

**MOLECULAR GENETIC MARKERS AS A BASIS FOR PERSONALIZED MEDICINE**

## MOLEKULARNO-GENETIČKI MARKERI KAO OSNOV ZA PERSONALIZOVANU MEDICINU

Sonja Pavlović, Branka Zukić, Maja Stojilković Petrović

Laboratory for Molecular Biomedicine, Institute of Molecular Genetics and Genetic Engineering,  
University of Belgrade, Belgrade, Serbia**Summary**

Nowadays, genetics and genomics are fully integrated into medical practice. Personalized medicine, also called genome-based medicine, uses the knowledge of the genetic basis of disease to individualize treatment for each patient. A number of genetic variants, molecular genetic markers, are already in use in medical practice for the diagnosis, prognosis and follow-up of diseases (monogenic hereditary disorders, fusion genes and rearrangements in pediatric and adult leukemia) and presymptomatic risk assessment (*BRCA 1/2* for breast cancer). Additionally, the application of pharmacogenomics in clinical practice has significantly contributed to the individualization of therapy in accordance with the patient's genotype and gene expression profile. Genetic testing for several pharmacogenomic markers (*TPMT, UGT1A1, CYP2C9, VKORC1*) is mandatory or recommended prior to the initiation of therapy. The most important achievement of genome-based medicine is molecular-targeted therapy, tailored to the genetic profile of a disease. Testing for gene variants in cancer (*BCR-ABL, PML/RAR $\alpha$ , RAS, BCL-2*) is part of the recommended evaluation for different cancers, in order to achieve better management of the disease. The ultimate goal of medical science is to develop gene therapy which will fight or prevent a disease by targeting the disease-causing genetic defect. Gene therapy technology is rapidly developing, and has already been used with success. Although medicine has always been essentially »personal« to each patient, personalized medicine today uses modern technology and knowledge in the field of molecular genetics and genomics, enabling a level of personalization which leads to significant improvement in health care.

**Keywords:** gene therapy, molecular diagnosis, molecular genetic markers, molecular-targeted therapy, personalized medicine, pharmacogenomics

**Kratik sadržaj**

Genetika i genomika su danas potpuno integrisane u medicinsku praksu. Personalizovana medicina, poznata i kao medicina zasnovana na genomu, koristi znanja o genetičkoj osnovi bolesti da bi se individualizovalo lečenje svakog pacijenta. Veliki broj genetičkih varijanti, molekularno-genetičkih markera, već se koristi u kliničkoj praksi za dijagnozu, prognozu i praćenje bolesti (monogenska nasledna oboljenja, fuzioni geni i reorganizirani u pedijatrijskim i adultnim leukemijama) i presimptomatsku procenu rizika od obolevanja (*BRCA1/2* za kancer dojke). Osim toga, primena farmakogenomike u kliničkoj praksi značajno je doprinela individualizaciji terapije u skladu sa genotipom i profilom ekspresije gena pacijenta. Genetičko testiranje za nekoliko farmakogenomičkih markera (*TPMT, UGT1A1, CYP2C9, VKORC1*) obavezno je ili se preporučuje pre započinjanja terapije. Najvažniji doprinos medicine zasnovane na genomu je ciljana molekularna terapija, prilagođena genetskom profilu bolesti. Testiranje genetičkih varijanti u malignim oboljenjima (*BCR-ABL, PML/RAR $\alpha$ , RAS, BCL-2, KIT, PDGFR, EGF*) doprinosi tačnijoj stratifikaciji različitih kancera i adekvatnom izboru terapije. Krajnji cilj medicinske nauke je da primeni gensku terapiju koja bi eliminisala uzrok bolesti ili prevenirala bolest, ciljajući genetički defekt koji leži u osnovi bolesti. Tehnologija koja prati gensku terapiju veoma se brzo razvija i već se uspešno primenjuje. Iako je medicina oduvek suštinski bila »personalizovana«, prilagođena svakom pacijentu, personalizovana medicina danas koristi modernu tehnologiju i znanja iz oblasti molekularne genetike i genomike, omogućujući stepen personalizacije koji vodi ka značajnom napretku medicinske prakse.

**Ključne reči:** genska terapija, molekularna dijagnostika, molekularno-genetički markeri, ciljana molekularna terapija, personalizovana medicina, farmakogenomika

Address for correspondence:

Sonja Pavlović  
Laboratory for Molecular Biomedicine  
Institute of Molecular Genetics and Genetic Engineering  
University of Belgrade  
Vojvode Stepe 444a, Belgrade, Serbia  
fax: +381 11 3975 808  
phone: +381 11 3976 445  
e-mail: sonya@sezampro.rs

## Personalized medicine

Nowadays, it is thought that virtually all human diseases, except perhaps trauma, have a genetic component. Genetic information is stored in the DNA molecule. Certain portions of DNA are unique to each individual. Any two unrelated people are 99.9 percent identical at the genetic level, with 0.1% being different and making us all individuals (genetic variation). Genetic variation influences every aspect of human physiology, development, and adaptation. Consequently, understanding human genetic variation could play an important role in promoting health and combating disease.

Fascinating recent developments in molecular genetics, especially the improvement in modern technology for human genetic profiling, as well as growing knowledge regarding the genetic base of diseases, have led to the introduction of the principles of personalized medicine in clinical practice.

Personalized medicine principles aim to reach an individualized treatment for each patient. These principles, shared by medical genetics and genomics, include the use of genetic variants as markers for diagnosis, prognosis and prevention, as well as targets for treatment (1) (Table I).

Personalized medicine is frequently called genome-based medicine. It is »a form of medicine that uses information about a person's genes, proteins, and environment to prevent, diagnose, and treat disease« (2). It is defined as »any clinical practice model that emphasizes the systematic use of preventive, diagnostic and therapeutic interventions that use genome and family history information to improve health outcome« (3).

There has long been interest in personalizing medicine. Hippocrates individualized diagnosis and treatment, for example, by giving cold food to a »phlegmatic« person (4). Personalized genomics follows several decades of scientific discovery and clinical translation in human genetics. Genetic analyses have been used in medicine for years. Genetics examines individual genes and their effects as they relate to diseases. Single gene diseases include thalassemia, phenylketonuria and cystic fibrosis. However, even these monogenic hereditary disorders can be influenced by other, modifier genes.

Genomic and personalized medicine aim to tackle more complex diseases, such as cancer, heart disease and diabetes. It is now well-known that these diseases have a strong polygenic background. Therefore, they can be better understood using a whole-genome approach.

High-throughput analysis of the whole genome (the complete set of DNA within a single cell of an organism), comprising the DNA sequencing analysis and functional genomic analysis (mainly concerned with the patterns of gene expression during various conditions), opened the door wide for personalized

medicine. The application of genomics in clinical practice is the best example of successful translational research, the research that aims to move »from bench to bedside« or from laboratory experiments through clinical trials to point-of-care patient applications.

## Molecular genetic markers

Molecular genetic markers represent one of the most powerful tools for the analysis of genomes and enable the association of heritable traits with underlying genomic variation. Availability of a wide array of molecular genetic markers offers tools for quick detection and characterization of genetic variation. Two forms of DNA sequence-based markers, single nucleotide polymorphisms (SNPs) and simple sequence repeats (SSRs), predominate in modern genetic analysis (5). The most studied molecular genetic markers, SNPs, are distributed over the whole genome. The number of SNPs is estimated to range from 0.5 to 1 SNP per 100 base pairs (bp). Besides SNPs, there are other important classes of genetic variants frequently used as molecular genetic markers, such as VNTRs (variable number of tandem repeats, a polymorphic sequence containing 20–50 copies of 6–100 bp repeats), STRs (short tandem repeats, also known as SSRs or microsatellites, a subclass of VNTR in which a repeat unit consists of only 2–7 nucleotides) and CNP (copy number polymorphisms, variation in the number of copies (CNV) of a DNA sequence in the >1 kb size range, which are common and widely distributed in the human genome) (6).

DNA sequence-based markers may affect levels and patterns of gene expression. The amount of transcript of each gene is treated as a phenotypic trait, since it reflects changes in protein function more reliably than DNA markers. Gene expression profiling represents a potent tool for exploring functional genetic variation using RNA molecular genetic markers (7).

The systematic study of protein structures, post-translational modifications, protein profiles, protein–protein, protein–nucleic acid, and protein–small molecule interactions, and the spatial and temporal expression of proteins in eukaryotic cells, are crucial to understanding complex biological phenomena. Proteins are essential to the structure of living cells and their functions. However, the technology for protein profiling is still very expensive and time consuming. Therefore, protein-based molecular markers are not widely used yet (8).

## High-throughput methodology for genome-wide genetic and gene expression profiling

There are several approaches for the comprehensive analysis of the genetic profiles of a large

**Table 1** Major genes and associated molecular genetic markers applied in personalized medicine.

| Gene/molecular genetic marker       | Disease  | Application                                       | Reference      |
|-------------------------------------|--|---|----------------|
| HBB                                 | thalassemia  | molecular diagnosis                               | 23, 26, 27     |
| PAH                                 | phenylketonuria                                      | molecular diagnosis                               | 24, 28         |
| CFTR                                | cystic fibrosis                                      | molecular diagnosis                               | 25, 29         |
| A1AT                                | alpha-1 antitrypsin deficiency                       | molecular diagnosis                               | 70             |
| HLA                                 | celiac disease                                       | molecular diagnosis<br>preventive medicine        | 74             |
| <i>t(9;22)(q34;q11) – BCR/ABL</i>   | ALL<br>CML   | risk stratification<br>molecular-targeted therapy | 37<br>83       |
| <i>t(4;11)(q21;q23) – MLL/AF4</i>   | ALL  | risk stratification                               | 37             |
| <i>t(12;21)(p13;q22) – TEL/AML1</i> | ALL  | risk stratification                               | 37             |
| <i>t(1;19)(q23;p13) – E2A/PBX1</i>  | ALL  | risk stratification                               | 37             |
| <i>BRCA 1/2</i>                     | breast, ovarian, prostate<br>and pancreatic cancers  | preventive medicine<br>molecular-targeted therapy | 57, 58,<br>107 |
| <i>TPMT</i>                         | ALL, IBD, transplantation<br>medicine                | pharmacogenomics                                  | 47, 48         |
| <i>UGT1A1</i>                       | Gilbert syndrome                                     | molecular diagnosis<br>pharmacogenomics           | 46             |
| <i>VCORC1</i>                       | thrombosis and<br>thromboembolism                    | pharmacogenomics                                  | 43, 44         |
| <i>CYP2C9</i>                       | thrombosis and<br>thromboembolism                    | pharmacogenomics                                  | 45             |
| <i>PML/RAR<math>\alpha</math></i>   | APL  | molecular-targeted therapy                        | 89–95          |
| <i>RAS</i>                          | various human<br>cancers                             | molecular-targeted therapy                        | 103            |
| <i>BCL-2</i>                        | AML  | molecular-targeted therapy                        | 104–106        |
| <i>KIT</i>                          | sarcoma, glioma, melanoma,<br>liver and renal cancer | molecular-targeted<br>therapy                     | 107            |
| <i>PDGFR</i>                        | sarcoma, glioma, melanoma,<br>liver and renal cancer | molecular-targeted<br>therapy                     | 107            |
| <i>EGFR</i>                         | lung cancer, glioblastoma                            | molecular-targeted therapy                        | 107            |
| <i>BRAF</i>                         | melanoma   | molecular-targeted therapy                        | 107            |
| <i>HER2</i>                         | breast cancer  | molecular-targeted therapy                        | 107            |
| <i>ADA</i>                          | SCID (ADA deficiency)                                | gene therapy                                      | 113–118        |
| <i>LPL</i>                          | LPL deficiency                                       | gene therapy                                      | 119–122        |

number of people which have provided sufficient data on molecular genetic markers that may be used in the diagnosis, prognosis and treatment of certain diseases (9). The best known are platforms for DNA analyses (DNA microarrays, genotyping arrays, SNP arrays, Next-generation sequencing) and hybridization platforms for the analyses of gene expression, or the amount of transcribed messenger RNA (hybridization microarrays, expression profiling) (10). A special kind of studies, GWAS (genome-wide association study) analyses, have contributed to the implementation of personalized medicine in clinical practice, analyzing a large number of genetic markers in different individuals suffering from the same disease. GWAS analyses establish the relationship of molecular genetic markers with the pathological phenotype (11). Biomedical professionals are keen to understand the personal genetic profile of every person. Sequencing the complete genome is therefore imposing as the ultimate genetic test. It can be performed once in a lifetime, as early as possible, and the data can be used throughout life, with the aim to achieve better health and longer life using the principles of preventive and personalized medicine (12). It is important to note that for all these methods, bioinformatics data processing has a significant role.

Several international projects have contributed to the development and permanent improvement of methodology for a comprehensive analysis of genetic profiles. Biobanks, the repository of human genetic material, as the major outcome of these projects, provided a sufficient number of samples for comprehensive studies. The Human Genome Project was completed in 2003. Its main achievement is the information on the first sequence of the entire human genome. Results of this research allowed a better understanding of the structure, organization and variability of the human genome, and also became the basis for the study of normal and abnormal gene functioning (13). International HapMap Project has identified the most common genetic variants in the human populations, which are later used in designing genotyping platforms (14).

DNA microarrays, known also as DNA chips, are used for detection of a large number of SNPs in populations and differences between patients and healthy controls. Gene expression profiling platforms use the same technology, except the starting material that is analyzed is total RNA of an individual (10).

Comparative Genomic Hybridization (CGH), as a step ahead of cytogenetic analysis and standard FISH analysis, is a molecular-cytogenetic method that detects copy number variants, very common and very heterogeneous in human DNA (15).

The most accurate method, the so-called gold standard, for determining nucleotide changes in DNA is the sequencing analysis. The sequencing method »reads« the DNA nucleotide by nucleotide. Automatic sequencing, based on the Sanger method, was absolutely dominating in genetics (16), but the need for

accurate genetic information to be obtained quickly and cheaply was a catalyst for a fundamental shift in the sequencing technology. Nowadays, there are various platforms for new »next-generation sequencing« technologies that are based on different strategies and are able to produce very large amounts of data. Today, using this methodology, up to 500 000 different DNA samples can be sequenced in one step (17–19).

Sequencing of the complete genome provides 3000 times more information than the platforms for the analysis of DNA variants that exist today. It is a method that allows the analysis of all the genes and regulatory sequences in an individual (20, 21). Comparison of genomes analyzed in two groups (patients and healthy controls) would contribute to a true understanding of the genetic basis of certain diseases.

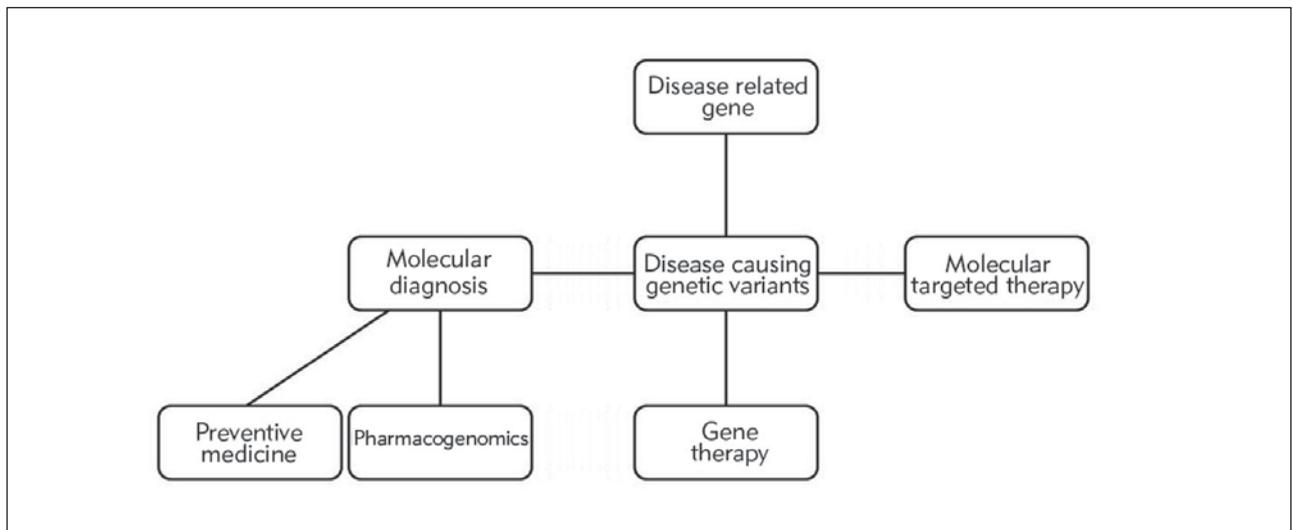
Sequencing of the complete genome does provide information on complete DNA in the genome of an individual. However, our knowledge is not sufficient to understand how to use this information in clinical practice and preventive medicine. Nowadays, many studies are devoted to the bioinformatics analysis of data obtained from genomic sequences and their possible applications in medicine. Sequencing of the entire genome of each newborn baby and monitoring its health until old age could give valuable information on the genotype–phenotype association for the future of medicine, personalized medicine.

Genome-wide association studies (GWAS) use modern methodology (next generation sequencing technologies, as well as expression profiling platforms) to examine the presence or absence of thousands or millions of genetic variants in the genomes of different individuals that have the same disease and compare it with genetic variants in the genomes of healthy individuals, with the aim to determine the associations of certain genetic variants with normal or pathologic conditions. DNA profiles from a group of healthy individuals are compared with DNA profiles from a group of patients carrying a certain disease. If a genetic variant is more frequent in the group of patients, then it could be an attribute of the disease and should be considered as a diagnostic, prognostic or targeted therapy marker (11).

In January 2008, the National Institute of Health (NIH), USA, decided to combine all the available GWAS studies and to put up their results for public health care usage. Thousands of people were tested for over 200 diseases in 1200 GWAS studies till the end of 2011, and over 4000 genetic variants associated with different diseases were discovered (22).

### **Molecular genetic markers and health care strategies**

Study of the genetic basis of different diseases and the analysis of a number of human genome-wide profiles have led to the implementation of the principles of personalized medicine in clinical practice.



**Figure 1** Molecular genetic markers and health care strategies.

Identification of disease related genes and disease causing genetic variants (molecular genetic markers) enables accurate diagnosis, prognosis and follow-up of the disease. It is also a basis for design of the strategies that minimize risk for developing the disease (preventive medicine) and establishment of the guidelines for using therapeutics according to a person's genotype (pharmacogenomics). The final achievement of the study of molecular genetic markers is the implementation of therapeutic approaches that »repair« the affected genes (gene therapy) and design of molecular therapeutics which target the biological mechanism that causes the disease (molecular targeted therapy).

There are several health care strategies based on the application of molecular genetic markers, such as: molecular diagnosis, prognosis and follow-up of the disease, predictive genetics, pharmacogenomics, molecular-targeted and gene therapy (Figure 1).

### **Molecular genetic markers in diagnosis, prognosis and follow-up of disease**

The association of molecular markers with human diseases has led to the identification of genes and genetic mutations responsible for many heritable diseases such as thalassemia, phenylketonuria, cystic fibrosis, etc. Thalassemias, the most frequent hereditary disorders in the world, are characterized by genetic defects in one or more globin genes which impair the synthesis of hemoglobin's polypeptide chains (23). Phenylketonuria is a metabolic disease inherited in an autosomal recessive fashion. PKU is caused by mutations in the human phenylalanine hydroxylase gene which affect the structure and/or function of the phenylalanine hydroxylase enzyme, thus decreasing catabolism of L-phenylalanine (24). Cystic fibrosis is an autosomal recessive genetic disorder caused by a mutation in the gene for the protein cystic fibrosis transmembrane conductance regulator (CFTR). It represents the most common genetic disorder among Caucasians (25).

The characterization of the most common mutations causing hereditary disorders has created the basis for screening, counseling and first trimester prenatal diagnosis.

Also, better definition of the genetic basis of the most common genetic disorders in the Serbian at risk population has improved the strategy for screening prospective parents and making prenatal diagnosis. Data on molecular genetic markers of the most common genetic disorders are the result of an over 20-year systematic survey in Serbia (26–29).

Molecular genetic markers have been proven to be crucial in the diagnosis of single gene disorders. However, genetic profiles nowadays have diagnostic, prognostic and therapeutic applications in several fields, especially cancer care. One of the most prominent examples for the role of genetic profiling in oncology is the detection of fusion genes and rearrangements in pediatric leukemia (30).

Acute lymphoblastic leukemia (ALL) is one of the most common malignancies in childhood and adolescence, with a successful treatment rate of 80 percent (31, 32). The treatment options were continuously improving during the past few decades, but still 10–15% of the patients develop relapse of the disease (33). Emerging new therapy concepts are focused on individualization of the therapy, that can be achieved through precise risk stratification based on the patients' specific genetic aberrations (34), detection of early treatment response and detection of minimal residual disease (MRD) (35, 36).

Genetic alterations that are the most important in ALL risk stratification, and the MRD follow-up, are translocations  $t(9;22)(q34;q11)$  – *BCR/ABL*,  $t(4;11)(q21;q23)$  – *MLL/AF4*,  $t(12;21)(p13;q22)$  – *TEL/AML1* and  $t(1;19)(q23;p13)$  – *E2A/PBX1* (37).

Minimal residual disease (MRD) studies of these translocations allow sensitive detection of leukemic cells undetectable by normal cytomorphologic examination, thereby providing accurate information about the *in vivo* efficacy of cytotoxic treatment (38).

### Pharmacogenetics and Pharmacogenomics

Pharmacogenomics is referred to as the study of variation in the DNA sequence and gene expression as related to drug efficacy and toxicity. It is a base for the implementation of personalized medicine, a young but rapidly advancing field of health care. The goal of pharmacogenomics is to identify genomic and clinical information in order to predict the response to treatment of a person. Pharmacogenomic research is being developed in two main directions: identification of specific genes and gene products correlated with different diseases, which could represent the target for new therapeutics, and identification of genes and gene allelic variants that might influence the response to a drug that has already been used in therapy (39).

Pharmacogenomics completely changes the old-fashioned therapeutic paradigm of »one dose fits all patients« and »trial-and-error« prescription, to a novel, personalized concept of »matching the right therapeutic and the right dose to the specific genetic signature of the patient«.

Due to the rapid development of technology, pharmacogenetics became more and more applied to the whole genome and grew into pharmacogenomics. Pharmacogenomic testing is provided through medical and research institutions that developed it in order to make the treatment of patients more efficient, and also by direct-to-consumer companies, mostly accessible through the Internet (40). Many pharmacogenomic tests are routinely used in clinical practice worldwide. Before administering certain medications to a patient, it is mandatory to perform some pharmacogenomic analyses. For some medications pharmacogenomic testing is just recommended, but for the majority of drugs, the testing used today is only informative (41). Introducing routine pharmacogenomic testing into clinical practice enables patients to get an adequate therapy (correct medications and correct drug dose) in accordance with their genotype (42). This approach reduces duration of treatment, saves the health care system a lot of money for unnecessary medications and provides minimal complications and adverse reactions to the drug. Detection of polymorphisms in certain genes involved in the metabolism of a particular drug defines the metabolic status of a person. This is a criterion for the adequacy of a particular drug and also for a drug dose. However, some other factors, including copy number of the gene, presence or absence of second-

ary or tertiary modifiers, interactions of different drugs and some environmental factors, can also influence the metabolic category of a person. Basic research gives us ever more information that makes the pharmacogenomic testing more accurate. The most clinically relevant pharmacogenomic markers are found in genes for *VKORC* (vitamin K epoxide reductase) (43, 44) and *CYP2C9* (member of the cytochrome P450 family) (45) and they have to be tested before administering anticoagulant therapy (coumarin derivatives and warfarin). Additionally, prior to application of irinotecan therapy, used for colorectal and pulmonary cancer treatment, pharmacogenomic markers in the *UGT1A1* (uridine diphosphate glucuronosyltransferase 1 family, polypeptide A1) gene need to be analyzed (46). Variants in the *TPMT* gene (thiopurine S-methyltransferase) are tested in order to adjust immunosuppressive therapy (6-mercaptopurine, azathioprine and thioguanine), used in the treatment of acute leukemia, inflammatory and autoimmune diseases and in transplantation medicine (47, 48).

Recently, the research in the field of population pharmacogenomics has shown that the study of pharmacogenomic markers in a population and in a certain ethnic community is of great importance (49). The international PGEnI project (Pharmacogenetic for Every Nation Initiative) coordinated by the University of North Carolina, USA, has a goal to help the incorporation of genomic risk data into medication decision-making in every country (50). PGEnI's model is to look at the genetic incidences of causative risk or drug-efficacy markers in a given population and then to try to individualize the health policy, rather than to introduce treatment individualization for each person. PGEnI's bioinformatics tool compares the SNPs (pharmacogenomic markers) found in a particular population with the World Health Organization's »clinical decision trees«, to come up with a prioritized list of medications that should be chosen for the treatment of each disease or trait. Classification of population-specific pharmacogenomic marker frequency profiles could lead to country-specific recommendations for drug efficacy and safety. Serbia is an active member of the PGEnI project. Our preliminary data showed that, due to high frequency of the *UGT1A1* and *CYP2C9* pharmacogenomic markers in the Serbian population, routine testing of these markers for every patient should be performed before administering irinotecan and warfarin drugs.

Easily accessible Internet databases on pharmacogenomics, designed by authoritative agencies, have an important role in building up awareness of the significance of pharmacogenomic testing in both the scientific community and general population. Pharmacogenomics Knowledge Base (PharmGKB) is a freely accessible web database that collects, curates and disseminates knowledge about the impact of human

genetic variation on drug responses (51). American Food and Drug Administration (FDA) (52), European Medicines Agency (EMA) (53) and Pharmaceutical and Medical Devices Agency (PMDA) (54) from Japan are the most relevant world agencies that work on the improvement of public health and safety by reviewing and evaluating clinical information on medications and medical devices, including dosing guidelines and drug labels, potentially clinically actionable gene–drug associations and genotype–phenotype correlations. Pharmacogenomic testing before administering drugs became validated and approved by those agencies, based on clinical studies. The international HapMap Project is focused on the identification and catalogization of genetic similarities and differences in the human population, thus enabling biomedical researchers from all over the world to find the genes involved in diseases and responses to therapeutic drugs (14).

The rapid development and application of »next-generation sequencing« technology have opened the possibility of successful application of pharmacogenomic testing in order to individualize therapy. The ultimate genetic test at a reasonable price, complete human genome sequencing, could change the future of pharmacogenomic testing. Before routine application of this modern technology, it is necessary to intensify basic research and find answers about the influence of genetic variants on phenotype in order to develop appropriate bioinformatics tools. At that point, pharmacogenomic testing will get true clinical significance and personalized medicine will really find the path to each patient.

### **From predictive genetics to preventive medicine**

Predictive genetic testing represents the genetic analysis of a healthy individual in order to predict risk for developing a certain disease before the appearance of early symptoms (presymptomatic risk assessment). The aim of predictive genetics is to define predictive genetic risks factors and determinants of health and disease, based on comprehensive epidemiological studies. Predictive genetic risk markers can be used separately or in combination with other markers in algorithms.

Predictive genetic testing can be very important for people that have any cancer history in the family (5–10% of familial adenomatous polyposis, hereditary nonpolypoid colon cancer, breast cancer and ovarian cancer) (55–57). If family cancer history suggests an increased risk of developing a certain disease, performing genetic testing could be particularly important for the denial of risk. As an illustration, for particular variants of the *BRCA1* and *BRCA2* genes, it has been demonstrated that they are associated with increased risk of breast and ovarian cancers. Variants of *BRCA1* gene account for 5 percent of all breast cancers and

about 50% of all inherited breast cancers. Variants of *BRCA2* gene account for about 30–40% of all inherited breast cancers. Furthermore, these genetic variants contribute to the risk of developing breast or prostate cancer in men. If a woman has a history of breast cancer in her family, preventive genetic testing could reveal her possible carrier status and risk could be assessed. In the case of positive testing results for risk contributing genetic variants, preventive measures could be carried out with the aim to »catch« the disease in the very beginning and to achieve better quality of life (57, 58).

A large number of predictive tests reveal the risk, but do not provide information that the disease will really develop, when it will happen and how severe the symptoms will be. Such tests are used for Crohn's disease (59, 60), cardiovascular disease (61), hypertension (62, 63), rheumatoid arthritis (64), ulcerative colitis (65, 66), venous thromboembolism (67). The positive side of knowing the genetic risks of various diseases and conditions is the awareness of the patient and the physician that preventive measures and diagnostic examinations should be performed on time. The most useful are predictive genetic tests accompanied by efficient diagnostic methods to determine the symptoms and the effectiveness of therapy.

Genetic tests for the diagnosis of hereditary diseases, such as alpha and beta thalassemia (23, 26), cystic fibrosis (25, 68), phenylketonuria (24), Gaucher's disease (69), alpha-1 antitrypsin deficiency (70), hemochromatosis (71), tyrosinemia (72), mucopolysaccharidosis (73) etc. should be performed after the first symptoms of the disease. Genetic tests have a predictive value in the patient's family members. The advantage of using such tests is that, if a person knows that they carry genetic risk at the time of planning the offspring, he/she may turn to genetic counseling for help. Because the sequencing of the human genome in the future will become a financially feasible option, it is possible that the complete genome sequence would be determined for the child at birth, instead of neonatal screening tests. Then, genetic tests that identify genetic disease will become truly predictive genetic tests. This approach would also be of great importance for the diagnosis of rare diseases, which are nowadays characterized by time-consuming diagnostic analyses.

Predictive tests are not only performed when searching for the risk of developing a serious disease. Predictive genetic tests can indicate that a person needs to modify the diet, to avoid the harmful effects of nutrients, for example gluten (for celiac disease) (74), lactose (for adult hypolactasia) (75), caffeine (for hypersensitivity) (76, 77) or fat (for obesity) (78–82). Nutrigenomics, based on the individual's genetic background, provides the ability to correct a congenital metabolic imbalance with proper diet or certain food supplements.

### Molecular genetic markers as therapeutic targets

Knowledge of the molecular structure of disease related genes is also changing the way researchers approach developing new drugs.

The best known molecular-targeted therapy is imatinib mesylate, a tyrosine-kinase inhibitor used in the treatment of Philadelphia chromosome-positive (Ph<sup>+</sup>) chronic myelogenous leukemia (CML) (83). The exact chromosomal defect in Philadelphia chromosome is a reciprocal translocation between chromosomes 9 and 22, designated as t(9;22). As a result of the translocation, the oncogenic *BCR-ABL* gene fusion is formed, producing a constitutive active tyrosine kinase enzyme, which phosphorylates subsequent proteins and initiates the signaling cascade necessary for cancer development. Imatinib mesylate works by preventing *BCR-ABL* enzyme from permanent activation of the »downstream« proteins, thus inhibiting the growth of cancer cells and leading to their death by apoptosis (83). The *BCR-ABL* tyrosine kinase enzyme exists only in cancer cells. Therefore, only cancer cells are killed through the drug's action (84, 85). Imatinib mesylate, an authentic molecular-targeted therapy, was not as efficient as it was expected. Analysis of the *BCR-ABL* enzyme active site of imatinib mesylate resistant CML patients revealed genetic-based changes which prevent binding of the drug. Consequently, new tyrosine-kinase inhibitors were designed to target these molecular defects (86). A brand new approach in the treatment of CML is based on RNA interference. Small interfering RNAs have been designed to inhibit *BCR-ABL* gene expression (87).

During the last decade, research in the field of molecular genetics has made substantial advances in understanding the molecular basis of acute myeloid leukemia (AML). A great number of specific genetic alterations in AML have been identified and characterized. These molecular genetic markers represent a target for the development of new therapeutic agents specifically directed toward leukemic cells (88).

Acute promyelocytic leukemia (APL) is the first AML subtype which is treated with an agent targeted to a molecular genetic aberration. More than 98% of APL cases are characterized by the presence of *PML/RAR $\alpha$*  fusion protein which blocks the differentiation of leukemia cells in the promyelocytic stage. *PML/RAR $\alpha$*  fusion gene was the target for the design of a specific therapeutic agent – all-trans retinoic acid (ATRA).

ATRA leads to a conformational change of the multifunctional complex which includes *PML-RAR $\alpha$* , leading to normal regulation of *RAR $\alpha$* -responsive genes and the induction of the terminal differentiation of APL cells (89). ATRA is commonly used in the treatment of newly diagnosed APL patients. Introduction of ATRA in therapeutic protocols for APL resulted in high clinical remission rates of APL

patients (90–92). However, some patients in time become resistant to ATRA. A new therapeutic agent, arsenic trioxide (ATO), emerged as an option for overcoming ATRA resistance (93, 94). ATO induces differentiation of APL cells, as well as their apoptosis, thus eliminating the effects of *PML-RAR $\alpha$*  genetic defect (95). Therapeutic approach based on ATRA and ATO used for the treatment of APL is the most successful example of differentiation therapy. It represents a prototype for the development of similar therapeutic agents for treatment of other hematological malignancies and cancers.

Another approach of molecular-targeted therapy in AML is based on the principle that the block in the differentiation process of cells can be reversed by abrogation of the epigenetic silencing (96, 97). This is a universal approach used in development of potential drugs for treatment of many diseases, including cancer. The two most common mechanisms of epigenetic silencing, altering the regulation of transcription, have led to the development of clinically applicable drugs. The first mechanism of epigenetic silencing is aberrant DNA methylation. Cytidine analogs such as 5-aza-cytidine or 5-aza-2-deoxycytidine integrate into DNA as alternative nucleotides and trap DNA methyltransferases, causing the formation of demethylated DNA (98). Due to this mechanism, hypermethylation of DNA in malignant cells is reversed (99), generally leading to the induction of differentiation and the inhibition of proliferation of the malignant cells (100). These drugs could replace cytotoxic chemotherapy in the near future (101). The second mechanism of epigenetic silencing, used as a target for molecular therapeutics, is the modification of histones. Deacetylation of histones results in their stronger binding to DNA and eventually to transcriptional repression. Newly developed histone deacetylase (HDAC) inhibitors work as modulators of transcriptional repression of tumor suppressors or factors responsible for normal differentiation and cell growth (102).

Farnesyltransferase inhibitors (FTIs) are small-molecule inhibitors that selectively inhibit farnesylation of a number of intracellular substrate proteins such as RAS. RAS is the most common oncogene in human cancer. Mutations that permanently activate the RAS protein are found in 20–25% of all human tumors and up to 90% in certain types of cancer (e.g. pancreatic cancer) (103). For this reason, RAS inhibitors are studied as a potential therapy for the treatment of malignancies and other diseases with RAS overexpression.

Another molecular target, successfully used in the design of molecular therapy, is apoptosis. Overexpression of the Bcl-2, an antiapoptotic protein, was observed in hematological malignancies. Cells that have less Bcl-2 are not only more susceptible to apoptosis, but also more sensitive to chemotherapy (104).

Antisense oligonucleotides block target mRNA specifically. An antisense oligonucleotide-based therapy inhibits Bcl-2 overexpression, promotes apoptosis and diminishes drug resistance in patients with AML (105, 106).

A number of therapies based on targeting gene variants responsible for malignant transformation have been used: genetic variants of *EGFR* gene in lung cancer and glioblastoma are treated with cetuximab, gefitinib, etc., *KIT* and *PDGFR* gene variants in sarcoma, glioma, melanoma, liver and renal cancer are treated with imatinib, nilotinib etc., *BRAF* gene variants in melanoma are treated with RAF inhibitors, *BRCA* gene variants in breast, ovarian, prostate and pancreatic cancers are treated with PARP inhibitors, *HER2*-positive breast cancer is treated with Herceptin (107).

Up until recently, revolutionary discoveries in the field of molecular genetics had to wait for years to be applied in medical practice. Today, each novel molecular mechanism is applied immediately after its identification. Therefore, new emerging molecular-targeted therapy is constantly being introduced into clinical practice.

### Gene therapy

Disease related genes and disease causing genetic variants can be treated by introducing genetic material into a cell to fight or prevent disease. The idea of gene therapy, the way to repair defective genes, was born thirty years ago and is still considered controversial in some scientific communities (108, 109). However, research on gene therapy has been conducted for a number of diseases, especially monogenic diseases (thalassemia, cystic fibrosis, hemophilia) and cancer, through various approaches (110).

Genetic material can be delivered to a cell using a »vector«. The most commonly used vectors in gene therapy are viruses, since they are natural deliverers of genetic material (their own) into a human cell. Viral genome is altered in a manner to make a virus safe and non-infective, and to carry a therapeutic gene (111, 112). A therapeutic gene is not only a »healthy« copy of the gene which replaces a mutated gene, but also a genetic material which inactivates a mutated gene that functions improperly or any other genetic material that can fight a disease, when introduced in a cell or incorporated in a human genome. Virtually all cells and tissues are potential targets for gene therapy. However, all gene therapy protocols in humans are directed to somatic cells which are non-reproductive. Somatic cell therapy affects only the targeted cells in the patient, and is not passed on to future generations. Germline gene therapy remains controversial and prohibited in most of the countries.

Somatic gene therapy is divided in three categories: *ex vivo*, *in vivo* and *in situ*. In *ex vivo* gene ther-

apy, patient's cells are removed from the body and then grown and genetically modified outside the body. After insertion of the therapeutic gene into the patient's cells, they are returned to the patient. Interior, *in vivo*, gene therapy means that genetic manipulation and the transfer of the therapeutic gene to cell is performed inside the patient's body, while *in situ* gene therapy means that the therapeutic gene is delivered directly to the tissue that has to be treated in order to restore the missing function (113).

In the early 1990s, gene therapy was successful in combating SCID (Severe Combined Immunodeficiency, also called ADA deficiency or »bubble baby disease«) for the first time. *Ex vivo* approach was applied. Retroviral vectors were used to introduce the normal allele of the adenosine deaminase (*ADA*) gene into the cells of a 4-year-old girl, born with ADA deficiency. In this disease, an abnormal variant of the *ADA* gene fails to make ADA, a protein indispensable for the correct function of T-lymphocytes. The girl, and many more after her suffering from SCID, was cured and had a normal life, although she had to repeat the gene therapy protocol every few months. From 1993, SCID immunodeficiency was considered 100% cured by gene therapy (113, 114). However, in 2002, two cases of T-cell ALL were newly diagnosed after retrovirus-mediated gene therapy of SCID immunodeficiency. It was confirmed that the therapeutic gene was integrated in the regulatory region of *LMO2* oncogene, most probably causing the malignant phenotype (115, 116). Moreover, an 18-year-old high-school graduate died after adenovirus-mediated gene therapy of ornithine transcarbamylase. Both incidents were the result of well-known weaknesses of the gene therapy of today, the use of viral vectors for the delivery of the therapeutic gene in the patient's cell (the position of viral integration in the human genome is hard to control and production of noninfective viral particles is not yet efficient enough) (117, 118).

Gene therapy had its best and worst times. However, researchers continue to improve gene therapy and develop new approaches. Today, there are more than a thousand on-going clinical trials for gene therapy. Finally, in November 2012, the first gene therapy received marketing authorization from the European Commission, for patients with lipoprotein lipase (LPL) deficiency (119). The gene therapy product, alipogene tiparvovec, is based on an adeno-associated virus vector and the replacement of the gene responsible for LPL expression, which is defective in patients with LPL deficiency (120). These patients have an extremely high level of serum triglycerides causing recurrent and life threatening pancreatitis (121). Definitely, the first commercially-approved gene therapy product in the West represents an outstanding medical achievement.

»Gene therapy, like every other major new technology, takes time to develop. It will succeed with

time. And it is important that it does succeed, because no other area of medicine holds as much promise for providing cures for the many devastating diseases that now ravage humankind« (122).

### Conclusion

The future of medicine, without a doubt, lies in the realization of the idea of personalized medicine. The final achievement of the Human Genome Project was the creation of a catalogue of human genes. Molecular biologists of today are facing an ambitious goal of understanding the function of all genes, associating DNA content with individual phenotype as well as medically relevant features. Gene expression profil-

ing will be able to reveal all genes relevant for certain pathologies, guiding medical doctors toward specific molecular therapy. As soon as gene manipulation begins to cure, a large number of people will have a long and better life, despite the predispositions. In this way, an old proverb will finally become true: »Fato prudentia maior est« (Wisdom is stronger than destiny).

*Acknowledgements.* This work was supported by the Ministry of Education, Science and Technological Development, Republic of Serbia (Grant No. III41004).

### Conflict of interest statement

The authors stated that there are no conflicts of interest regarding the publication of this article.

### References

1. Offit K. Personalized medicine: new genomics, old lessons. *Hum Genet* 2011; 130: 3–14.
2. National Cancer Institute, USNIH (2011). <http://www.cancer.gov/dictionary/?CdrID=561717>. (Accessed 27 Jan 2011)
3. H.R. 5440 (2010) Genomics and personalized medicine act of 2010. <http://www.opencongress.org/bill/111-h5440/text>. Accessed 27 Jan 2011
4. Steele FR. Personalized medicine: something old, something new. *Pers Med* 2009; 6: 1–5.
5. Gužvić M. The history of DNA sequencing. *J Med Biochem* 2013; 32: 301–12.
6. Pavlović S. TPMT gene polymorphisms: on the doorstep of personalized medicine. *Indian J Med Res* 2009; 129(5): 478–80.
7. Pavlović S, Zukić B, Stojiljković M. Personalizovana medicina. In: Antonić S and Popović A, editors. *Matične ćelije i genetika u službi čovečanstva*. Beograd: Univerzitetna biblioteka Svetozar Marković, 2013: 8–26.
8. Lueking A, Possling A, Huber O, Beveridge A, Horn M, Eickhoff H, Schuchardt J, et al. A nonredundant human protein chip for antibody screening and serum profiling. *Mol Cell Proteomics* 2003; 2: 1342–9.
9. Soon WW, Hariharan M, Snyder MP. High-throughput sequencing for biology and medicine. *Mol Syst Biol* 2013; 9: 640.
10. Gorreta F, Carbone W, Barzaghi D. Genomic profiling: cDNA arrays and oligoarrays. *Methods Mol Biol* 2012; 823: 89–105.
11. Bush WS, Moore JH. Chapter 11: Genome-wide association studies. *PLoS Comput Biol* 2012; 8(12): e1002822.
12. Drmanac R. Medicine. The ultimate genetic test. *Science* 2012; 336(6085): 1110–12.
13. Human Genome Project ([http://www.ornl.gov/sci/techresources/Human\\_Genome/home.shtml](http://www.ornl.gov/sci/techresources/Human_Genome/home.shtml))
14. HapMap Project (<http://hapmap.ncbi.nlm.nih.gov/>)
15. Wan TS, Ma ES. Molecular cytogenetics: an indispensable tool for cancer diagnosis. *Chang Gung Med J* 2012; 35(2): 96–110.
16. Sanger F, Nicklen S, Coulson AR. DNA sequencing with chain-terminating inhibitors. *Proc Natl Acad Sci USA* 1977; 74(12): 5463–7.
17. Liu L, Li Y, Li S, Hu N, He Y, Pong R, et al. Comparison of next-generation sequencing systems. *J Biomed Biotechnol* 2012; 2012: 251364.
18. Henson J, Tischler G, Ning Z. Next-generation sequencing and large genome assemblies. *Pharmacogenomics* 2012; 13(8): 901–15.
19. Harrison RJ. Understanding genetic variation and function – the applications of next generation sequencing. *Semin Cell Dev Biol* 2012; 23(2): 230–6.
20. Wheeler DA, Srinivasan M, Egholm M, Shen Y, Chen L, McGuire A, et al. The complete genome of an individual by massively parallel DNA sequencing. *Nature* 2008; 452(7189): 872–6.
21. Peters BA, Kermani BG, Sparks AB, Alferov O, Hong P, Alexeev A, et al. Accurate whole-genome sequencing and haplotyping from 10 to 20 human cells. *Nature* 2012; 487(7406): 190–5.
22. National Institute of Health (NIH), A Catalog of Published Genome-Wide Association Studies (<http://www.genome.gov/gwastudies/>)
23. Antonarakis SE, Kazazian HH Jr, Orkin SH. DNA polymorphism and molecular pathology of the human globin gene clusters. *Hum Genet* 1985; 69: 1–14.
24. J. Zschocke. Phenylketonuria mutations in Europe. *Hum Mutat* 2003; 21: 345–56.
25. Cystic Fibrosis Mutation Data Base (<http://www.genet.sickkids.on.ca/cftr/>)
26. Pavlović S, Urošević J, Poznančić J, Perišić Lj, Petručev B, Tošić N, et al. Molecular basis of thalassemia syndromes in Serbia and Montenegro. *Acta Haematol* 2005; 113: 175–80.

27. Radmilović M, Zukić B, Stanković B, Karan-Đurašević T, Stojiljković M, Spasovski, et al. Thalassemia Syndromes in Serbia: An update. *Hemoglobin* 2010; 34(5): 477–85.
28. Stojiljković M, Jovanović J, Đorđević M, Grković S, Cvorkov Dražić M, Petručev B, et al. Molecular and phenotypic characteristics of phenylketonuria patients in Serbia and Montenegro. *Clin Genet* 2006; 70: 151–5.
29. Dabović B, Radojković D, Minić P, Savić J, Savić A. Frequency of the delta F508 deletion and G551D, R553X and G542X mutations in Yugoslav CF patients. *Hum Genet* 1992; 88(6): 699–700.
30. Lazić J, Tošić N, Dokmanović L, Krstovski N, Rodić P, Pavlović S, et al. Clinical features of the most common fusion genes in childhood acute lymphoblastic leukemia. *Med Oncol* 2010; 27(2): 449–53.
31. Pui CH, Relling MV, Downing R. Acute lymphoblastic leukemia. *N Engl J Med* 2004; 350: 1535–48.
32. Silverman LB, Gelber RD, Dalton VK, Asselin BL, Barr RD, Clavell LA, et al. Improved outcome for children with acute lymphoblastic leukemia: results of Dana-Faber Consortium Protocol 91–01. *Blood* 2001; 97: 1211–18.
33. Gaynon PS. Childhood ALL and relapse. *Br J Haematol* 2005; 131: 579–87.
34. Carroll WL, Bhojwani D, Min DJ, Moskowitz N, Raetz EA. Childhood Acute Lymphoblastic leukemia in the age of genomics. *Pediatr Blood Cancer* 2006; 46: 570–8.
35. Panzer-Grumayer ER, Schneider M, Pancer S, Fasching K, Gadner H. Rapid molecular response during early induction therapy predicts a good outcome in childhood acute lymphoblastic leukemia. *Blood* 2000; 95: 790–4.
36. Biondi A, Valesecchi MG, Seriu T, D'Aniello E, Willemse MJ, Fasching K. Molecular detection of minimal residual disease is a strong predictive factor of relapse in childhood B-lineage acute lymphoblastic leukemia with medium risk features. A case control study of the International BFM study group. *Leukemia* 2000; 14: 1939–43.
37. Mitelman F, Johansson B, Mertens F. The impact of translocations and gene fusions on cancer causation. *Nat Rev Cancer* 2007; 7(4): 233–5.
38. Szczepanski T, Orfao A, van der Velden VHJ, San Miguel JF, van Dongen JJM. Minimal residual disease in leukemia patients. *Lancet Oncol* 2001; 2: 409–17.
39. Wolf CR, Smith G, Smith RL. Science, medicine, and the future: Pharmacogenetics. *BMJ* 2000; 320(7240): 987–90.
40. Chua EW, Kennedy MA. Current State and Future Prospects of Direct-to-Consumer Pharmacogenetics. *Front Pharmacol* 2012; 3: 152.
41. Tauser RG. Matching the right foundation at personalized medicine in the right genomic era. In: Sanoudou D, editor. *Clinical Applications of Pharmacogenetics*, InTech, Rijeka, Croatia, 2012: 3–34.
42. Pavlović S, Zukić B, Nikčević G. Pharmacogenomics of Thiopurine S-Methyltransferase: Clinical Applicability of Genetic Variants. In: Sanoudou D, editor. *Clinical Applications of Pharmacogenetics*, InTech, Rijeka, Croatia, 2012: 75–94.
43. D'Andrea G, D'Ambrosio RL, Di Perna P, Chetta M, Santacroce R, Brancaccio V, et al. A polymorphism in the VKORC1 gene is associated with an interindividual variability in the dose-anticoagulant effect of warfarin. *Blood* 2005; 105: 645–9.
44. Kovač M, Rakičević L, Kušić-Tišma J, Radojković D. Pharmacogenetic tests could be helpful in predicting of VKA maintenance dose in elderly patients at treatment initiation. *J Thromb Thrombolysis* 2013; 35(1): 90–4.
45. Aithal GP, Day CP, Kesteven PJ, Daly AK. Association of polymorphisms in the cytochrome P450 CYP2C9 with warfarin dose requirement and risk of bleeding complications. *Lancet* 1999; 353: 717–19.
46. Hoskins JM, Goldberg RM, Qu P, Ibrahim JG, McLeod HL. UGT1A1\*28 genotype and irinotecan-induced neutropenia: dose matters. *J Natl Cancer Inst* 2007; 99: 1290–5.
47. Dokmanović L, Urošević J, Janić D, Jovanović N, Petručev B, Tošić N, et al. Analysis of thiopurine S-methyltransferase Polymorphism in the Population of Serbia and Montenegro and Mercaptopurine Therapy Tolerance in Childhood Acute Lymphoblastic Leukemia. *Ther Drug Monit* 2006; 28: 800–6.
48. Dokmanović L, Janić D, Krstovski N, Zukić B, Tošić N, Pavlović S. Importance of genotyping of thiopurine S-methyltransferase in children with acute lymphoblastic leukaemia during maintenance therapy. *Srp Arh Celok Lek* 2008; 136(11–12): 609–16.
49. Mette L, Mitropoulos K, Vozikis A, Patrinos GP. Pharmacogenomics and public health: implementing 'populationalized' medicine. *Pharmacogenomics* 2012; 13(7): 803–13.
50. Pharmacogenetic for Every Nation Initiative Project (<http://www.pgeni.org/>)
51. Pharmacogenetics and Pharmacogenomics Knowledge Base (PharmGKB) (<http://www.pharmgkb.org/>)
52. American Food and Drug Administration (<http://www.fda.gov/>)
53. European Medicines Agency (<http://www.ema.europa.eu/ema/>)
54. Pharmaceutical and Medical Devices Agency (<http://www.pmda.go.jp/english/>)
55. Lin OS. Colorectal cancer screening in patients at moderately increased risk due to family history. *World J Gastrointest Oncol* 2012; 4(6): 125–30.
56. Jasperson KW, Tuohy TM, Neklason DW, Burt RW. Hereditary and familial colon cancer. *Gastroenterology* 2010; 138(6): 2044–58.
57. Christinat A, Pagani O. Practical aspects of genetic counseling in breast cancer: Lights and shadows. *Breast* 2013; doi: S0960-9776(13)00087-8. 10.1016/j.breast.2013.04.006.
58. Dobričić J, Krivokuća A, Brotto K, Mališić E, Radulović S, Branković-Magić M. Serbian high-risk families: extensive results on BRCA mutation spectra and frequency. *J Hum Genet* 2013; doi: 10.1038/jhg.2013.30.

59. Protić M, Pavlović S, Bojić D, Krstić M, Radojčić Z, Tarabar D, et al. CARD15 gene polymorphism in Serbian patients with Crohn's disease: genotype-phenotype analysis. *Eur J Gastroenterol Hepatol* 2008; 20(10): 978–84.
60. Anderson CA, Massey DC, Barrett JC, Prescott NJ, Tremelling M, Fisher SA, et al. Investigation of Crohn's disease risk loci in ulcerative colitis further defines their molecular relationship. *Gastroenterology* 2009; 136(2): 523–9.
61. O'Donnell CJ, Nabel EG. Genomics of cardiovascular disease. *N Engl J Med* 2011; 365: 2098–109.
62. Baudin B. Polymorphism in angiotensin II receptor genes and hypertension. *Exp Physiol* 2005; 90(3): 277–82.
63. Tabara Y, Kohara K, Miki T. Millennium Genome Project for Hypertension. Hunting for genes for hypertension: the Millennium Genome Project for Hypertension. *Hypertens Res* 2012; 35(6): 567–73.
64. McClure A, Lunt M, Eyre S, Ke X, Thomson W, Hinks A, Bowes J, et al. Investigating the viability of genetic screening/testing for RA susceptibility using combinations of five confirmed risk loci. *Rheumatology (Oxford)* 2009; 48(11):1369–74. Erratum in: *Rheumatology (Oxford)* 2011; 50(6): 1178.
65. Cummings SA, Rubin DT. The complexity and challenges of genetic counseling and testing for inflammatory bowel disease. *J Genet Couns* 2006; 15(6): 465–76.
66. Vermeire S. Review article: genetic susceptibility and application of genetic testing in clinical management of inflammatory bowel disease. *Aliment Pharmacol Ther* 2006; 24 Suppl 3: 2–10.
67. Đorđević V, Rakičević L, Radojković D. An overview of genetic risk factors in thrombophilia. *Srp Arh Celok Lek* 2010; 138: Suppl 1: 79–8.
68. Castellani C, Cuppens H, Macek M Jr, Cassiman JJ, Kerem E, et al. Consensus on the use and interpretation of cystic fibrosis mutation analysis in clinical practice. *J Cyst Fibros* 2008; 7(3): 179–96.
69. Rodić P, Pavlović S, Kostić T, Suvajdžić Vuković N, Đorđević M, Šumarac Z, et al. Gammopathy and B lymphocyte clonality in patients with Gaucher type I disease. *Blood Cells Mol Dis* 2013; 50(3): 222–5.
70. Topić A, Stanković M, Divac-Rankov A, Petrović-Stanojević N, Mitić-Milikić M, Nagorni-Obradović L, et al. Alpha-1-antitrypsin deficiency in Serbian adults with lung diseases. *Genet Test Mol Biomarkers* 2012; 16(11): 1282–6.
71. Sarić M, Zamurović Lj, Keckarević-Marković M, Keckarević D, Stevanović M, Savić-Pavićević D, et al. Frequency of the hemochromatosis gene mutations in the population of Serbia and Montenegro. *Clin Genet* 2006; 70(2): 170–2.
72. Sniderman King L, Trahms C, Scott CR. Tyrosinemia Type 1. In: Pagon RA, Bird TD, Dolan CR, Stephens K, Adam MP, editors. *SourceGeneReviews™* [Internet]. Seattle (WA): University of Washington, Seattle; 1993–2006 Jul 24 [updated 2011 Aug 25].
73. Clarke LA, Heppner J. Mucopolysaccharidosis Type I. In: Pagon RA, Bird TD, Dolan CR, Stephens K, Adam MP, editors. *GeneReviews™* [Internet]. Seattle (WA): University of Washington, Seattle; 1993–2002 Oct 31 [updated 2011 Jul 21].
74. Lavant EH, Agardh DJ, Nilsson A, Carlson JA. A new PCR-SSP method for HLA DR-DQ risk assessment for celiac disease. *Clin Chim Acta* 2011; 412(9–10): 782–4.
75. Usai-Satta P, Scarpa M, Oppia F, Cabras F. Lactose malabsorption and intolerance: What should be the best clinical management? *World J Gastrointest Pharmacol Ther* 2012; 3(3): 29–33.
76. Cornelis MC, El-Sohemy A, Campos H. Genetic polymorphism of CYP1A2 increases the risk of myocardial infarction. *J Med Genet* 2004; 41(10): 758–62.
77. Cornelis MC, El-Sohemy A, Kabagambe EK, Campos H. Coffee, CYP1A2 genotype, and risk of myocardial infarction. *JAMA* 2006; 295(10): 1135–41.
78. Kilpeläinen TO, Qi L, Brage S, Sharp SJ, Sonestedt E, Demerath E, et al. Physical activity attenuates the influence of FTO variants on obesity risk: a meta-analysis of 218,166 adults and 19,268 children. *PLoS Med* 2011; 8(11): e1001116.
79. Levy E, Ménard D, Delvin E, Stan S, Mitchell G, Lambert M, et al. The polymorphism at codon 54 of the FABP2 gene increases fat absorption in human intestinal explants. *J Biol Chem* 2001; 276(43): 39679–84.
80. Ukkola O, Tremblay A, Bouchard C. Beta-2 adrenergic receptor variants are associated with subcutaneous fat accumulation in response to long-term overfeeding. *Int J Obes Relat Metab Disord* 2001; 25(11): 1604–8.
81. Memisoglu A, Hu FB, Hankinson SE, Manson JE, De Vivo I, Willett WC, et al. Interaction between a peroxisome proliferator-activated receptor gamma gene polymorphism and dietary fat intake in relation to body mass. *Hum Mol Genet* 2003; 12(22): 2923–9.
82. Marti A, Corbalán MS, Martínez-Gonzalez MA, Martínez JA. TRP64ARG polymorphism of the beta 3-adrenergic receptor gene and obesity risk: effect modification by a sedentary lifestyle. *Diabetes Obes Metab* 2002; 4(6): 428–30.
83. Goldman JM, Melo JV. Chronic myeloid leukemia—advances in biology and new approaches to treatment. *N Engl J Med* 2003; 349(15): 1451–64.
84. Fausel C. Targeted chronic myeloid leukemia therapy: Seeking a cure. *Am J Health Syst Pharm* 2007; 64: 9–15.
85. Stegmeier F, Warmuth M, Sellers WR, Dorsch M. Targeted Cancer Therapies in the Twenty-First Century: Lessons From Imatinib. *Clin Pharmacol Ther* 2010; 87(5): 543–52.
86. Talpaz M, Shah NP, Kantarjian H, Donato N, Nicoll J, Paquette R, et al. Dasatinib in imatinib-resistant Philadelphia chromosome-positive leukemias. *N Engl J Med* 2006; 354(24): 2531–41.
87. Koldehoff M, Kordelas L, Beelen DW, Elmaagacli AH. Small interfering RNA against BCR-ABL transcripts sen-

- sitize mutated T315I cells to nilotinib. *Haematol* 2010; 95(3): 388–97.
88. Pavlović S, Tošić N. Molecular genetics of acute myeloid leukemia. *Global J Biochem* 2012; 3(8): 1–16.
89. Grignani F, De Matteis S, Nervi C, Tomassoni L, Gelmetti V, Cioce M, et al. Fusion proteins of the retinoic acid receptor-alpha recruit histone deacetylase in promyelocytic leukaemia. *Nature* 1998; 391: 815–18.
90. Tallman MS, Andersen JW, Schiffer CA, Appelbaum FR, Feusner JH, Ogden A, et al. All-trans retinoic acid in acute promyelocytic leukemia. *N Engl J Med* 1997; 337: 1021–8.
91. Fenaux P, Chastang C, Chevret S, Sanz M, Dombret H, Archimbaud E, et al. A randomized comparison of all-trans retinoic acid (ATRA) followed by chemotherapy and ATRA plus chemotherapy and the role of maintenance therapy in newly diagnosed acute promyelocytic leukemia. *Blood* 1999; 94: 1192–200.
92. Tallman MS, Andersen JW, Schiffer CA, Appelbaum FR, Feusner JH, Woods WG, et al. All-trans retinoic acid in acute promyelocytic leukemia: long-term outcome and prognostic factor analysis from the North American Intergroup Protocol. *Blood* 2002; 100: 4298–302.
93. Soignet SL, Frankel SR, Douer D, Tallman MS, Kantarjian H, Calleja E, et al. United States multicenter study of arsenic trioxide in relapsed acute promyelocytic leukemia. *J Clin Oncol* 2001; 19: 3852–60.
94. Estey E, Garcia-Manero G, Ferrajoli A, Faderl S, Verstovsek S, Jones D, et al. Use of all-trans retinoic acid plus arsenic trioxide as an alternative to chemotherapy in untreated acute promyelocytic leukemia. *Blood* 2006; 107: 3469–73.
95. Chen GQ, Shi XG, Tang W, Xiong SM, Zhu J, Cai X, et al. Use of arsenic trioxide (As<sub>2</sub>O<sub>3</sub>) in the treatment of acute promyelocytic leukemia (APL): As<sub>2</sub>O<sub>3</sub> exerts dose dependent dual effects on APL cells. *Blood* 1997; 89: 3345–53.
96. Chim CS, Wong AS, Kwong YL. Infrequent hypermethylation of CEBPA promotor in acute myeloid leukaemia. *Br J Haematol* 2002; 119: 988–90.
97. Figueroa ME, Reimers M, Thompson RF, Ye K, Li Y, Selzer RR, et al. An integrative genomic and epigenomic approach for the study of transcriptional regulation. *PLoS ONE* 2008; 3: e1882.
98. Egger G, Liang G, Aparicio A, Jones PA. Epigenetics in human disease and prospects for epigenetic therapy. *Nature* 2004; 429: 457–63.
99. Daskalakis M, Nguyen TT, Nguyen C, Guldborg P, Köhler G, Wijermans P, et al. Demethylation of a hypermethylated P15/INK4B gene in patients with myelodysplastic syndrome by 5-Aza-2-deoxycytidine (decitabine) treatment. *Blood* 2002; 100: 2957–64.
100. Issa JP, Garcia-Manero G, Giles FJ, Mannari R, Thomas D, Faderl S, et al. Phase 1 study of low-dose prolonged exposure schedules of the hypomethylating agent 5-aza-2-deoxycytidine (decitabine) in hematopoietic malignancies. *Blood* 2004; 103: 1635–40.
101. Garcia-Manero G. Demethylating agents in myeloid malignancies. *Curr Opin Oncol* 2008; 20: 705–10.
102. Bolden JE, Peart MJ, Johnstone RW. Anticancer activities of histone deacetylase inhibitors. *Nat Rev Drug Discov* 2006; 5: 769–84.
103. Downward J. Targeting RAS signaling pathways in cancer therapy. *Nat Rev Cancer* 2003; 3(1): 11–22.
104. Miyashita T, Reed JC. Bcl2 oncoprotein blocks chemotherapy-induced apoptosis in human leukemia cell line. *Blood* 1993; 81: 151–7.
105. Marcucci G, Byrd JC, Dai G, Klisović MI, Kourlas PJ, Young DC, et al. Phase 1 and pharmacokinetic studies of G3139, a Bcl-2 antisense oligonucleotide, in combination with chemotherapy in refractory or relapsed acute leukemia. *Blood* 2003; 101: 425–32.
106. Marcucci G, Stock W, Dai G, Klisovic RB, Liu S, Klisovic MI, et al. Phase II study of oblimersen sodium an antisense to Bcl2 in untreated older patients with AML. *J Clin Oncol* 2005; 23: 3404–11.
107. MacConaill LE, Garraway LA. Clinical Implications of the Cancer Genome. *J Clin Oncol* 2010; 28: 5219–28.
108. Anderson WF. Prospects for human gene therapy. *Science* 1984; 226: 401–9.
109. Friedmann T. Progress toward human gene therapy. *Science* 1989; 244: 1275–81.
110. Yamamoto M, Curiel DT. Cancer gene therapy. *Technol Cancer Res Treat* 2005; 4(4): 315–30.
111. Goff S, Berg P. Construction of hybrid viruses containing SV40 and lambda; phage DNA segments and their propagation in cultured monkey cells. *Cell* 1976; 9(4): 695–705.
112. Eglitis MA, Anderson WF. Retroviral vectors for introduction of genes into mammalian cells. *Biotechniques* 1988; 6(7): 608–14.
113. Anderson WF. Human gene therapy. *Science* 1992; 256(5058): 808–13.
114. Cavazzana-Calvo M, Lagresle C, Hacein-Bey-Abina S, Fischer A. Gene therapy for severe combined immunodeficiency. *Annu Rev Med* 2005; 56: 585–602.
115. Babić N. Clinical pharmacogenomics and concept of personalized medicine. *J Med Biochem* 2012; 31: 281–6.
116. Johnston J, Baylis F. What ever happened to gene therapy? A review of recent events. *Clin Res* 2004; 4: 11–15.
117. Hacein-Bey-Abina S, Von Kalle C, Schmidt M, McCormack MP, Wulffraat N, Leboulch P, et al. LMO2-Associated Clonal T Cell Proliferation in Two Patients after Gene Therapy for SCID-X1. *Science* 2003; 302(5644): 415–19.
118. Lehrman S. Virus treatment questioned after gene therapy death. *Nature* 1999; 401: 517–18.
119. Kastelein JJP, Colin JD, Ross CJD, Hayden M. From Mutation Identification to Therapy: Discovery and Origins of the First Approved Gene Therapy in the Western World. *Hum Gene Ther* 2013; 24: 472–8.

120. Carpentier AC, Frisch F, Labbé SM, Gagnon R, de Wal J, Greentree S, et al. Effect of alipogene tiparvovec (AAV1-LPL(S447X)) on postprandial chylomicron metabolism in lipoprotein lipase-deficient patients. *J Clin Endocrinol Metab* 2012; 97: 1635–44.
121. Brunzell J, Deeb S. Familial lipoprotein lipase deficiency, ApoCII deficiency, and hepatic lipase deficiency. In: CR Scriver, AI Beaudet, WS Sly, D Vale, editors. *The Metabolic and Molecular Bases of Inherited Disease*, 8<sup>th</sup> ed. New York, NY: McGraw-Hill Inc, 2000: 2789–816.
122. Anderson WF. The best of times, the worst of times. *Science* 2000; 288: 627–9.

*Received: May 31, 2013*

*Accepted: July 9, 2013*