

THE EFFECT OF HYPERGLYCEMIA ON THE VALUES OF ANTIOXIDATIVE PARAMETERS IN TYPE 2 DIABETIC PATIENTS WITH CARDIOVASCULAR COMPLICATIONS

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Summary: Oxidative stress and hyperglycemia play an important role in the aetiology of vascular complications in diabetes. Hyperglycemia induces a large number of alterations in vascular tissue that potentially accelerate the atherosclerotic process. Nonenzymatic glycosylation of proteins and lipids which can interfere with their normal function by disrupting molecular conformation, alter enzymatic activity, reduce degradative capacity and interfere with receptor recognition, oxidative stress and protein kinase C activations are the major processes that encompass most of the pathological alterations in the diabetic vasculature. Diabetes-associated oxidative stress is a consequence of both increased production of free radicals and reduced capacity for antioxidative defense. The aim of this study was to test the parameters of antioxidative defense: superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), glutathione reductase (GR) and total antioxidant status (TAS), in type 2 diabetic patients with and without cardiovascular complications in order to determine the effect of hyperglycemia on the extent of disorders of antioxidative parameters. Sixty nine type 2 diabetic patients with cardiovascular complications, along with 48 diabetics without complications and 42 healthy subjects were examined. Determination of antioxidant parameters was performed using commercial tests manufactured by Randox Laboratories, UK, based on spectrophotometer determination methods. Statistical data processing revealed significantly lower SOD, GSH-Px, GR and TAS values in type 2 diabetic patients with cardiovascular complications in comparison to the control group ($p < 0.0001$), as well as in comparison to diabetics without complications ($p < 0.01$).

Key words: oxidative stress, antioxidative defense, type 2 diabetic patients, cardiovascular complications

Introduction

Diabetes-associated oxidative stress is a consequence of both increased production of free radicals and reduced capacity for antioxidative defense. Prolonged hyperglycemia is the major factor in the pathogenesis of atherosclerosis in diabetes which is the cause of 80% of total mortality in diabetic patients, while more than 75% of total hospitalizations due to

diabetic complications are attributable to cardiovascular diseases. Hyperglycemia induces a large number of alterations in the vascular tissue that potentially accelerate the atherosclerotic processes. There are three major mechanisms that encompass most of the pathological alterations observed in the diabetic vasculature: 1) nonenzymatic glycosylation of proteins and lipids which can interfere with their normal function by disrupting molecular conformation, alter enzymatic activity, reduce degradative capacity and interfere with receptor recognition; 2) oxidative stress and 3) protein kinase C activation with subsequent alteration in growth factor expression (1, 2).

One of the most important mechanisms responsible for accelerated atherosclerosis in diabetes is the nonenzymatic reaction between glucose and proteins or lipoproteins in arterial walls, leading to for-

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mation of advanced glycosylation end products (AGEs). Once formed, AGE protein adducts are stable and irreversible and continuously accumulate with aging and accelerated rate of diabetes on the long-lived vessel wall proteins. The degree of nonenzymatic glycation is determined mainly by glucose concentration and time of exposure (3). AGEs can accelerate the atherosclerotic process by diverse mechanisms, which can be classified as non-receptor dependent and receptor mediated.

The most studied example of a non-receptor dependent mechanism is the interference of normal physiology of the low-density lipoprotein particle. The glycosylation process occurs both on the apoprotein B and phospholipids components of LDL, leading to functional alteration of LDL clearance and increased susceptibility to oxidative modifications (4–7). LDL-oxidation following AGE-LDL formation occurs in direct proportion to glucose concentration and it is considered a critical step in the atherosclerotic process (8).

The presence of AGE receptor (RAGE) has been demonstrated in all cells relevant to the atherosclerotic process including monocyte-derived macrophages, endothelial cells and smooth muscle cells (9). AGE interaction with RAGE on endothelial cells results in introduction of oxidative stress and consequently of transcription factor NF- κ B and Vascular Cell Adhesion Molecule-1 (VCAM-1) (10) resulting in reduced barrier function with increased permeability of endothelial cell monolayers (11). This reaction can initiate events in atherogenesis as well as monocyte-macrophage interaction of RAGE and AGE resulting in the production of mediators such as interleukin-1 (IL-1), tumor necrosis factor- β (TNF- β), platelet-derived growth factor (PDGF) and insulin growth factor-1 (IGF-1) (12–14). In smooth muscle cells, binding of AGE-modified proteins with RAGE is associated with increased cellular proliferation (15).

High glucose concentration activates protein kinase C (PKC) by increasing the formation of diacylglycerol (DAG) which is the major endogenous cellular co-factor of PKC activation (16). The PKC system is involved in the transcription of several growth factors and in signal transduction as response to growth factors (17,18). PKC activation increases the expression of transforming growth factor- β (TGF- β), which is one of the most important growth factors regulating extracellular matrix production (19). Increased expression of TGF- β is thought to lead to the thickening of capillary basement membrane, one of early structural abnormalities observed in almost all tissues in diabetes.

Hyperglycemia can increase oxidative stress through several pathways. The main mechanism is hyperglycemia-induced intracellular reactive oxygen species, produced by the proton electrochemical gradient generated by mitochondrial electron transport

chain and resulting in increased production of superoxide (2). The other mechanism involves the transition metal-catalysed auto-oxidation of free glucose yielding superoxide anion and hydrogen peroxide (20). The third mechanism involves the transition metal-catalysed auto-oxidation of protein-bound Amadori products, which yields superoxide and hydroxyl radicals and highly reactive dicarbonyl compounds (21).

There is also evidence that hyperglycemia may compromise natural antioxidant defense. Under normal circumstances free radicals are rapidly eliminated by antioxidants such as reduced glutathione, vitamin C and vitamin E (22, 23). Reduced glutathione content, as well as reduced vitamin E, have been reported in diabetic patients (24). Plasma and tissue levels of vitamin C are 40–50% lower in diabetic patients compared with non-diabetic subjects. Non-enzymatic glycation of enzymes, especially intra- and extracellular SOD, compromise their catalytic activity.

The aim of this study was to test the parameters of antioxidative defense: SOD, GSH-Px, GR and TAS in type 2 diabetic patients with and without cardiovascular complications in order to discover the effect of hyperglycemia on the extent of disorders of antioxidative parameters.

Material and Methods

A total of 159 subjects: 71 males and 88 females aged 32 to 90 years, were included in the study. They were divided into three groups: 69 of them, aged 57.9 ± 8.7 years (37 males and 32 females), were type 2 diabetics with cardiovascular complications (DM + CVC), 48 of them were type 2 diabetics without complications (DM) (25 males and 23 females), aged 58.1 ± 10 years, while the control group consisted of 42 age-matched healthy subjects (9 males and 33 females). Type 2 diabetic patients were tested angiographically in order to establish the presence and type of complications. All of type 2 diabetic patients with cardiovascular complications had coronary artery disease (CAD) as primary complication, 32 (46.4%) of them had CAD and hypertension (CAD + HTA), 7 (10.1%) patients had CAD and acute myocardial infarction (CAD + AMI), while 13 (18.8%) had all three types of complications (CAD + HTA + AMI).

Determination of antioxidant parameters: SOD, GSH-Px, GR and TAS, was performed using commercial tests manufactured by Randox Laboratories, UK, based on spectrophotometer determination methods. SOD was determined in a hemolysate prepared from blood collected in lithium-heparinized Vacutainer test-tubes. The hemolysate preparation for SOD determination consisted of erythrocyte separation by centrifugation (10 minutes at 3 000 rpm), fol-

lowed by washing four times with 3 mL of 154 mmol/L NaCl and centrifuged at 3 000 rpm. After the last supernatant decantation, erythrocytes were lysed with 2 mL of cold deionized water and left for 15 minutes at 4 °C in order to finish hemolysis. To obtain a linear measurement, it was necessary to dilute the lysates 26 times with 10 mmol/L of pH 7.0 phosphate buffer. SOD determination was performed using Ransod test kit (Randox Laboratories Ltd, UK) based on the method described by Goldstein (26).

For GSH-Px determination, the whole blood was diluted with dilution solution (obtained through test-reagents) and lysed with a doubly concentrated Drabkin reagent. In this way, blood was diluted 41 times with addition of equal amounts of indicated solution, and then examined using Ransel kits based on the method described by Paglia and Valentine (27).

TAS and GR were determined in plasma obtained after the 10-minute centrifugation of Li-heparinized blood at 3 000 rpm using commercial kits developed by the same manufacturer, based on methods described by Miller (28) and Goldberg (29), respectively.

Fasting glucose levels were also determined in sera for all subjects using a standard method.

For statistical evaluation, basic methods of descriptive statistics were used: mean values with dispersion measure (standard deviation). Statistical significance was determined using Student's t-test, Mann-Whitney U-test and one-way analysis of variance, as well as Spearman rank correlation test.

Results

The obtained values of antioxidative parameters and glucose concentration are presented in *Table I*.

Statistical data processing revealed significantly lower SOD values in diabetics with cardiovascular complications compared to the control group ($p < 0.0001$) and to diabetics without complications ($p < 0.001$), as well as significantly lower GSH-Px values in both pathological groups in comparison to the control group ($p < 0.05$). There was also a significant difference of GSH-Px values between diabetics with and without complications ($p < 0.001$). Mean GR and TAS values of diabetics with complications were also significantly lower in comparison to the control group ($p < 0.0001$ and $p = 0.0002$ respectively). Lower GR and TAS values were also obtained in diabetics without complications, but such difference was not statistically significant.

Diabetics with coronary disease as their complication had significantly lower values of TAS ($t = 2.57$, $p < 0.05$), GR ($t = 1.96$, $p < 0.05$) and glucose ($t = 1.965$, $p < 0.05$) in relation to patients with coronary disease and experienced acute myocardial infarction as complications (*Table II*). Patients with coronary disease and AMI manifested significantly higher values of TAS in relation to DM patients having CD with HTA ($t = 1.965$, $p < 0.05$). Reviewing as a whole the diabetics with complications and interrelating the values of antioxidative parameters, significant positive correlation was found between SOD and GPx, where Spearman's factor was $r = 0.259$ for $p < 0.05$, while there was negative correlation between TAS and GPx and GR, but it was not statistically significant.

In diabetics without complications, Spearman's correlation coefficient revealed significant positive

Table I Demographic and biochemical characteristics of the tested groups: DM + CVC – Type 2 diabetic patients with cardiovascular complications; DM – Type 2 diabetic patients without complications; CG – Control group

Characteristics	DM + CVC	DM	CG
N	69	48	42
Sex (male/female)	37/32	25/23	9/33
Age (years)	57.9 ± 8.7	58.1 ± 10	51.6 ± 11.4
Duration of DM (years)	9.34 ± 9.7	7.36 ± 7.5	–
SOD (U/gHb)	806.5 ± 103.6*	961.0 ± 92.9♠	969.0 ± 104.8
GSH-Px (U/gHb)	23.6 ± 4.6*	27.2 ± 5.3*♠	29.1 ± 3.5
GR (U/L)	55.1 ± 9.5*	59.3 ± 8.8♠	62.5 ± 8.0
TAS (mmol/L)	1.17 ± 0.19*	1.27 ± 0.21♠	1.35 ± 0.23
Glucose (mmol/L)	8.6 ± 3.2*	9.2 ± 3.4*	5.0 ± 0.8
Significance of differences: * $p < 0.05$ – difference between pathological group and control group ♠ $p < 0.05$ – difference between DM and DM + CVC			

Table II Characteristics of type 2 diabetic patients with cardiovascular complications:
 CAD – coronary artery disease; CAD+HTA – coronary artery disease with hypertension;
 CAD+AMI – coronary artery disease and acute myocardial infarction;
 CAD+HTA+AMI – coronary artery disease, hypertension and myocardial infarction

Characteristics	CAD	CAD+HTA	CAD+AMI	CAD+AMI+HTA
N	17	32	7	13
Sex (M/F)	10/7	16/16	3/4	8/5
SOD (U/gHb)	791.6 ± 106.6	799.4 ± 112.7	842.4 ± 53.4	805 ± 119.6
GSH-Px (U/gHb)	23.8 ± 5.3	23.1 ± 4.0	25.9 ± 4.5 ♦	23.1 ± 5.8
GR (U/L)	52.4 ± 10.5*	55 ± 10.3	60.4 ± 8.5	57.8 ± 6.0 ♥
TAS (mmol/L)	1.15 ± 0.15*	1.17 ± 0.19	1.34 ± 0.1 ♦	1.18 ± 0.25
Glucose (mmol/L)	9.1 ± 2.3*	8.9 ± 3.8	7.3 ± 2.1	7.5 ± 1.8 ♥
Significance of differences: * p<0.05 difference between CAD and CAD + AMI ♦ p<0.05 difference between CAD + AMI and CAD + HTA ♥ p<0.05 difference between CAD + AMI +HTA and CAD ♠ p<0.05 difference between CAD + HTA and CAD				

correlation between SOD and glucose ($r = 0.375$, $p < 0.05$), and between GPx and glucose ($r = 0.384$, $p < 0.05$). If the diabetics with complications were observed as a heterogeneous group consisting of 4 subgroups, Spearman's correlation coefficient disclosed important information: significant negative correlation was found between GPx and glucose ($r = -0.382$, $p < 0.05$) in DM group with CD and HTA, and also in DM group with CD and AIM ($r = -0.860$, $p < 0.05$); in DM group with all three types of complications highly significant negative correlation was found between SOD and glucose ($r = -0.590$, $p < 0.05$).

These data suggest a direct correlation between glucose concentrations and the activities of studied enzymes: SOD and GPx, as well as a negative impact of glycosylation of proteins and proteinaceous enzymes involved in this process, which is manifested by reduced catalytic activity of the enzymes.

Discussion

On the basis of the obtained results, it may be concluded that the values of studied antioxidative parameters (SOD, GPx, GR and TAS) were significantly lower in diabetics with cardiovascular complications both in relation to the controls ($p < 0.001$) and to the diabetics without complications ($p < 0.05$) (30, 31). Cardiovascular complications alter the antioxidative defense of diabetic patients in the way that TAS and GR values are significantly lower ($p < 0.05$) in diabetics with coronary disease, regardless of the fact whether it is associated with hypertension or not, in

relation to those diabetics who have experienced acute myocardial infarction in the last 8 years. It is important to highlight that, in the diabetic group without complications, the increase of glucose concentration is followed by higher activity of SOD and GPx, which means that in these patients hyperglycemia induces a positive response from the antioxidative defense system; on the contrary, in diabetics with cardiovascular complications, higher glucose concentration is associated with lower GPx activity in diabetic subgroups who had coronary artery disease with hypertension and AIM, respectively ($p < 0.05$), as well as with lower SOD activity in diabetics with all three forms of complications (CAD+HTA+AIM). Such negative response of the antioxidative defense system may be related to the effect of protein glycosylation and the impact of oxidative stress on reduced catalytic SOD and GPx activity, all contributing to impaired total antioxidative defense of diabetics with cardiovascular complications.

Similar results have been obtained by other authors who had studied this issue.

Oda et al. (30) proved, by *in vitro* experiment, that incubation of Cu,Zn-SOD with increasing glucose concentration ranging from 10–100 mmol/L in a time period of 2–120 hours produces increased glycosylation of this enzyme and reduces its activity by 40%. The same authors confirmed this *in vitro* experiment by an *in vivo* one, where the activity of erythrocytic Cu,Zn-SOD in insulin-independent diabetics correlated negatively with glucose concentration, suggesting that hyperglycemia brings about the glycosylation and inactivity of this enzyme.

Kesavulu et al. (33) obtained low SOD and GPx values in type 2 diabetics with microvascular complications such as retinopathy and nephropathy which were associated with higher concentrations of lipid peroxide and HbA_{1c} (>10%). Siemianowich et al. (34) obtained low values of GPx and catalase in children who were at high risk of coronary arterial disease because they had been born in families with positive history.

Valabhji et al. (35) proved that patients with type 1 DM and coronary arterial calcification had lower TAS value in relation to non-diabetics without calcification, which correlated negatively with diabetes duration, age, degree of calcification, cholesterol and creatinine concentrations, and arterial pressure level. Therefore, they suggest that this parameter be accepted as an independent predictive index of coronary arterial calcification.

Lapenna and assoc. (36) confirmed low GPx and GR values in carotid arterial plaque in 13 patients with marked atherosclerosis of carotid arteries. It is known that GPx and GR may be inactivated by free radicals, particularly hypochloric acid which results from myeloperoxidase action. GPx may also be inactivated by 4-hydroxynonenal, byproduct of lipid peroxidation, which originates from peroxidation of LDL in circulation and inactivates GPx in erythrocytes (36). Therefore, it is believed that these two enzymes are significant in conditions of oxidative stress in athero-

sclerotic lesions. GPx is an essential enzyme for the elimination of organic and inorganic peroxides, and it is a crucial intracellular antioxidative enzyme in mammals (38). Peroxides are cytotoxic to vascular cells, especially in the presence of transitory redox-active metals which are available in catalytic active form in human atherosclerotic plaque (38). The key role of GPx in vascular antioxidative defense is based on the fact that catalase is deficient while SOD is poorly effective in vascular cells (40). Accordingly, deficiency of GPx and redox glutathione cycle in atherosclerotic tissue may considerably weaken their antioxidative potential and therefore favor the pro-oxidative and atherosclerotic processes, even if there is a normal concentration of low-molecular »scavenging« antioxidants (41).

Determination of markers of antioxidative defense as very sensitive parameters not only contributes to a better understanding of the effect of oxidative stress on the development of diabetes and diabetic complications, but also opens new perspectives for the treatment of diabetic complications; in addition, it is particularly important in the prevention of atherosclerosis and diabetic micro- and macrovascular complications.

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UTICAJ HIPERGLIKEMIJE NA VREDNOST ANTIOKSIDATIVNIH PARAMETARA KOD DIJABETIČARA TIPA 2 SA KARDIOVASKULARNIM KOMPLIKACIJAMA

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Kratak sadržaj: Oksidativni stres i hiperglikemija igraju značajnu ulogu u etiologiji vaskularnih komplikacija u dijabetesu. Hiperglikemija indukuje veliki broj oštećenja vaskularnog tkiva koja potencijalno ubrzavaju aterosklerotske procese. Neenzimatska glikozilacija proteina i lipida koja interferira sa njihovom normalnom funkcijom tako što oštećuje molekularnu konformaciju, remeti enzimsku aktivnost, smanjuje kapacitet razgradnje i interferira sa prepoznavanjem proteinskih struktura od strane receptora, zatim oksidativni stres i aktivacija protein kinaze C predstavljaju glavne procese koji su odgovorni za većinu patoloških oštećenja vaskulature kod dijabetičara. Oksidativni stres koji se sreće kod dijabetičara je posledica kako povećane produkcije slobodnih radikala, tako i smanjenog kapaciteta antioksidativne zaštite. Cilj ovoga rada je bio da se ispituju parametri antioksidativne zaštite: superoksid dizmutaza (SOD), glutation peroksidaza (GSH-Px), glutation reduktaza (GR) i totalni antioksidantni status (TAS) kod dijabetičara tipa 2 sa i bez kardiovaskularnih komplikacija kako bi se utvrdio uticaj hiperglikemije na stepen poremećaja antioksidativne zaštite. Ispitano je 69 dijabetičara tipa 2 sa kardiovaskularnim komplikacijama, 48 dijabetičara tipa 2 bez komplikacija i 42 zdrava ispitanika. Određivanje parametara antioksidativne zaštite vršeno je komercijalnim testovima firme Randox Laboratories Ltd, UK, i zasnovano je na spektrofotometrijskim metodama određivanja. Statističkom obradom podataka zaključeno je da su vrednosti SOD, GSH-Px, GR i TAS-a značajno niže u dijabetičara tipa 2 sa prisutnim kardiovaskularnim komplikacijama, kako u odnosu na kontrolnu grupu zdravih ($p < 0,0001$), tako i u odnosu na dijabetičare bez komplikacija ($p < 0,01$).

Ključne reči: oksidativni stres, antioksidativna zaštita, dijabetes mellitus tipa 2, kardiovaskularne komplikacije

References

1. Aronson D, Rayfield EJ. How hyperglycemia promotes atherosclerosis: molecular mechanisms. *Cardiovascular Diabetology* 2002; 1: 1.
2. Nishikawa T, Edelstein D, Du XL, Yamagishi S, Matsumura T, Kaneda Y, et al. Normalizing mitochondrial superoxide production blocks three pathways of hyperglycemic damage. *Nature* 2000; 404: 787–90.
3. Brownlee M, Cerami A, Vlassara H. Advanced glycosylation end products in tissue and the biochemical basis of diabetic complications. *N Engl J Med* 1998; 318: 1315–21.
4. Bucala R, Mitchell R, Arnold K, Innerarity T, Vlassara H, Cerami A. Identification of the major site of apolipoprotein B modification by advanced glycosylation end products blocking uptake by the low density lipoprotein receptor. *J Biol Chem* 1995; 270: 10828–32.
5. Bucala R, Makita Z, Koschinsky T, Cerami A, Vlassara H. Lipid advanced glycosylation: pathway for lipid oxidation *in vivo*. *Proc Natl Acad Sci USA* 1993; 90: 6434–38.
6. Stocker R, Keaney JF Jr. Role of oxidative modifications in atherosclerosis. *Physiol Rev* 2004; 84: 1381–478.
7. Touyz RM. Reactive oxygen species, vascular oxidative stress and redox signaling in hypertension: what is the clinical significance? *Hypertension* 2004; 44: 248–52.
8. Bowie A, Owens D, Collins P, Johnson A, Tomkin GH. Glycosylated low density lipoprotein is more sensitive to oxidation: implications for the diabetic patients? *Atherosclerosis* 1993; 102: 63–7.
9. Schmidt AM, Hori O, Brett J, Yan SD, Wantier JL, Stern D. Cellular receptors for advanced glycation end products. Implications for induction of oxidant stress and cellular dysfunction in the pathogenesis of vascular lesions. *Atheroscler Thromb* 1994; 14: 1521–8.
10. Schmidt AM, Hori O, Chen JX, Li JF, Zhang J, Cao R et al. Advanced glycation end products interacting with their endothelial receptor induce expression of vascular cell adhesion molecule-1 (VCAM-1) in cultured human endothelial cells and in mice. A potential mechanism for accelerated vasculopathy of diabetes. *J Clin Invest* 1995; 96: 1395–1403.
11. Wantier JL, Zoukourian C, Chappay O, Wantier MP, Guillausseau PJ, Cao R, et al. Receptor-mediated endothelial cell dysfunction in diabetic vasculopathy. Soluble receptor for advanced glycation end products blocks hyperpermeability in diabetic rats. *J Clin Invest* 1996; 97: 238–43.
12. Kirstein M, Brett J, Radoff S, Ogawa S, Stern D, Vlassara H. Advanced protein glycosylation induces transendothelial human monocyte chemotaxis and secretion of platelet-derived growth factor: role in vascular disease of diabetes and aging. *Proc Natl Acad Sci USA* 1990; 87: 9010–14.
13. Vlassara H, Brownlee M, Monogue KR, Dinarello CA, Pasagian A. Cachectin/TNF and IL-1 induced by glucose-modified proteins: role in normal tissue remodeling. *Science* 1998; 240: 1546–8.
14. Kirstein M, Aston C, Hints R, Vlassara H. Receptor-specific induction of insulin-like growth factor 1 in human monocytes by advanced glycosylation end product-modified proteins. *J Clin Invest* 1992; 90: 439–46.
15. Vlassara H, Bucala R, Striker L. Pathogenic effects of advanced glycosylation: biochemical biologic and clinical implications for diabetes and aging. *Lab Invest* 1994; 70: 138–51.
16. Xia P, Inoguchi T, Kern TS, Engerman RL, Oates PJ, Kuning GL. Characterization of the mechanism for the chronic activation of diacylglycerol-protein kinase C pathway in diabetes and hyperglycemia. *Diabetes* 1994; 43: 1122–9.
17. Koya D, Haneda M, Nakagawa H, Isshiki K, Soto H, Maeda S et al. Amelioration of accelerated diabetic mesangial expansion by treatment with a PKC beta inhibitor in diabetic DB/db mice, a rodent model for type 2 diabetes. *FASEB J* 2000; 14: 439–47.
18. Park JY, Takohara N, Gabriele A, Chou E, Naruse K, Suzuma K et al. Induction of endothelin expression by glucose: an effect of protein kinase C activation. *Diabetes* 2000; 49: 1239–48.
19. Nabel E, Shum L, Pompili V, Yang ZY, San H, Liptay S et al. Direct transfer of transforming growth factor b1 gene into arteries stimulates fibrocellular hyperplasia. *Proc Natl Acad Sci USA* 1993; 90: 10759–63.
20. Wolf SP. Diabetes mellitus and free radicals. Free radical transition metals and oxidative stress in the aetiology of diabetes mellitus and complications. *Br Med Bull* 1993; 49: 642–52.
21. Baynes JW, Thorpe SR. Role of oxidative stress in diabetic complications: a new prospective on an old paradigm. *Diabetes* 1999; 48:1–9.
22. Yoshida K, Hirokawa J, Tagami S, Kawakami Y, Urata Y, Kondo T. Weakened cellular scavenging activity against oxidative stress in diabetes mellitus: regulation of glutathione synthesis and efflux. *Diabetologia* 1995; 38: 201–10.
23. Dominguez C, Ruiz E, Gussinye M, Carracosa A. Oxidative stress at onset and in early stages of type 1 diabetes in children and adolescents. *Diabetes Care* 1998; 21: 1736–42.
24. Chen MS, Hutchinson ML, Pecoraro RE, Lee WY, Labbe RF. Hyperglycemia-induced intracellular depletion of ascorbic acid in human mononuclear leukocytes. *Diabetes* 1983; 32: 1078–81.
25. Yue DK, McLennan S, Fisher E, Hefferman S, Capogreco C, Ross GR et al. Ascorbic acid metabolism and polyol pathway in diabetes. *Diabetes* 1989; 38: 257–61.

26. Goldstein S, Michel C, Boors A, Saran M, Czapsky G. A critical re-evaluation of some assay methods for superoxide dismutase activity. *Free Radical Biol Med* 1988; 4: 295–303.
27. Paglia DE, Valentine WN. Studies on the quantitative and qualitative characterisation of glutathione peroxidase. *J Lab Clin Med* 1967; 70: 158–63.
28. Miller NJ, Rice-Evans C, Davies MJ, Gopinathan V, Milner A. A novel method for measuring antioxidant capacity and its application to monitoring the antioxidant status in premature neonates. *Clin Sci* 1993; 84: 407–12.
29. Goldberg DM, Spooner RJ. In Bergmeyer HU, editor. *Methods of enzymatic analysis*. 3rd ed. New York: Academic Press 1974, Vol 3, p. 258–65.
30. Čolak E, Majkić-Singh N, Stanković S, Srećković-Dimitrijević V, Đorđević PB, Lalić K et al. Parameters of antioxidative defense in type 2 diabetic patients with cardiovascular complications. *Ann Med* 2005; 37: 613–20.
31. Dimitrijević-Srećković SV, Đorđević PB, Popović SS, Gostiljac D, Canović F, Srećković MB et al. Metabolic syndrome and antioxidative activity in early glycoregulation disorders. *Diabetologia* 2004; 47 (Suppl 1): A143 (Abstract book of the 40th Annual Meeting of the EASD, Munich, Germany, 5–9 September 2004).
32. Oda A, Bnnai C, Yamaoka T, Katori T, Matsushima T, Yamashita K. Inactivation of Cu,Zn-SOD by *in vitro* glycosylation and in erythrocytes of diabetic patients. *Horn metab res* 1994; 26 (1): 1–4.
33. Kesavulu MM, Giri R, Kameswara RB, Apparo Ch. Lipid peroxidation and antioxidant levels in type 2 diabetics with microvascular complications. *Diabetes & Metabolism* 2000; 25 (5): 387–95.
34. Siemianowich K, Gminski J, Francuz T, Wojcik A, Polesiezna B. Activity of antioxidant enzymes in children from families at high risk of premature coronary heart disease. *Scand J Clin Lab Invest* 2003; 63 (2): 151–8.
35. Valabhji J, McColl AJ, Richmond W, Schacher M, Rubens MB, Elkeles RS. Total antioxidant status and coronary artery calcification in type 1 diabetes. *Diabetes Care* 2001; 24: 1608–13.
36. Lapenna D, De Gioia S, Ciofani G, Mezzetti A, Ucchino S, Calafiore AM et al. Glutathione-related antioxidant defenses in human atherosclerotic plaques. *Circulation* 1998; 97: 1930–34.
37. Kinter M, Roberts RJ. Glutathione consumption and glutathione peroxidase inactivation in fibroblast cell lines by 4-hydroxy-2-nonenal. *Free Radic Biol Med* 1996; 21: 457–62.
38. Blankenberg S, Rupprecht HJ, Bickel C, Torzewski M, Hafner G, Tiret L et al. Glutathione peroxidase 1 activity and cardiovascular events in patients with coronary artery disease. *N Engl J Med* 2003; 349 (17): 1605–13.
39. Forgione MA, Cap A, Liao R, Moldovan NI, Eberhardt RT, Lim CC et al. Heterozygous cellular glutathione peroxidase deficiency in the mouse: abnormalities in vascular and cardiac function and structure. *Circulation* 2002; 106 (9): 1154–8.
40. Raes M, Michiels C, Remacle J. Comparative study of the enzymatic defense system against oxygen-derived free radicals: the key role of glutathione peroxidase. *Free Radic Biol Med* 1987; 3: 3–7.
41. Suarna C, Dean RT, May J, Stocker R. Human atherosclerotic plaques contain both oxidized lipids and relatively large amounts of α -tocopherol and ascorbate. *Arterioscler Thromb Vasc Biol* 1995; 15: 1616–24.

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