

JMB 28: 166–171, 2009

Original paper  
Originalni naučni rad

## IMMUNE COMPLEXES AND COMPLEMENT IN SERUM AND SYNOVIAL FLUID OF RHEUMATOID ARTHRITIS PATIENTS

IMUNSKI KOMPLEKSI I KOMPLEMENT U SERUMU I SINOVIJALNOJ TEČNOSTI KOD BOLESNIKA SA REUMATOIDNIM ARTRITISOM

*Zoran Mijušković<sup>1</sup>, Ljiljana Rackov<sup>2</sup>, Janko Pejović<sup>1</sup>, Sandra Živanović<sup>4</sup>,  
Jelica Stojanović<sup>5</sup>, Zoran Kovačević<sup>3</sup>*

<sup>1</sup>*Institute of Medical Biochemistry,*

<sup>2</sup>*Clinic for Rheumatology and Clinical Immunology,*

<sup>3</sup>*Clinic for Nephrology, Military Medical Academy, Belgrade,*

<sup>4</sup>*Primary Health Center, Kragujevac,*

<sup>5</sup>*Faculty of Mathematics and Natural Sciences, University of Kragujevac, Serbia*

**Summary:** Rheumatoid arthritis (RA) is predominantly an intraarticular inflammatory and autoimmune disease that involves different autoantibodies and effector mechanisms. The aim of the study was to determine the utility of Circulating Immune Complexes (CIC) and complement components (C3c, C4) as possible markers for the disease activity in laboratory diagnostics. In a cross-section study 59 patients, according to the clinical criteria, were categorized into two groups: group with moderate (MA, n=24), and group with severe activity (SA, n=35) of RA. The concentration of CIC, C3c and C4 in sera (S) and synovial fluids (SF) was examined by an immunonephelometric method in both groups and compared with values in the control group (n=15) of patients with lesions of the menisci. Obtained results showed that there was no statistical significance in the values of C3c and C4, in both biological fluids, among all tested groups. Significant differences were found in the levels of CIC in both fluids, while testing the parameters ( $\bar{x} \pm SD$ , IU/mL) in the sera of groups with SA and MA of RA:  $7.43 \pm 13.40$ ;  $3.01 \pm 2.92$  ( $p < 0.05$ ) and SF:  $13.47 \pm 21.1$ ,  $5.33 \pm 7.53$  ( $p < 0.001$ ), respectively. These differences were higher between the group with SA and CG. Results for the concentrations of CIC were significantly higher in SF compared to sera: in the RA group with SA by 77% and group with MA by about 82%. These data could provide a confirmation of the

**Kratak sadržaj:** Reumatoидни artritis (RA) jeste predomniantno intraartikularna zapaljenska i autoimunska bolest u koju su uključena različita autoantitela i efektorni mehanizmi. Cilj ispitivanja je bio da se ustanovi značaj cirkulirajućih imunskih kompleksa (CIK) i komponenti komplementa (C3c, C4), kao pokazatelja stepena aktivnosti RA za laboratorijsku dijagnostiku. U studiji preseka stanja je ispitano 59 bolesnika koji su prema kliničkim kriterijumima za aktivnost RA podeljeni u dve grupe: grupu sa umerenom (UA, n=24) i grupu sa visokom aktivnošću (VA, n=35) RA. Koncentracije CIK, C3c i C4 u serumu i sinovijalnoj tečnosti (ST) određivane su imunonefelometrijskom metodom u obe grupe ispitanih i upoređene sa vrednostima u kontrolnoj grupi od 15 pacijenata s povredama meniskusa. Rezultati su pokazali da nije bilo statistički značajnih razlika u koncentracijama za C3c i C4 u oba biološka uzorka između ispitivanih grupa. Statistički značajne razlike u koncentracijama CIK utvrđene testiranjem vrednosti ( $\bar{x} \pm SD$ , IU/mL) u serumu između grupe sa VA i grupe sa UA RA bile su:  $7.43 \pm 13.40$ ;  $3.01 \pm 2.92$  ( $p < 0.05$ ) i za vrednosti u ST:  $13.47 \pm 21.1$ ,  $5.33 \pm 7.53$  ( $p < 0.001$ ). Razlike su bile više izražene između grupe sa VA RA i KG. Rezultati koncentracija CIK su bili značajno viši u ST u odnosu na serum u obe grupe bolesnika: u grupi sa UA za 77% a u grupi sa VA RA za 82%. Ti podaci idu u prilog potvrđi hipoteze o lokalnoj, intraartikularnoj produkciji autoantitela, odnosno

---

Address for correspondence:

Zoran V. Mijušković  
Military Medical Academy  
Institute of Medical Biochemistry  
Crnotravska 17, 11002 Belgrade, Serbia  
e-mail: zmijusko@gmail.com

hypothesis about local, intraarticular autoantibodies and subsequent CIC production. It can be concluded that the examination of CIC concentration in serum, and where it is possible in SF, is a useful marker of disease activity in RA patients, in contrast to the tested components of the complement. This statement does not exclude their consumption within immune effector mechanisms, but elicits the possibility that lower molecular fragments (C3d, C4d), as well as the novel activation products, could be better disease activity markers in RA patients.

**Keywords:** rheumatoid arthritis, circulating immune complexes, complement components, disease activity

## Introduction

Rheumatoid arthritis (RA) is a chronic autoimmune synovitis affecting about 1.0% of the world's population (1), but the mechanisms underlying the initiation and progression of RA are not yet completely understood. The presence of immune complexes (ICs) in the blood and synovial fluid of patients with RA has been well described (2, 3), and there is evidence that they are involved in the activation of the complement cascade in RA synovial tissue (5). However, apart from rheumatoid factor (RF-IgM) and different RF isotypes (6–8), other autoantibodies have been added to the growing list of autoantigens such as: filaggrin (10), Sa (11, 12), calpastatin (13), RA33 (14), collagen type II (15) and a number of citrullinated proteins (16–18). Due to their ability to form ICs, they may promote complement activation. The ICs from RA synovial fluid can induce cytokine and RF production (19), but the identity of antigens involved in ICs remains obscure. In addition to dietary factors, speculation on and investigation of RA etiology has also been focused on genetic susceptibility, abnormal bowel permeability and microorganisms. Therefore, RA is a classic example of a multifactorial disease in which a combination of genetic and environmental factors contributes to the disease progression.

The aim of this study was to evaluate the concentrations of circulating immune complexes (CIC), C3 and C4 complement components of the classical pathway as possible biochemical parameters for disease activity and progression of RA. Taking into account that RA is predominantly an articular disease, it was interesting to compare the levels of mentioned parameters in synovial fluid (SF) and sera of different RA activity groups with the control group (CG) with non-inflamed synovitis.

## Patients and Methods

In this multicentric study fifty-nine RF positive sera and SF were assayed in RA patients for the level of CIC, C3c and C4 components of the complement and compared with the same biological fluids of the control

CIK. Može se zaključiti da je laboratorijsko određivanje koncentracije CIK korisan pokazatelj stepena aktivnosti RA, što se ne odnosi na ispitivane komponente komplementa. To ne isključuje njihovu aktivnost u okviru efektornog imunskega mehanizma, ali ukazuje na to da bi manji molekulski fragmenti (C3d, C4d) i novi aktivacioni produkti mogli biti bolji pokazatelji stepena aktivnosti RA.

**Ključne reči:** reumatoidni artritis, cirkulišуci imunski kompleksi, komponente komplementa, aktivnost bolesti

group of 15 patients with lesions menisci. All RA patients satisfied the American Rheumatism Association criteria (20) and were classified into two groups: one with moderate ( $n=24$ ) and another with severe ( $n=35$ ) disease activity. The RA activity was classified according to adequate clinical (number of painful and swollen joints, Steinbrocker's radiographic changes, Ritchie articular index, Disease Activity Score) and biochemical inflammatory parameters: erythrocyte sedimentation rate, fibrinogen, C-reactive protein and serum amyloid A concentration.

The levels of CIC and complement components in both fluids were assayed by an immunonephelometric (DADE Behring) method. Prior to testing, all SF samples were pretreated with hyaluronidase (*Streptomyces hyalurolyticus*). Statistical analyses were performed using the Basic Statistic software for the comparison of means (Mann-Whitney U test), standard deviation and distribution data (21).

## Results

Average values of C3c and C4 components in both RA activity groups as well as in the control group were approximately lower by 50 to 70 percent in SF as compared to the sera values (Table I), with the statistical significance of  $p<0.001$ .

In the groups with moderate and severe RA activity, the average values of CIC were more than 80 percent higher in SF, which is also statistically highly significant ( $p<0.001$ ). The comparison of complement components and CIC concentration in the tested biological fluids in two different RA activity groups and the control group is presented in Tables II–IV.

There were no significant differences for the examined complement components in the three tested groups. High statistical significance was only found for the CIC concentrations in both fluids (serum and SF) between moderate and severe activity RA groups. The level of statistical significance in RA patients with MA and SA was higher in SF as compared to sera values. There were no significant differences between MA and CG (Table II, Figure 1).

**Table I** Comparison of C3c, C4 and CIC levels in sera and SF in three tested groups

Control group (N=15)			$\bar{x} \pm SD$ (g/L)	$\bar{x} \pm SD$ (g/L)	Statistical significance	% $\bar{x}$ , C3,C4, CIC in SF vs. serum
Serum	vs.	SF				
C3c	vs.	C3c SF	1.12 ± 0.26	0.28 ± 0.16	p < 0.001	- 75 %
C4	vs.	C4 SF	0.25 ± 0.09	0.07 ± 0.06	p < 0.001	- 72 %
CIC	vs.	CIC SF (IU mL)	1.81 ± 0.76	2.42 ± 0.66	p < 0.05	+ 34 %
Moderate activity N=24			serum	SF		
C3c	vs.	C3c SF	1.22 ± 0.37	0.41 ± 0.28	p < 0.001	- 66 %
C4	vs.	C4 SF	0.25 ± 0.08	0.12 ± 0.18	p < 0.001	- 52 %
CIC	vs.	CIC SF (IU mL)	3.01 ± 2.92	5.33 ± 7.53	p < 0.05	+ 77 % *
Severe activity N=35			serum	SF		
C3c	vs.	C3c SF	1.33 ± 0.36	0.44 ± 0.3	p < 0.001	- 67 %
C4	vs.	C4 SF	0.29 ± 0.13	0.10 ± 0.1	p < 0.001	- 69 %
CIC	vs.	CIC SF (IU mL)	7.43 ± 13.4	13.5 ± 21.1	p < 0.001	+ 82 % *

Abbreviations: CIC – circulating immune complexes; SF – synovial fluid; vs. – versus; N – number of patients;  $\bar{x}$  – arithmetic mean; SD – standard deviation; p – statistical significance, NS – not significant by Mann Whitney U test. \* – p < 0.05 in spite of the adequate value in control group (t-test proportion).

**Table II** Comparison of C3c, C4 levels vs. CIC in serum and SF between groups with moderate activity of RA and control group.

RA activity groups Moderate vs. Control (N = 24) (N = 15)	$\bar{x} \pm SD$ (g/L)	$\bar{x} \pm SD$ (g/L)	Statistical significance
C3c vs. C3c	1.22 ± 0.37	1.12 ± 0.26	NS
C3c SF vs. C3c SF	0.41 ± 0.28	0.28 ± 0.16	NS
C4 vs. C4	0.25 ± 0.08	0.25 ± 0.08	NS
C4 SF vs. C4 SF	0.12 ± 0.18	0.07 ± 0.06	NS
CIC vs. CIC (IU/mL)	3.01 ± 2.92	1.81 ± 0.76	NS
CIC SF vs. CIC SF (IU/mL)	5.33 ± 7.53	2.42 ± 0.66	NS

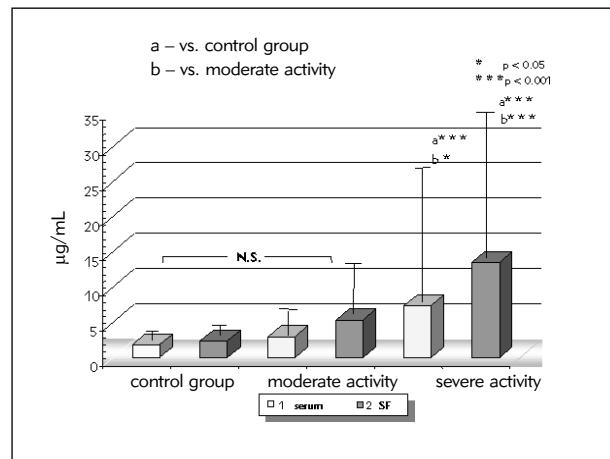
**Table III** Comparison of C3c, C4 levels vs. CIC in serum and SF among groups with moderate and severe activity of RA.

RA activity groups Moderate vs. Severe (N = 24) (N = 15)	$\bar{x} \pm SD$ (g/L)	$\bar{x} \pm SD$ (g/L)	Statistical significance
C3c vs. C3c	1.33 ± 0.40	1.22 ± 0.37	NS
C3c SF vs. C3c SF	0.44 ± 0.30	0.41 ± 0.28	NS
C4 vs. C4	0.29 ± 0.10	0.25 ± 0.08	NS
C4 SF vs. C4 SF	0.10 ± 0.10	0.12 ± 0.18	NS
CIC vs. CIC (IU/ml)	7.43 ± 13.40	3.01 ± 2.92	p < 0.05
CIC SF vs. CIC SF (IU/mL)	13.47 ± 21.1	5.33 ± 7.53	p < 0.001

**Table IV** Comparison of C3c, C4 levels vs. CIC in serum and SF between control group and group with severe activity of RA.

RA activity groups Control vs. Severe (N = 24) (N = 15)	$\bar{x} \pm SD$ (g/L)	$\bar{x} \pm SD$ (g/L)	Statistical significance
C3c vs. C3c	1.33 ± 0.40	1.12 ± 0.26	NS
C3c SF vs. C3c SF	0.44 ± 0.30	0.28 ± 0.16	NS
C4 vs. C4	0.29 ± 0.10	0.25 ± 0.08	NS
C4 SF vs. C4 SF	0.10 ± 0.10	0.07 ± 0.06	NS
CIC vs. CIC (IU/mL)	7.43 ± 13.40	1.81 ± 0.76	p < 0.001
CIC SF vs. CIC SF (IU/mL)	13.47 ± 21.1	2.42 ± 0.66	p < 0.001

Abbreviations: CIC – circulating immune complexes; SF – synovial fluid; vs. – versus; N – number of patients;  $\bar{x}$  – arithmetic mean; SD – standard deviation; p – statistical significance.



**Figure 1** Comparison of CIC values in sera and SF in the control and groups with moderate and severe activity of RA.

## Discussion

Under normal conditions, ICs are rapidly removed from the blood stream by macrophages in the spleen and Kupffer cells in the liver. Large ICs can be easily removed, while small ones do not provoke pathological states. Therefore, ICs of intermediate size usually have the largest potential for developing autoaggression. However, ICs continue to circulate in some circumstances. Eventually, they become trapped in the tissues of the kidneys, lung, skin, joints, or blood vessels. Certain properties influence their potential pathogenicity. Features of particular importance include nature, size, and concentration of the antigen, nature, size, and concentration of the antibody (22), rate of formation and clearance of the ICs (23). Just where they end up they set off reactions that lead to inflammation and tissue damage. ICs develop in autoimmune disease, resulting in the continuous production of autoantibodies which overload the ICs removal system. Up to date, unclear etiopathology of RA involves a large spectrum of biological molecules with several antigens or autoantigens and adequate antibodies, producing high

levels of CIC, which are important for the severity of RA. Higher CIC level in SF than in serum implies its local production, that can be seen from the results obtained in this examination. However, to what extent CIC levels belong to the mentioned types of complexes and complement activation via the classical pathway remains an open question. Complement proteins are a major determinant for the size and solubility of an immune complex, which affects clearance. The evidence regarding intraarticular activation of the complement system in RA could elicit their involvement in pathogenesis and inflammation. The obtained results show no evidence of complement consumption in the sera or SF. Complement activation or consumption through C3 and C4 in the sera is hard to detect because all mentioned molecules belong to the family of acute phase proteins. Their production as a consequence of inflammation possibly masks the C3 and C4 consumption. Some authors for that reason suggest novel activation products (C1q-C4 complex, C3dg), as more relevant for complement activation in RA (24, 25). Our previous results revealed the connection between RA severity and the high level of different RF isotype combinations in both fluids (9), and confirmed the hypothesis about their involvement as a part of ICs in the pathological mechanisms.

More than 40 assay techniques used to detect or measure CIC have been described. Those tests include the Raji cell assay, C1q deviation test, conglutinin test, fluid and solid phase C1q binding procedures, RF assay, PEG precipitin test, etc. Since the size and physicochemical properties of CIC vary markedly, none of these assays has been accepted as a standard. A collaborative study (26) sponsored by the World Health Organization determined that no single method was appropriate in all suspected disease states and recommended that at least two different assay techniques should be performed to detect and measure CIC adequately. Everything mentioned above makes the interpretation of CIC results difficult for clinicians (27, 28). CIC concentration is relevant only in cases with clearly established clinical symptoms of the disease. Our results support the hypothesis (28,

29) that circulating as well as intraarticular ICs together with complement split products may play an important role in some RA pathogenetic aspects. The C1q binding test with anticomplementary activity may be the method of choice for evaluating CIC levels in RA.

The findings presented here imply a possible putative role of the CIC interaction in complement

activation in RA. Quantification of CIC concentrations in sera and in SF showed that it could be a good laboratory parameter as a marker for the RA high activity, especially in combination with IgMRF (9) and anti-CCP Ab (30, 31). Measurement of total C3 and C4 in serum as a routine laboratory method is time-consuming without marked relevant data.

## References

1. Firestein GS. Evolving concepts of rheumatoid arthritis. *Nature* 2003; 423: 356–61.
2. Zubler RH, Nydegger U, Perrin LH, Fehr K, McCormick J, Lambert PH, Miescher PA. Circulating and intra-articular immune complexes in patients with rheumatoid arthritis. Correlation of 125I-C1q binding activity with clinical and biological features of the disease. *J Clin Invest* 1976; 57: 1308–19.
3. Antes U, Heinz HP, Schultz D, Brackertz D, Loos M. C1q-bearing immune complexes detected by a monoclonal antibody to human C1q in rheumatoid arthritis sera and synovial fluids. *Rheumatol Int* 1991; 10: 245–50.
4. Low JM, Moore TL. A role for the complement system in rheumatoid arthritis. *Curr Pharm Des* 2005; 11: 655–70.
5. Newkirk MM, Fournier MJ, Shiroky J. Rheumatoid factor avidity in patients with rheumatoid arthritis: identification of pathogenic RFs which correlate with disease parameters and with the gal(0) glycoform of IgG. *J Clin Immunol* 1995; 15: 250–7.
6. Wolfe F, Sharp J. Radiographic outcome of recent-onset rheumatoid arthritis: a 19-year study of radiographic progression. *Arthritis Rheum* 1998; 41: 1571–82.
7. Bakhari M, Lunt M, Harrison B, Scott D, Symmons D, Silman A. Rheumatoid factor is the major predictor of increasing severity of radiographic erosions in rheumatoid arthritis: results from the Norfolk Arthritis Register Study, a large inception cohort. *Arthritis Rheum* 2002; 46: 906–12.
8. Mijušković VZ, Zgradić I, Nikolajević R, Popović M, Neđović J, Lazarević M, Stojanović J. Rheumatoid factor isotypes as the prognostic markers in Rheumatoid arthritis. Proceedings of the 10<sup>th</sup> International Congress of Immunology 1–6 Nov. 1998; New Delhi, India, Monduzzi Editore Sp. A – Bologna, 1251–5.
9. Mijušković VZ, Zgradić I, Čolić M, Đurović M. Synovial fluid RF isotypes superior to the same sera markers for the course and the disease activity in rheumatoid arthritis. Monduzzi editore: Proceedings of the 15<sup>th</sup> Congress of Clinical Biochemistry and Laboratory Medicine, Barcelona, Spain, June 1–5 2003; pp. 713–6.
10. Girbal-Neuhauser E, Durieux J, Arnaud M, et al. The epitopes targeted by the rheumatoid arthritis-associated antifilaggrin autoantibodies are post-translationally generated on various sites of (pro)filaggrin by deimination of arginine residues. *J Immunol* 1999; 162: 585–94.
11. Menard H, Boire G, Lopez-Longo F, Lapointe S, Larose A. Insights into rheumatoid arthritis derived from the Sa immune system. *Arthritis Res* 2000; 2: 429–32.
12. Vossenaar ER, Despres N, Lapointe E, Van der Heijden A, Lora M, Senshu T, Van Venrooij WJ, Menard HA. Rheumatoid arthritis specific anti-Sa antibodies target citrullinated vimentin. *Arthritis research & therapy* 2004; 6 (2): 142–50.
13. Vittecoq O, Salle V, Jouen-Beades F, et al. Autoantibodies to the 27 C-terminal amino acids of calpastatin are detected in a restricted set of connective tissue diseases and may be useful for diagnosis of rheumatoid arthritis in community cases of very early arthritis. *Rheumatology* 2001; 40: 1126–34.
14. Hassfeld W, Steiner G, Graninger W, Witzmann G, Schweitzer H, Smolen J. Autoantibody to the nuclear antigen RA33: A marker for early rheumatoid arthritis. *Br J Rheumatol* 1993; 32: 199–203.
15. Garner P, Gineys E, Christgau S, Finck B, Delmas P. Association of baseline levels of urinary glucosyl-galactosyl-pyridinoline and type II collagen C-telopeptide with progression of joint destruction in patients with early rheumatoid arthritis. *Arthritis Rheum* 2002; 46: 21–30.
16. Van Venrooij WJ, Zendman AJ, Pruijn GJ. Autoantibodies to citrullinated antigens in early rheumatoid arthritis. *Autoimmunity reviews* 2006; 6 (1): 37–41.
17. Van Venrooij WJ, Zendman AJ. Anti-CCP2 antibodies: An overview and perspective of the diagnostic abilities of this serological marker for early rheumatoid arthritis. *Clinical reviews in allergy & immunology* 2008; 34 (1): 36–9.
18. Van Venrooij WJ, Van Beers JJ, Pruijn GJ. Anti-CCP antibody, a marker for the early detection of rheumatoid arthritis. *Annals of the New York Academy of Science* 2008; 1143: 268–85.
19. Mathsson L, Lampa J, Mullazehi M, Rönnelid J. Immune complexes from rheumatoid arthritis synovial fluid induce FcγRIIa-dependent and rheumatoid factor correlated production of tumour necrosis factor-α by peripheral blood mononuclear cells. *Arthritis Research & Therapy* 2006; 8:R64 <http://arthritis-research.com/content/8/3/R64>

20. Arnett FC, and Committee: The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum* 1988; 31: 315–9.
21. Aslan D, Sandberg S. Simple statistics in diagnostic tests. *Journal of Medical Biochemistry* 2007; 26: 309–13.
22. McDougal JS, McDuffie FC. Immune Complexes in Man: Detection and Clinical Significance. *Adv Clin Chem* 1985; 24: 1–60.
23. Endo L, Corman LC, Panush RS. Clinical Utility of Assays for Circulating Immune Complexes. *Med Clin North Am* 1985; 69 (4): 623–36.
24. Wouters D, Voskuyl EA, Molenaar THE, Dijkmans ACB, Hack CE. Evaluation of classical pathway activation in rheumatoid arthritis: measurement of C1q-C4 complex as novel activation products. *Arthritis care & Research* 2006; 54 (4): 1143–50.
25. Solomon S, Kassahn D, Illges H. The role of the complement system and the FcyR system in the pathogenesis of arthritis. *Arthritis Research & Therapy* 2005; 7: 129–35.
26. Lambert PH, Dixon FJ, Zubler RH, Agnello V, Cambiaso C, Casali P, et al. A WHO collaborative study for the evaluation of eighteen methods for detecting immune complexes in serum. *J Clin Lab Immunol* 1978; 1 (1): 1–15.
27. Wener HM. Immune complexes. In: *Current molecular medicine: Principles of molecular rheumatology 2000*; Ed. G C Tsokos, Humana Press Inc., Totowa, NJ.
28. Low JM, Moore TL. A Role for the Complement System in Rheumatoid Arthritis. *Current Pharmaceutical Design* 2005; 11 (5): 655–70.
29. Okroj M, Heinegard M, Holmdahl R, Blom AM. Rheumatoid arthritis and the complement system. *Annals of medicine* 2007; 39 (7): 517–21.
30. Agrawal S, Misra R, Aggarwal A. Autoantibodies in rheumatoid arthritis: association with severity of disease in established RA. *Clin Rheumatol* 2007; 26 (2): 201–4.
31. Gupta R, Tabah MM, Aneja R, Kumar A, Varghese T, Chandrasenan PJ. Usefulness of anti-CCP antibodies in rheumatic diseases in Indian patients. *Indian J Med Sci* 2009; 63 (3): 92–9.

Received: April 23, 2009

Accepted: June 26, 2009