent on chronological aging, the aim of our study was to (1) determine the age-related alterations of plasma amino acids and serum AGEs and (2) investigate whether any correlation exists between levels of specific amino acids and the formation of AGEs with regards to different age groups.

## Materials and Methods

### *Sampling method*

This cross-sectional study was conducted from January to March 2016. A total of 78 healthy subjects in three age groups were recruited by convenience sampling from the people who attended the public health centers and old people's homes affiliated to Tabriz University of Medical Sciences, Tabriz, Iran. Members of three different age groups were randomly selected in this study as follows: 26 young adults (20–30 years), 26 middle-aged adults (35–50 years) and 26 older adults (over 60 years). All of the participants provided written informed consent to participate in the trial. The protocol was approved by the ethical committee of the Tabriz University of Medical Sciences, Tabriz, Iran (reference number: TBZMED.REC.1394.1032). The exclusion criteria were as follows: those suffering from any disease including chronic diseases and mental illness, regular drug therapy, a history of smoking or a history of alcohol consumption. Each group included an equal number of males and females.

### *Collection of basic characteristics and anthropometric indices*

Body weight was determined to the nearest 0.1 kg on an electronic digital scale while the subjects were wearing light clothing, and height was measured to the nearest 0.1 cm. Body mass index (BMI), defined as the weight in kilograms of the individual divided by the square of the height in meters was then calculated. Waist circumference (WC) was measured as the minimum circumference between the iliac crest and the rib cage. Hip circumference was measured at the maximum protuberance of the buttocks, and the waist-to-hip ratio (WHR) defined as WC (cm) divided by hip circumference (cm).

### *Blood chemistry*

Blood samples were taken in the morning after 12–14 h fasting from the antecubital fossa and divided into two different glass tubes; one was heparinized containing 5.9 mg K<sub>2</sub>EDTA for amino acid measurement and another with a clot activator and gel separator for the measurement of AGEs. The serum and plasma samples were centrifuged for 10 min at 300 g and were then frozen at –20 °C until being thawed for analysis.

### *Determination of amino acids*

Plasma amino acids levels were determined by an HPLC (High-performance Liquid Chromatography) method (KNAUER, Germany), coupled with a fluorescence detector set at excitation/emission of 280/320

nm (KNAUER, Germany). A gradient mobile phase of sodium acetate (0.1 mol/L pH 7.2) and methanol with 70:30% v/v was used to separate the compounds on inertsil ODS-2 C18 (250 mm × 4 mm ID, 5 µm particle size) column. The column temperature was maintained at 40 °C. For internal standardization, homoserine powder was mixed in methanol and added to 200 µL of the prepared sample and 800 µL methanol. The pre-column derivatization was conducted by addition of 50 µL O-phthaldialdehyde (OPA), 100 µL borate and 25 µL HCL. Then, 50 µL of the prepared liquid was added to the mobile phase, and finally an amount of 40 µL was injected into the HPLC system. Flow rate was set at 1 mL/min.

#### Determination of advanced glycation end products

Commercially available kits were used to determine the levels of serum pentosidine or CML (Cat. No. E0004Hu and E1413Hu; Bioassay Technology Laboratory, Shanghai, China, respectively) following the manufacturers' instructions. Briefly, the standards and samples were pipetted into the 96-well pre-coated microplate with an antibody specific for pentosidine or CML, and then incubated at 37 °C. After five washes, avidin conjugated horseradish peroxidase was added to the wells and incubated for 15–25 minutes at 37 °C. Finally, the reaction was stopped and the optical density was measured at 450 nm with an ELISA reader.

#### Statistics

Amino acid and AGEs concentrations in the three age groups were compared by one-way analysis of variance (ANOVA) followed by the Tukey's HSD *post hoc* test. The partial correlation analysis was applied to determine the correlations between amino acids and AGEs. The independent sample *t* test was

performed to compare the means of serum AGEs concentrations in each sex group. A *P* value < 0.05 was considered significant. All of the data were checked for distribution using the Kolmogorov–Smirnov test. Analyses were performed using SPSS software, version 23.0 (SPSS, Chicago, Illinois, USA).

## Results

A total of 78 subjects were included in this study. Demographic and clinical characteristics are presented in *Table I*. The average age of the young, middle and oldest subjects was 25.2±3.0, 39.4±6.9 and 69.8±10.0 years, respectively. These data indicated that the three study groups did not differ with regards to BMI. The mean body weight was 66.88±9.64 kg, 72.95±12.60 kg and 64.11±19.26 kg in each study group, respectively. The younger group had a significantly higher WHR compared to either the middle-aged or oldest groups (*p*=0.039, 0.008 respectively). The dietary intake and physical activity of the subjects were assessed and found not to differ significantly between the study groups (data not shown).

*Table II* shows the normal reference ranges for concentrations of amino acids in the plasma. All of the measured amino acids concentrations were within their reference ranges, except for Arg which was higher than normal range in all three study groups and Met which was lower than normal range in all of the study groups. Asn and Ala were only higher than normal range in the oldest group and Leu was lower in the oldest group. Glu was higher than normal range in the middle group. Alterations in the plasma levels of Asp, Asn, Gly, Thr, Lys, Val, Leu and Ileu were also seen with aging.

*Figure 1* shows the levels of serum pentosidine and CML in the three age groups. The concentration of pentosidine was 3.71±0.63 (ng/mL) in the young group, 3.49±0.57 in the middle group and 3.22±

**Table I** Clinical characteristics of subjects (mean ± SD).

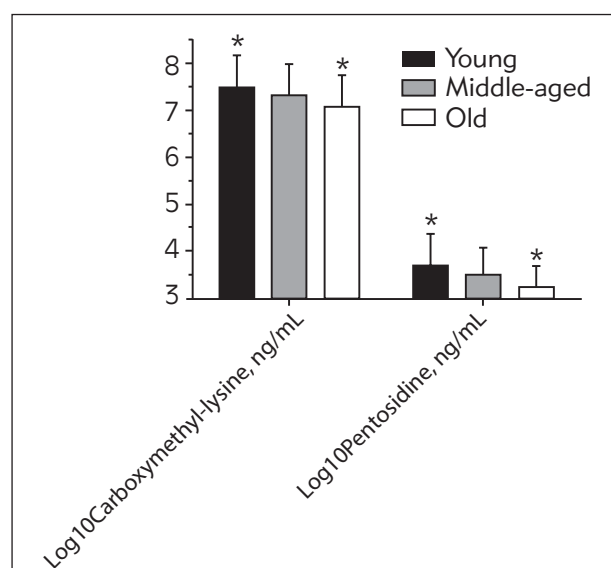
	Group			P. value***
	Young (n=26)	Middle (n=26)	Old (n=26)	
Age (years)	25.20±3.03 <sup>a</sup>	39.46±6.97 <sup>b</sup>	69.80±10.01 <sup>c</sup>	<0.001
Body weight (kg)	66.88±9.64	72.95±12.60	64.11±19.26	0.058
Height (cm)	168.18±9.22 <sup>a</sup>	165.66±10.37 <sup>ab</sup>	161.51±10.98 <sup>bc</sup>	0.043
BMI*	23.75±3.64	26.57±3.64	24.37±6.40	0.059
WHR**	0.85±0.07 <sup>a</sup>	0.90±0.09 <sup>ab</sup>	0.95±0.07 <sup>bc</sup>	0.008

\*Body mass index = weight (kg)/height (m<sup>2</sup>). \*\*Waist to hip ratio=waist circumference (cm)/hip circumference (cm) \*\*\*Based on one-way ANOVA

**Table II** Plasma concentration of amino acids (mean±SEM).

	Young (n=26)	Middle (n=26)	Old (n=26)	p.value*	Normal Range
Asp	17.74±1.55 <sup>a</sup>	12.60±1.22 <sup>b</sup>	12.87±1.39 <sup>bc</sup>	0.017	2–25
Glu	230.07±27.11	257.90±29.55	230.54±19.13	0.681	26–240
Asn	64.96±6.11 <sup>a</sup>	74.29±7.15 <sup>ab</sup>	105.12±14.08 <sup>bc</sup>	0.012	20–80
Ser	144.16±13.02	109.05±12.56	133.48±12.36	0.538	80–230
His	74.24±6.48	75.56±8.31	81.70±8.02	0.764	54–120
Gly	163.21±11.32 <sup>a</sup>	170.27±13.18 <sup>ab</sup>	238.88±12.52 <sup>c</sup>	<0.001	135–350
Thr	106.52±8.47 <sup>a</sup>	101.35±9.03 <sup>ab</sup>	82.09±8.10 <sup>c</sup>	0.001	60–205
Cit	27.19±1.58 <sup>a</sup>	26.23±2.26 <sup>ab</sup>	32.53±1.94 <sup>c</sup>	0.054	10–45
Arg	42.24±5.37	43.03±5.49	43.61±3.57	0.981	12–40
Ala	287.51±24.19	294.25±25.22	336.38±27.50	0.350	123–310
Tyr	62.80±7.37	59.30±5.51	53.39±4.72	0.534	32–85
Met	13.63±1.79	15.04±2.79	16.75±2.41	0.650	28–110
Val	151.41±13.48 <sup>a</sup>	142.33±14.76 <sup>ab</sup>	106.64±10.79 <sup>c</sup>	0.043	60–190
Phe	33.75±3.31 <sup>a</sup>	32.400±3.41 <sup>ab</sup>	43.50±3.60 <sup>c</sup>	0.051	32–85
Ileu	47.84±4.06 <sup>a</sup>	43.98±3.92 <sup>ab</sup>	33.13±3.55 <sup>bc</sup>	0.024	28–110
Leu	79.52±6.64 <sup>a</sup>	72.87±6.96 <sup>a</sup>	52.49±6.18 <sup>bc</sup>	0.013	60–190
Orn	76.42±6.85	78.93±8.10	93.19±7.65	0.244	28–110
Lys	104.64±8.00 <sup>a</sup>	104.36±10.50 <sup>ab</sup>	143.90±11.84 <sup>c</sup>	0.009	80–240

\*Based on Post Hoc, Tukey Test

unit :  $\mu\text{mol/L}$ **Figure 1** Serum carboxymethyl-lysine and pentosidine levels in study groups by age. \* $p < 0.05$ .

0.53 (ng/mL) in the oldest group ( $p = 0.026$ ). Levels of CML were  $7.52 \pm 0.69$  and  $7.32 \pm 0.65$  (ng/mL) in the young and middle-aged groups respectively and decreased to  $7.12 \pm 0.61$  (ng/mL) in the oldest group ( $p = 0.029$ ).

Table III presents the correlations among CML or pentosidine and amino acids using the partial correlation test after adjustment for sex, BMI and WHR. Serum pentosidine was significantly correlated with levels of plasma Tyr ( $r = 0.211$ ,  $p < 0.05$ ), Val ( $r = 0.224$ ,  $p < 0.05$ ) and Ileu ( $r = 0.203$ ,  $p < 0.05$ ). We also found correlations between CML and Tyr ( $r = 0.224$ ,  $p < 0.05$ ), Val ( $r = 0.247$ ,  $p < 0.05$ ) and Ileu ( $r = 0.247$ ,  $p < 0.05$ ). These two AGEs had no significant correlations with the other amino acids that were measured in this study.

Table IV shows the concentrations of AGEs in different sex groups. Serum pentosidine and CML were higher in female than in male individuals ( $p = 0.046$  and  $0.049$  respectively).

**Table III** Correlations among carboxymethyl-lysine, pentosidine and amino acids.

	Log10Carboxymethyl-lysine*		Log10Pentosidine*	
	R	P	R	P
Asp	0.022	0.837	0.018	0.870
Glu	0.191	0.076	0.147	0.174
Asn	0.135	0.212	0.119	0.272
Ser	-0.057	0.599	-0.079	0.469
His	-0.033	0.758	-0.038	0.726
Gly	-0.163	0.132	-0.143	0.187
Thr	-0.005	0.961	0.051	0.636
Cit	0.071	0.515	0.036	0.739
Arg	0.124	0.254	0.079	0.469
Ala	0.122	0.260	0.146	0.178
Tyr	0.224	0.037	0.211	0.032
Met	0.049	0.651	0.057	0.598
Val	0.247	0.021	0.224	0.037
Phe	0.154	0.154	0.156	0.148
Ileu	0.247	0.021	0.203	0.041
Leu	0.156	0.150	0.089	0.412
Orn	0.117	0.279	0.075	0.491
Lys	0.140	0.197	0.127	0.243

\*Pearson analysis\*  
age, sex, BMI and WHR as covariates

**Table IV** Serum AGEs concentration based on sex groups (mean±SD).

Variables	Sex		
	Men (n=39)	Women (n=39)	P. value*
Carboxymethyl-lysine (ng/mL)	3.10±0.24	3.23±0.29	0.049
Pentosidine (ng/mL)	1.43±0.22	1.56±0.25	0.046

\*Based on Independent samples T-test

## Discussion

We demonstrated that concentrations of serum pentosidine and CML had positive correlations with levels of plasma Tyr, Val and Ileu amino acids. It is known that amino acids heated in the presence of reducing sugars can produce AGEs. A decrease in glycosylated proteins and amino acids can lead to a reduction in AGEs. Since the accumulation of AGEs is involved in the pathogenesis of many age-related diseases, these findings may provide the basis for the application of appropriate dietary regimens and food supplements to improve health conditions in the elderly.

These observations on the correlation between levels of selected amino acids and specific AGEs have not been reported elsewhere. Exner et al. (19) reported that the production of p-hydroxyphenylacetaldehyde that causes CML accumulation is barely quantitative at physiological concentrations of tyrosine amino acid (19–21). Furthermore, Lyer et al. (22) have reported that supplementation with curry leaf powder induced a transient reduction in glycosylated proteins and amino acids by reducing the level of total amino acids (22). Overall, it appears that selected amino acids are involved in the specific modulation of glycation. Judging from the lower levels of three plasma amino acids (Tyr, Val, and Ileu) and the concurrent reduction in serum AGEs levels, it is suggested that these selected amino acids are the main precursors in the formation of AGEs during aging. To the best of our knowledge, this is a new finding in the current literature.

We have also determined that the profile of plasma amino acids was affected by the age of the study participants. For instance, plasma Asp, Thr, Leu, Ileu and Val in middle and oldest groups were lower than in the young adults; and levels of Asn, Gly and Lys were higher in the young group as compared with the middle or oldest groups. It is known that health in the elderly could be affected by alteration in the levels of different amino acids. These alterations indicate differences in protein metabolism in the elderly. Indeed, alterations in amino acids during aging might be related to the age-related deterioration in liver function, since enzymes needed for protein metabolism in older individuals possess a lower catalytic ability compared to those of younger individuals (23). In line with our study, Yamamoto et al. (24) reported a correlation between age and plasma amino acid levels in 1890 Japanese individuals. They noticed that the highest plasma amino acid was Cit (24). De Alvaro et al. (25) found that BCAA concentrations reduced and Phe increased with aging. Lawton et al. (26) revealed that Cys, Gly and Pro levels were lower in young adults than in elderly individuals. Menni et al. (27) reported that Val, Met, Ileu, Leu, Phe and Lys were lower in the older subjects than in younger adults. These observations could be explained in part by alterations in steroid hormones such as estrogens and



androgens, as a reduction in the serum levels of these hormones has adverse effects on nitrogen retention which decreases the plasma levels of amino acids (28). Adequate dietary intake, metabolic rate and nutrient interaction within the gastrointestinal systems are also factors that influence plasma amino acid status (29). Thus, careful regulation of amino acid intake may result in a better plasma amino acid balance and in turn regulate many biogenic factors in protein synthesis.

We further compared serum pentosidine and CML between the sexes. We found that women had higher AGEs in comparison to men. This finding was consistent with previously published data. For instance, studies showed higher pentosidine in the serum, urine, skin and cartilage of women (30–32). Van Deemter et al. (33) revealed that pentosidine concentrations were higher in the vitreous base and posterior vitreous of the eyes in women. It can be hypothesized that the hormones involved in protein metabolism, the content of body water or content and distribution of body fat may influence AGE concentrations. Further studies should elucidate whether the accumulation of AGEs is directly related to decreased hormone levels.

In this study, the oldest group had significantly lower serum pentosidine and CML levels than the two younger groups. Our results are surprising, since it has generally been accepted that the formation of AGEs increases in an age-dependent manner. For example, Kandarakis et al. (34) reported that there was a linear age-dependent increase in pentosidine and CML concentrations on Bruch's membranes, choroidal extracellular matrix, and vessel walls. Other studies reported that serum, plasma and cortical bone pentosidine in healthy and non-diabetic subjects increased exponentially during aging by 200% (35–37). Furthermore, Takahashi et al. (38) demonstrated that serum pentosidine was significantly higher in those over the age of 50 than in younger individuals, and that the value for the group aged 80–93 years was threefold that of the group aged 20–29 years. However, there are some contradictory findings too. Rattiste et al. (39) reported that skin pentosidine levels of male common gulls (*Laruscanus*), aging from 2 to 30 years, did not correlate with the chronological

age of the birds. Hein et al. (40) also reported that levels of pentosidine and CML in patients classified into different osteoporosis scales had no correlations with the levels of AGEs or the age of the participants. Knecht et al. (41) revealed a decrease in urine fructoselysine (FL) concentrations with aging, which is the CML precursor. This study indicated that the CML to FL ratio had no correlation with age in either diabetic or control subjects. Serum AGEs are partly exogenously derived from some foods, in addition to their endogenous production by non-enzymatically glycosylated proteins. Intake of higher levels of animal-derived foods and high- and dry-heat processed foods may raise AGEs accumulation in the body (42). The fact that the majority of old subjects in the current study lived in old people's homes that received healthy packaged food daily as opposed to the young subjects who may have received a more unhealthy diet and fast food could be effective in the serum AGEs falling by aging. On the other hand, in line with our study, it seems that AGEs are not only related to changes in physiology, but are also influenced by the availability of specific amino acids (Tyr, Val, Ileu). Thus, regular monitoring of these amino acids and subsequent supplementation may help improve protein synthesis and decrease AGEs accumulation in the elderly.

## Conclusion

In conclusion, there was a correlation between serum AGEs and plasma Tyr, Val and Ileu. Thus, better understanding of the role of these selective amino acids might provide new perception of how glycation pathways may be altered and pave the way for new therapeutic principles.

*Acknowledgements.* The data published was the M.Sc. thesis of the first author. This study was supported by a Grant from the Tabriz University of Medical Sciences.

## Conflict of interest statement

The authors stated that they have no conflicts of interest regarding the publication of this article.

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Received: November 22, 2017

Accepted: January 2, 2018