## UDK 577.1:61

J Med Biochem 31: 287-294, 2012

**ISSN 1452-8258** 

Review article Pregledni članak

# PATHOPHYSIOLOGICAL IMPORTANCE OF NITRIC OXIDE IN CORONARY HEART DISEASE

PATOFIZIOLOŠKI ZNAČAJ AZOT-MONOKSIDA U KORONARNOJ BOLESTI SRCA

Vidosava B. Đorđević<sup>1,2</sup>, Ivana Stojanović<sup>1</sup>, Slavica Kundalić<sup>2</sup>, Tatjana Ristić<sup>2</sup>, Radmila Pavlović<sup>1</sup>, Vladan Ćosić<sup>2</sup>, Tatjana Cvetković<sup>1</sup>

> <sup>1</sup>Institute of Biochemistry, Faculty of Medicine, Niš, Serbia <sup>2</sup>Centre for Medical Biochemistry, Clinical Centre Niš, Serbia

Summary: Nitric oxide (NO) is produced by many cells in the body; however, its production by vascular endothelium is particularly important in the r equilation of blood flow Vascular actions of NO include the following: dir ect vasodilation, indirect vasodilation by inhibiting the vasoconstrictor influences, anti-thrombotic, anti-inflammatory and anti-proliferative effects. Due to its importance in vascular function, abnormal production of NO, occurring in different diseases can adversely affect blood flow and other vascular functions. It has been suggested that alterations in NO generation ar e a critical cause of injury in the ischemic heart. A biologic link between the endothelial damage and ather osclerotic coronary arterial disease has been pr esumably related to de creased arterial bioavailability of NO thr ough the increased leucocyte and platelet adhesions, vasoconstriction and smooth muscle cell proliferation. However, the precise mechanism of the impair ed NO generation is not known, and there is a considerable controversy regarding whether myocardial ischemia results in increased or decreased NO formation. Asymmetric dimethylarginine (ADMA) is a natural, competitive inhibitor, and one of the primary factors controlling the nitric oxide production. ADMA was found to be elevated and closely cor related with the impair ed vasodilator function in conditions associated with the endothelial dysfunction, such as hypercholesterolemia, hypertension, insulin resistance and type 2 diabetes, and renal failure. But ADMA also seems to be involved in myocar dial ischemia, since its plasma levels predict future coronary events in patients with the elevated cardiovascular risk. It has been r ecently reported that the elevated plasma ADMA concentrations in the acute coronary events ar e an independent car diovascular risk factor.

**Keywords:** nitric oxide, endothelial dysfunction, cor onary heart disease, risk factors

Kratak sadr`aj: Azot-monoksid (NO) se sintetiše u mno gim ćelijama organizma ali je njegova pr odukcija u vaskularnom endotelu nar očito važna u r egulaciji protoka krvi. Vaskularni efekti uključuju: dir ektnu vazodilataciju, indir ektnu vazodilataciju inhibicijom vazokonstriktora, anti-tr ombotični efekat, anti-inflamatorni i anti-proliferativni efekat. Zbog njegove važnosti u vaskularnoj funkciji, abnormalna produkcija NO, koja se javlja u različitim bolestima, može imati nepovoljan efekat na protok krvi i druge vaskularne funkcije. Smatra se da je por emećaj sinteze NO kritičan uzr ok oštećenja u ishemičnom srcu. Biološka veza između endotelnog oštećenia i aterosklerotske bolesti koronarnih arterija je pre svega vezivana za smanjenje raspoloživog NO kroz povećane adhezije leukocita i trombocita, vazokonstrikcije i proliferacije glatkomišićnih ćelija. Međutim, precizni mehanizam poremećaja sinteze NO još uvek nije jasan i postoje kontr overze o tome da li miokar dna ishemija dovodi do povećanog ili smanjenog stvaranja NO. Asimetrični dimetilarginin (ADMA) je prirodni, kompetitivni inhibitor i jedan od primar nih faktora koji kontrolišu produkciju NO. Koncentracija asimetričnog dimetilarginina je povećana i usko korelira sa poremećenom vazodilatacijom u uslovima kada postoji endotelna disfunkcija kao što su hiperholesterolemija, hipertenzija, rezistenca na insulin i dijabet Tip 2 i renalna insuficijencija. Takođe, ADMA je uključen u miokardnu ishemiju i na osnovu njegove vr ednosti u plazmi mogu se predvideti budući koronarni događaji kod pacijenata sa povećanim kardiovaskularnim rizikom. Nedavno je saopšteno da je povećana koncentracija ADMA u akutnim koronarnim događajima nezavisan kardiovaskularni rizik faktor.

**Klju~nere~i:** azot-monoksid, endotelna disfunkcija, ishe - mijska bolest srca, faktori rizika

Address for correspondence: Vidosava B. Đorđević Institute of Biochemistry, Faculty of Medicine, Niš, Serbia

# Introduction

Nitric oxide (NO) is a simple, diatomic gaseous molecule that is a key signaling messenger in the cardiovascular system (1). NO ser ves many important biological functions in the car diovascular physiology where it is produced in both endothelial and smooth muscle cells. NO pr oduced in endothelium contr ols vascular tonus and per meability, maintains vascular integrity by inhibiting the platelet aggregation, leukocyte endothelium adhesion and vascular smooth muscle proliferation. NO pr oduced in car diac smooth muscle cells r equlates cardiac contractility (2). In order for NO to preserve normal vascular physiology, its adequate levels have to be pr oduced. NO is synthesized from amino acid L -arginine and molecular oxygen by one of three nitric oxide synthases (NOS): neuronal NOS (nNOS, NOS1), endothelial NOS (eNOS, NOS3), and inducible NOS (iNOS, NOS2). These isoforms are encoded by the genes located on different chromosomes (12g24.2; 7g35-36; 17cenq42), and show 50%-60% of homology in their ami no acid sequences in the oxidase and r eductase domains that bind cofactors FAD, FMN, NADPH and BH4 (3), NOS isofor mes exert different characteristics wich r eflect their various functions in vivo (4). Endothelial NOS and nNOS ar e expressed constitutively and their activity is primarily r equilated by the levels of intracellular calcium and calmodulin concentrations, while iNOS is expressed during pathological processes, such as heart failure (5), after induction by cytokines and other inflammatory mediators, and produces high levels of nitric oxide (6).

nNOS is predominantly expressed in some neurons and skeletal muscles, eNOS in endothelial cells, and iNOS in macrophages and monocytes. In spite of their names, different types of cells express these isoforms, and numerous tissues express more than one isoform. Endothelial cells expr ess eNOS and iNOS, and cardiomyocytes nNOS. Further on, the inner vation and vascular str uctures in all tissues expr ess nNOS and eNOS, while circulating cells may express iNOS.

In order to be activated, NOS pr oteins have to bind cofactors and dimerize (9). eNOS consists of two identical monomers, and each monomer has two principal domains: C-terminal reductase domain with binding sites for NADPH, FMN, and FAD, and N-terminal oxidase domain which takes away an electr on from L-arginine and contains binding sites for (ir on) hem, BH4, and L-arginine (9,10). NOS proteins first bind the FAD and FMN cofactors. The addition of L arginine, BH4, and hem enables the proteins to form dimers. Hem is essential for the process of dimerization. The lack of these cofactors leads to NOS dysfunction (11). Low concentration or lack of L-arginine catalyzes the reduction of oxygen to super oxide or hydrogen peroxide (12), and dr op of BH4 level to simultaneous production of NO and super oxide, the products which may interact to for m peroxynitrite (13). The formed dimers eNOS and nNOS ar e inactive till the binding of calmodulin, effectuated via the increased concentration of the intracellular calcium. iNOS is activated even in low calcium concentrations due to high enzyme affinity towards calmodulin (14). This means that the main »switch« for the activa-

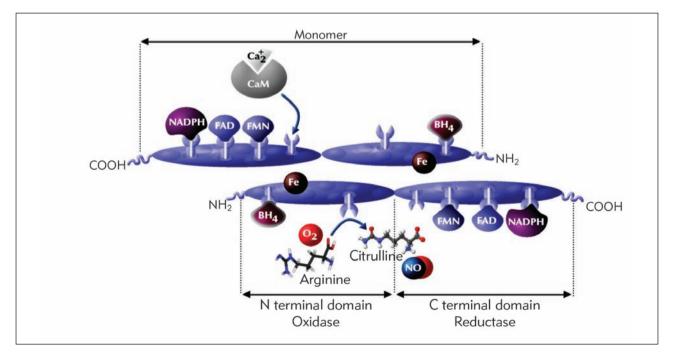


Figure 1 Model proposed for the dimeric eNOS structure.

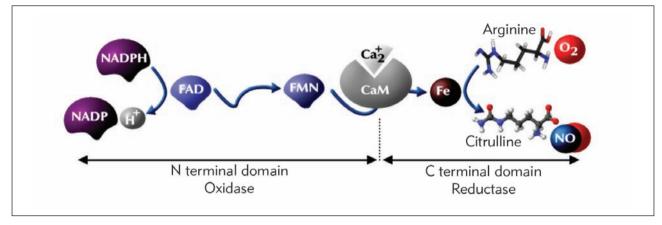


Figure 2 Electron transfer between cofactors and substrate of the enzyme str ucture.

tion of nNOS and eNOS is a transient elevation of calcium concentration, and for iNOS, the transcription process.

The process of catalysis of the constitutive NOS involves two oxidation stages: first, L -arginine is hydroxylated to NG -hydroxy-L-arginine, and then the oxidation of this inter mediary occurs with the use of one electron of NADPH and for mation of L-citrulline and NO (15). In the r eaction, 1.5 moles of NADPH and 2 moles of oxygen per a mole of citr ulline are consumed (16).

# Regulation of eNOS activity and gene expression

eNOS gene is expressed constitutively thus providing the basal concentration of eNOS pr otein. Stable concentrations of mRNA ar e maintained by the complex regulatory mechanism of gene expr ession (17). The eNOS gene promoter has multiple cisregulatory DNA sequences, including CCAT box, Sp1 sites, GATA motifs, CACCC box, AP-1 and AP-2 sites, p53 binding sites, NF-1 elements and the sequences responsive to sterol elements and shear str ess (18). The positive regulatory domains I and II (PRDI and PRDII) are located in the pr oximal promoter and involved in the baseline regulation of gene transcription by transcription factors such as Sp-1, Sp-3, Ets-1, Elf-1 YY1 and MY C-associated zinc finger pr otein (19). Shear stress activation of the promoter is mediated by NF B which binds to the r esponsive element GAGACC located upstream of the transcription side (20). Laminar flow shows a nine-fold incr ease of mRNA in bovine endothelial cells, and this effect is mediated by the transient increase of gene transcription and prolonged mRNA half-life (21).

The post-transcription r egulation is accomplished by the dimerization of the protein subunits of NOSs and by the interaction with the caveolin protein and heat shock protein hsp90 (22). The nNOS gene encodes a PDZ domain in exon 2 that is required for membrane association. During nNOS splicing several variants may be formed lacking exon 2 thus resulting in the expression of cytoplasmic nNOS that lacks subcellular localization sequences (2). In the endothelial cells eNOS is localized to caveolae by N-terminal fatty acid modifications as well as interactions with the heat shock protein hsp90 and caveolins (23). eNOS binds to caveolin-1 in the endothelial cells, and in the cardiac muscle to caveolin-3. eNOS activation r equires eNOS trafficking from the plasma membrane to the Golgi apparatus and enzyme phosphor vlation of the amino acid serine at position 1177 by the Akt. PKA or AMP kinases, which is the main mechanism of eNOS activation and increases in eNOS sensitivity to basal concentrations of calcium/calmodulin (24, 25). Tonic or phasic eNOS activation in r esponse to blood flow is independent of calcium concentration changes and constitutes the shear str ess. Although phosphorylation of the serine 1177 r esidue plays an essential role in eNOS enzyme activation, its r egulation is dependent on the phosphor ylation of other amino acid residues such as the serine633 r esidue, which also incr eases eNOS activity, or the thr eonine495 residue, which inter feres with the calmodulin binding domain, thus down r egulating NO synthesis (22).

### **Molecular targets of NO**

In many cells and for numer ous signaling roles of NO, the physiologic tar get is soluble guanylate cyclase (26). NO activates guanylate cyclase by binding to the hem ir on, which increases cGMP level in the cell. Via cGMP, NO leads to the relaxation of vascular smooth muscles and vasodilatation. In the brain, NO activates NMDA receptors, and in autonomous nerve system it pr oduces a transmitter which mediates the relaxation of smooth muscles of the gastrointestinal, urinary, and respiratory tract. The next NO target is sulphydryl groups of proteins with which it cr eates nitrosothiols (27). Nitrosylated hemoglobin serves as a natural transporter and pool of NO (28). In the car diac muscle, NO nitrosylates the SH group of ryanodine receptor in the membrane, thus activating the r eceptor (29). Nitrosylation of N-ethylmaleimide-sensitive factor is significant for the r egulation of exocytosis. P eroxynitrite anion is created in the reaction with superoxide.

In the case of pr oduction of large amounts of NO, which occurs in iNOS activation, this molecule can directly inhibit mitochondrial complexes I and IV. It induces energetic depletion in the cell by way of poly(ADP-riboso)polymerase activation (2). In gene - ral, biologic r oles of NOS ar e effectuated via the effects of soluble guanylate cyclase and S-nitr osylation of proteins, while other mechanisms lead to toxic effects of NO.

### **Endothelial dysfunction and NO**

Vascular endothelium is an active or gan which in physiologic conditions expresses a number of useful effects, such as vasodilatator y, antioxidative, antiinflammatory, anticoagulant, and profibrinolytic ones; it inhibits adhesion and migration of leukocytes, in hibits proliferation and migration of smooth muscle cells; inhibits aggregation and adhesion of thrombocytes (30). These atheroprotective effects depend on the balance of substances synthesized and r eleased by healthy endothelium, among which the most important vasoactive component is NO, critical in the pathophysiology of vascular disease and endothelial dysfunction concept. The disor der of endotheliumdependent vasodilatation is a systemic disorder recognized as endothelial dysfunction leading to ather 0sclerosis and its complications due to absence of normal endothelial functions. It occurs in ather osclerosis, hypertension, hyper cholesterolemia, and the process of normal aging (2, 31). Endothelial dysfunction in atherosclerotic coronary arteries was described for the first time by Ludmer et al. (32), and its association with the bioavailability of nitric oxide was later described as well. Reduced bioavailability of NO is the most important mechanism in the multifactorial pr ocess of the endothelial dysfunction and is involved in most important cardiovascular dysfunctions.

There are several potential mechanisms leading to endothelial dysfunction that can be divided into three categories: r eduction of eNOS expr ession, reduction of eNOS activity, and rapid elimination of NO. First, altered expression of mRNA for eNOS or protein synthesis lead to reduced eNOS activity (33). However, most evidence from the animal models and humans suggest an increased eNOS concentration in diabetes and ather osclerosis rather than decr eased one. Second, the L -arginine substrate may be deficient in the tissues or its transport to the cells may be disturbed. The presence of endogenous competitive inhibitor asymmetric dimethylar ginine (ADMA) may reduce the production of NO, even in the presence of physiologic substrate concentrations (34). Third, eNOS requires numerous cofactors for its activity . BH4 is especially important among them, the synthesis of which is controlled by GTP cyclohydrolase, and in the absence of which the transport of electr ons through eNOS becomes uncoupled, and superoxide is created instead of NO (35). Fourth, in order to be activated, eNOS has to be dimerized and adequately localized in the caveolae via caveolin and hsp90 (36). F ifth, eNOS is phosphorylated in the S1179 position via Akt or some other kinase (37). Sixth, with dysfunctional epithelium, there is an incr eased ROS production, above all via NADPH oxidase activity or uncoupling of eNOS, which may reduce NO levels via several different pathways: direct inactivation by super oxide, with peroxynitrite formation (38), reduction of expression and activity of NOS as the consequence of substrate or cofactor reduction, due to increased ADMA concentration (39), and due to uncoupling of NOS induced by the incr eased oxidation of tetrahydr obiopterin (24).

The results of recent studies have demonstrated that some drugs, such as antioxidants and r enin-angiotensin system blockers, may r educe the endothelial dysfunction via the mechanism of activation of eNOS by phosphorylation of the amino acid residues in specific positions. Dias et al. (22) have shown that Talmisartan, a blocker of angiotensin II r eceptor, reduces endothelial dysfunction by eNOS activation, via phosphorylation of serine residues in the positions 1177 and 635.

Since eNOS plays a significant role in the regulation of blood vessel function, an excessive NO pr oduction may contribute to development of atherosclerosis. The sour ce of NO may be iNOS and nNOS expressed in blood vessel smooth muscle cells in the atherosclerotic lesions, as well as iNOS expr essed in the activated macr ophages and monocytes (40). These isoforms produce NO, which with peroxynitrite, can increase the oxidative stress and oxidative modification of LDL particles (41). The evidence of the presence of per oxynitrite in human ather osclerotic lesions is the finding of nitr otyrosine. NO can affect redox-sensitive transcription of the genes involved in the process of activation of the endothelial cells (42).

# Risk factors for ischemic heart disease and NO

### Hypertension and NO

Blood pressure is controlled via the interaction of several homeostatic r egulatory mechanisms, in cluding the r enin-angiotensin system, autonomous nerve system, and the local mediators such as NO. The role of NO in the regulation of blood pressure is very important. NOS inhibition induces blood pr essure elevation in many animal species (43). Blood pressure in eNOS knockout mice is 30% higher than in their wild-type counterparts. It is still unclear why other homeostatic mechanisms cannot compensate for eNOS. One of possible explanations is that the renin-angiotensin system and the autonomous ner ve system serve primarily to prevent hypertension. Alternatively, there have been infor mation that eNOS is involved in the control of baroreceptors (44). Hypertension is associated with the incr eased release of vasoconstrictive endothelial mediators, including the angiotensin II as one of the most potent vasoconstrictors which induces the pr oduction of endothelin via the MAP kinase pathway (45). It also stimulates ROS production (46) and increases BH4 consumption, inhibiting NO production. The question could be rightfully asked whether endothelial dysfunction in hypertension is a cause or a consequence. On one hand, there are evidence indicating the defective phosphoinositol, NOS-activating pathway, and the suggestion that it is responsible for endothelial dysfunction in the essential hypertension (47). On the other hand, Zizek (48) has demonstrated that ther e is an endothelial dysfunction in normotensive children of hypertensive patients, and there are evidence of the association of the essential hypertension with the eNOS gene polymorphism (49, 50). Since endothelial dysfunction in hypertension is genetically deter mined, hypertension could be the cause of endothelial dysfunction. How ever, the endothelial dysfunction is encounter ed in patients with secondar y hypertension as well, so it could well be the consequence of hypertension. In view of the above facts, the endothelial dysfunction could equally be the cause and consequence of hypertension.

### Dyslipidemia, NO and atherosclerosis

It has been well known that some of the abnormalities of lipid metabolism, mostly high cholester ol and/or triglyceride levels, are usually encountered in most patients with the ischemic heart disease (IHD). The incidence of IHD in individuals with the disturbed lipoprotein metabolism is 60%-65% (51). Ather ogenic lipoproteins are the crucial factors in the initiation and promotion of ather osclerosis, the direct conseguence of which is IHD. A cute and chronic manifestations of ather osclerosis are the result of chronic inflammation, partly initiated and maintained by LDL particles penetrating into the subendothelial space from the circulation, where they are oxidatively modified (oxLDL) (52). By the activation of NF B, oxLDL induce the expression of the adhesion molecules in endothelial cells, enabling the adhesion of circulating inflammatory cells to the endothelium and their transition into the subendothelial space (53). The elevation of L-selectin, a vascular cellular adhesion molecule-1 and inter cellular adhesion molecule-1 in the sera of patients with cor onary arterial disease is an

indicant of the endothelial activation and dysfunction. Endothelial activation can be induced by triglyceriderich lipoproteins, such as chylomicr on remnants and VLDL remnants penetrating the endothelium and reaching the intima. In a prospective cohort study on 7587 women and 6394 men, it has been shown that the increased triglyceride concentration is associated with the increased risk of IHD, myocar dial infarction, and mortality in both genders (54).

Highly reactive free oxygen radicals are released in the blood vessel wall fr om inflammatory and also endothelial cells themselves (55), which oxidatively modify lipoproteins. In the culture of endothelial cells, eNOS may produce large amounts of super oxide after the addition of LDL particles to the medium (56). Increased LDL and decreased HDL concentrations induce disintegration of the caveolae complex, where NOS is bound (57). V ascular smooth muscle cells of rats, in which hypertension is induced by angiotensin II, also produce superoxide by the activation of membrane NADPH oxidase (58). Super oxide and other oxygen radicals may oxidize NO to the metabolites which cannot activate guanylate cyclase, being potentially toxic to the endothelium (such as peroxynitrite). The fact that ather osclerotic rabbit aorta produces more NO supports the notion that dysfunctional endothelium synthesizes mor e NO compared to normal one. oxLDL particles also stimulate transcription and synthesis of eNOS (59). An increased iNOS expression, producing large amounts of NO, has been demonstrated in human atherosclerotic plagues (60). These findings show that vascular cells in hypercholesterolemia and atherosclerosis synthesize more NO than dormant cells, but NO is rapidly inactivated or converted into toxic oxides due to increased production of free radicals. On the other hand, oxLDL may inhibit NOS and consequentially reduce NO production (61). Moreover, an increased production of ADMA, demonstrated in persons with hypercholesterolemia, competitively inhibits NOS (62). The increases of lipoproteins (a), encountered in the impaired coronary endothelium function (63), inhibits NOS with its oxidatively modified components or oxidizes and inactivates NO (64). R educed NO may stimulate the synthesis and r elease of the endothelin and proinflammatory cytokines, release of growth factors, hyperplasia, and migration of the smooth muscle cells and thrombocyte adhesion to the endothe lium. All these consequences of endothelial dysfunction are significant in the initiation, pr ogression and clinical manifestation of atherosclerosis, i.e. IHD (65).

## Diabetes and NO

In thin individuals, the insulin stimulates blood flow and reduces vascular resistance in the skeletal muscles (66). Using L-NMMA and by BH4 synthesis inhibition, it has been shown that blood flow stimulation and release of glucose are NO-mediated (67). In healthy individuals, the insulin increases NOS activity stimulating the phosphatidylinositol-3 kinase and Akt kinase. In insulin r esistant patients, the signal transduction is disturbed via phosphatidylinositol-3 kinase pathway, responsible also for glucose uptake by the cells. Due to r educed stimulation of NOS by the insulin, NO production is reduced, and consequentially the endothelium-dependent vasodilatation (68). However, the transduction by insulin via MAPK is preserved, resulting in the enhanced pr oduction of endothelin and stimulation of the inflammation and thrombosis (70). Ther eupon, hypertension occurs associated with the incr eased ADMA concentration (71). The occur rence of metabolic disor ders (oxidatively modified LDL) leads to downregulation of eNOS expression (72). Clinical studies with A CE inhibitors and statins have shown that these dr ugs not only reduce coronary disease and mortality fr om cardiovascular diseases, but also pr event the development of diabetes type 2 (73, 74), confir ming the role of endothelial dysfunction in the pathophysiology of the insulin resistance.

### References

- 1. Bredt DS, Snyder SH. Nitric oxide: a physiologic messenger molecule. Ann Rev Biochem 1994; 63: 175–95.
- 2. Liu VWT, Huang PL. Cardiovascular roles of nitric oxide: a review of insights from nitric oxide synthase gene disrupted mice. Cardiovas Res 2008; 77: 19–29.
- Govers R, Rabelink TJ. Cellular regulation of endothelial nitric oxide synthase. Am J Physiol R enal Physiol 2001; 280: F193–206.
- Stuehr DJ. Structure-function aspects in the nitric oxide synthases. Annu R ev Pharmacol Toxicol 1997; 37: 339–59.
- Ferreiro CR, Chagas AC, Carvalho MH, Dantas AP, Scavone C, Souza LC, et al. Expr ession of inducible nitric oxide synthase in increased in patients with heart failur e due to ischemic disease. Braz J Med Biol R es 2004; 37: 1313–20.
- 6. Andrew PJ, Mayer B. Enzymatic function of nitric oxide synthases. Cardiovasc Res 1999; 43: 521–31.
- Barouch LA, Harrison RW, Skaf MW, Rosas GO, Cappola TP, Kobeissi ZA, Hobai IA, Lemmon C A, Burnett AL, O'Rourke B, Rodriguez ER, Huang PL, Lima JA, Berko witz DE, Hare JM. Nitric oxide regulates the heart by spatial confinement of nitric oxide synthase isoforms. Nature 2002; 416(6878): 337–9.
- Balligand JL, Kobzik L, Han X, K aye DM, Belhassen L, O'Hara DS, Kelly RA, Smith TW, Michel T. Nitric oxidedependent parasympathetic signaling is due to activation of constitutive endothelial (type III) nitric oxide synthase in cardiac myocytes. J Biol Chem 1995; 270(24): 14582–6.
- 9. Alderton WK, Cooper CE, Knowles RG. Nitric oxide syn-

Hyperglycemia, accompanying the diabetes, increases superoxide production in the electr on-transport chain in the mitochondria (73). Super oxide activates protein kinase C, and the kinase activates NADPH oxidase to pr oduce even more superoxide. The reaction of NO and super oxide produces peroxynitrite which oxidizes BH4, uncoupling NOS, which produces superoxide instead of NO. Superoxide increases the production of the advanced glycation end products (74), and they incr ease the production of superoxide and other ROS, r educing thus NO. The resulting hyperglycemia-induced oxidative str ess inhibits dimethylaminohydrolase (DDAH) (75), with consequential ADMA increase and the final r esult of reduced NO synthesis.

### **Conflict of interest statement**

The authors stated that ther e were no conflicts of interest regarding the publication of this article.

thases: structure, function and inhibition. Biochem J 2001; 357: 593–615.

- 10. Andrew PJ, Mayer B. Enzymatic function of nitric oxide synthases. Cardiovasc Res 1999; 43: 521–31.
- Vásquez-Vivar J, Kalyanaraman B, Martásek P, Hogg N, Masters BS, Karoui H, Tordo P, Pritchard KA. Superoxide generation by endothelial nitric oxide synthase: the influence of cofactors. P roc Natl A cad Sci U S A 1998; 95(16): 9220–5.
- Rabelink TJ, Luscher TF. Endothelial nitric oxide synthase: host defense enzyme of the endothelium. Arterioscler Thromb Vasc Biol 2006; 26: 267–1.
- Beckman JS, K oppenol WH. Nitric oxide, super oxide, and peroxynitrite: the good, the bad, and ugly . Am J Physiol 1996, 271: C1424–37.
- Michel T, Feron O. Nitric oxide synthases: which, where, how, and why? J Clin Invest 1997; 100: 2146–52.
- Albrecht EW, Stegeman CA, Heeringa P, Henning RH, van Goor H. Protective role of endothelial nitric oxide synthase. J Pathol 2003; 199: 8–17.
- Korth HG, Sustmann R, Thater C, Butler AR, Ingold KU. On the mechanism of the nitric oxide synthase-catalyzed conversion of N omega-hydr oxyl-L-arginine to citr ulline and nitric oxide. J Biol Chem 1994; 269(27): 17776–9.
- Searles CD. Transcriptional and posttranscriptional regulation of endothelial nitric oxide synthase expression. Am J Physiol Cell Physiol. 2006; 291(5): C803–16.
- Marsden J, Heng HH, Scherer SW, Stewart RJ, Hall AV, Shi XM, et al. Structure and chromosomal localization of the human constitutive endothelial nitric oxide synthase gene. J Biol Chem 1993; 268: 17478–88.

- Karantzoulis-Fegaras F, Antoniou H, Lai SL, Kulkarni G, D'Abreo C, Wong GK, et al. Characterization of the human endothelial nitric-oxide synthase promoter. J Biol Chem 1999; 274: 3076–93.
- Davis ME, Grumbach IM, Fukai T, Cutchins A, Harrison DG. Shear stress regulates endothelial nitric-oxide synthase promoter activity through nuclear factor kappaB binding. J Biol Chem 2004; 279: 163–8.
- Davis ME, Cai H, Drummond GR, Har rison DG. Shear stress regulates endothelial nitric-oxide synthase expression through c-Src by divergent signaling pathways. Circ Res 2001; 89: 1073–80.
- Dias RG, Negrao CE, Krieger MH. Nitric oxide and cardiovascular system: cell activation, vascular reactivity and genetic variant. Arq Bras Cardiol 2011; 96(1): 68–75.
- Garcia-Cardena G, Oh P, Liu J, Schnitzer JE, Sessa WC. Targeting of nitric oxide synthase to endothelial cell caveolae via palmitoylation: implications for nitric oxide signaling. Proc Natl Acad Sci USA 1996; 93: 6448–53.
- 24. Sessa WC. eNOS at a glance. J Cell Sci 2004; 117: 2427–9.
- Boo YC, Kim HJ, Song H, Fulton D, Sessa W. Coordinated regulation of endothelial nitric oxide synthase activity by phosphorylation and subcellular localization. Free Radic Biol Med 2006; 41(1): 144–53.
- 26. Snyder SH, Bredt DS. Biological roles of nitric oxide. Sci Am 1992; 266: 68–77.
- 27. Stamler JS. Redox signaling: nitrosylation and related target interactions of nitric oxide. Cell 1994; 78. 931–6.
- Stamler JS, Jia L, Eu JP, Mcmahon TJ, Demchenko IT, Bonaventura J, et al. Blood flow r egulation by S-nitrosohemoglobin in the physiological oxygen gradient. Science 1997; 276: 2034–37.
- Xu L, Eu JP, Meissner G, Stamler JS. Activation of the cardiac calcium r elease channel (r yanodine receptor) by poly-S-nitrosylation. Science 1998; 279: 234–7.
- Forstermann U. Janus-faces role of endothelial NO synthase in vascular disease: uncoupling of oxygen r eduction from NO synthesis and its pharmacological reversal. Biol Chem 2006; 387(12): 1521–33.
- Cai H, Harrison DG. Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. Circ Res 2000; 87: 840–44.
- Ludmer PL, Selwyn AP, Shook TL, W ayne RR, Mudge GH, Alexander RW, et al. P aradoxical vasoconstriction induced by acetylcholine in ather osclerotic coronary arteries. N Engl J Med 1986; 315: 1046–51.
- 33. Wang Y, Marsden PA. Nitric oxide synthases: gene structure and regulation. Adv Pharmacol 1995; 34: 71–90.
- Cooke JP. Does ADMA cause endothelial dysfunction? Arterioscler Thromb Vasc Biol 2000; 20: 2032–7.
- Cosentino F, Patton S, d'Uscio L V, Werner ER, Werner-Felmayer G, Moreau P, et al. Tetrahydrobiopterin alters superoxide and nitric oxide r elease in pr ehypertensive rats. J Clin Invest 1998; 101: 1530–7.

- Shaul PW. Regulation of endothelial nitric oxide synthase: location, location, location. Annu Rev Physiol 2002; 64: 749–774.
- Fulton D, Gratton JP, McCabe TJ, F ontana J, F ujio Y, Walsh K, et al. R egulation of endothelium-derived nitric oxide production by the protein kinase Akt. Nature 1999; 399: 597–601.
- Gao L, Mann GE. V ascular NAD(P)H oxidase activation in diabetes: a double-edged swor d in r edox signaling. Cardiovasc Res 2009; 82(1): 9–20.
- De Gennaro Colonna V, Bianchi M, Pascale V, Ferrario P, Morelli F, Pascale W, et al. Asymmetric dimethylar ginine (ADMA): an endogenous inhibitor of nitric oxide synthase and a novel car diovascular risk molecule. Med Sci Monit 2009; 15(4): RA91–101.
- Sobey CG, Br ooks RM, Heistad DD. Evidence that expression of inducible nitric oxide synthase in r esponse to endotoxin is augmented in atherosclerotic rabbits. Circ Res 1995; 77: 536–43.
- Darley-Usmar VM, Hogg N, O 'Leary VJ, W ilson MT, Moncada S. The simultaneous generation of super oxide and nitric oxide can initiate lipid per oxidation in human low density lipoprotein. Free Radic Res Commun 1992; 17(1): 9–20.
- 42. Marui N, Offermann MK, Swerlick R, K unsch C, Rosen CA, Ahmad M, Alexander RW, Medford RM. Vascular cell adhesion molecule-1 (VC AM-1) gene transcription and expression are regulated through an antioxidant-sensitive mechanism in human vascular endothelial cells. Clin Invest 1993; 92(4): 1866–74.
- 43. Isakuma I, Togashi H, Yoshioka M, Saito H, Yanagida M, Tamura M. NG-methyl-L-arginine, an inhibitor of L-arginine-derived nitric oxide synthesis, stimulates r enal sympathetic nerve activity in vivo. A role for nitric oxide in the central regulation of sympathetic tone? Cir c Res 1992; 70: 607–11.
- Matsuda T, Bates JN, Lewis SJ, Abboud FM, Chapleau MW. Modulation of bar oreceptor activity by nitric oxide and S-nitrosocysteine. Circ Res 1995; 76: 426–33.
- 45. Cengel A, Sahinarslan A. Nitric oxide and car diovascular system. Anadolu Kardiyol derg 2006; 6: 364–8.
- Griending CC, Minieri C A, Ollerenshaw JD, Alexander RW. Angiotensin II stimulates NADPH oxidase activity in cultured vascular smooth muscle cells. Cir c Res 1994; 74: 1141–8.
- Cardillo C, Kilcoyne CM, Quyyumi A A, Cannon RO, Panza JA. Selective defect in nitric oxide synthesis may explain the impair ed endothelium-dependent vasodilation in patients with essential hypertension. Cir culation 1998; 97: 851–6.
- Zizek B, Poredos P, Videenik V. Endothelial dysfunction in hypertensive patients and in nor motensive offspring subjects with essential hypertension. Heart 2001; 85: 215–7.
- Shoji M, Tsutaya S, Saito R, T akamatu H, Yasujima M. Positive association of endothelial nitric oxide synthase gene polymorphism with hypertension in northern Japan. Life Sci 2000; 66: 2557–62.

- Miyamoto Y, Saito Y, Kajiyama N, Yoshimura M, Shimasaki Y, Nakayama M, et al. Endothelial nitric oxide synthase gene is positively associated with essential hypertension. Hypertension 1998; 32: 3–8.
- 51. Wada M, Wada K, Mise J. Ischemic heart disease and hyperlipidemia. Jpn Circ J 1975; 39(3): 325–30.
- Cannon RO. Role of nitric oxide in car diovascular disease: Focus on the endothelium. Clin Chem 1998; 44(8): 1809–19.
- Cybulsky MI, Gimbrone MA. Endothelial expression of a mononuclear leukocyte adhesion molecule during atherogenesis. Science 1991; 252: 788–91.
- Nordestgaard BG, Ben M, Schnohr P, Tybjaerg-Hansen A. Nonfasting triglycerides and risk of myocar dial infarction, ischemic heart disease, and death in men and women. JAMA 2007; 298(3): 299–308.
- Ohara Y, Peterson TE, Harrison DG. Hypercholesterolemia increases superoxide anion production. J Clin Invest 1993; 91: 2541–51.
- Pritchard KA Jr, Groszek L, Smalley DM, Sessa WC, W u M, Willalon P, et al. Native low -density lipoprotein increases endothelial cell nitric oxide synthase generation of superoxide anion. Circ Res 1995; 77: 510–8.
- 57. Drab M, Verkade P, Elger M, Kasper M, Lohn M, Lauterbach B, et al. Loss of caveolae, vascular dysfunction and pulmonary defects in caveolin-1 gene-disr upted mice. Science 2001; 293: 2449–52.
- Rajagopalan S, Kurz S, Münzel T, Tarpey M, Freeman BA, Griendling KK, Har rison DG. Angiotensin II-mediated hypertension in the rat increases vascular superoxide production via membrane NADH/NADPH oxidase activation. Contribution to alterations of vasomotor tone. J Clin Invest 1996; 97: 1916–23.
- 59. Hirata K, Miki N, K uroda Y, Sakoda T, Kawashima S, Yokoyama M. Low concentration of oxidized low -density lipoprotein and lysophosphatidylcholine upr egulate constitutive nitric oxide synthase mRNA expression in bovine aortic endothelial cells. CircRes 1995; 76: 958–62.
- Buttery LDK, Springall DR, Chester AH, et al. Inducible nitric oxide synthase is present within human atherosclerotic lesions and promotes the formation and activity of peroxynitrite. Lab Invest. 1996; 75: 77–85.
- Shimokawa H, Flavahan NA, van Houtte PM,. Loss of endothelial pertussis toxin-sensitive G protein function in atherosclerotic porcine coronary arteries. Circulation 1991; 83: 652–60.
- Boger RH, Bode-Boger SM, Szuba A, Tsao PS, Chan JR, Cooke JP. Asymmetric dimethylarginine: a novel risk factor for endothelial dysfunction. Cir culation 1997; 96: I-173.
- Tsurumi Y, Nagashima H, Ischkawa K, Sumiyoshi T, Hososda S. Influence of plasma lipopr otein(a) levels on coronary vasomotor response to acetylcholine. J Am Coll Cardiol 1995; 26: 1242–50.

- Galle J, Bengen J, Schollmeyer P, Wanner C. Impairment of endothelium-dependent dilation in rabbit renal arteries by oxidized lipoprotein(a). Role of oxygen-derived radicals. Circulation 1995; 92: 1582–9.
- 65. Sypniewska G, Ber gmann K, Krintus M, K ozinski M, Kubica J. How do apolipoproteins ApoB and ApoA-I perform in patients with acute coronary syndromes. Journal of Medical Biochemistry 2011; 30: 237–43.
- Cook S. Coronary artery disease, nitric oxide and oxidative stress: the »Yin-Yang« effect-a Chinese concept for a worldwide pandemicdisease. Swiss Med Wkly 2006; 136: 103–13.
- Scherrer U, R andin D, Vollenweider P, Vollenweider L, Nicod P. Nitric oxide release accounts for insulin's vascular effects in humans. J Clin Invest 1994; 94: 2511–5.
- Scherrer U, Sartori C. Insulin as a vascular and sympathoexcitatory hormone. Implications for blood pr essure, insulin sensitivity and car diovascular morbidity. Circulation 1997; 96: 4104–13.
- De Vriese AS, Verbeuren TJ, Van de Voorde J, Lameire NH, Vanhoutte PM. Endothelial dysfunction in diabetes. Br J Pharmacol 2000; 130: 963–74.
- Hübner-Woźniak E, Okecka-Szymańska J, Stupnicki R, Malara M, K ozdroń E. Age-Related blood antioxidant capacity in men and women. Jour nal of Medical Bio chemistry 2011; 30: 103–8.
- Ito A, Tsao PS, Adimoolam S, Kimoto M, Ogawa T, Cook JP. Novel mechanism for endothelial dysfunction: dysregulation of dimethylar ginine dimethylaminohydrolase. Circulation 1999; 99: 3092–5.
- 72. Lamarche B, Lemieux I, Despr es JP. The small, dense LDL phenotype and the risk of cor onary heart disease: epidemiology, pathophysiology and therapeutic aspects. Diabetes Metabol 1999; 25: 199–211.
- 73. Freeman DJ, Norrie J, Sattar N, Neely RD, Cobbe SM, Ford I, et al. Pravastatin and the development of the diabetes mellitus: evidence for a protective treatment effect in the W est Scotland Cor onary prevention Study. Circulation 2001; 103: 357–62.
- 74. Yusuf S, Sleight P, Pogue J, Bosch J, Davies R, Dagenais G. Effects of an angiotensin-converting-enzyme inhibitor, ramipril, on car diovascular events in high-risk patients. The Heart Outcomes P revention Evaluation Study In vestigators. N Engl J med 2000; 342: 145–53.
- Nishikawa T, Edelstein D, Du XL, Yamagishi S, Matsumura T, Kaneda Y, et al. Nor malizing mitochondria superoxide production blocks three pathways of hyperglycemic damage. Nature 2000; 404: 787–90.
- Lin KY, Ito A, Asagami T, Tsao PS, Adimoolam S, Kimoto M, et al. Impair ed nitric oxide synthase pathway in diabetes mellitus: role of a symmetric dimethylar ginine and dimethylaminohydrolase. Circulation 2002; 106: 987–92.

Received: May 15, 2012 Accepted: June 6, 2012