UC 577,1;61

Jugoslov Med Biohem 24: 265-269, 2005

ISSN 0354-3447

Originalni naučni rad Original paper

EFFECTS OF PHENYLETHYLAMINE ON ISOLATED UTERUS IN RATS TREATED WITH ESTROGEN

Mirko Rosić, Gvozden Rosić, Zorica Lazić, Suzana Pantović, Rajko Lazarević

Medical Faculty, University of Kragujevac, Serbia and Montenegro

Summary: It is well known that histamine produces relaxation of precontracted rat uterus in oestrogenized animals. This effect of histamine was caused by the stimulation of H₂ histamine receptors. Our results confirm these findings (histamine relaxation effect is $EC_{50} = 40.52 \pm 1.1 \times 10^{-6} \text{ mol/L}$). Phenylethylamine also produces relaxation of precontracted rat uterus in oestrogenized animals ($EC_{50} = 40.98 \pm 1.1 \times 10^{-6} \text{ mol/L}$). Phenylethylamine receptors antagonist. Actually, H₂ blockade has shifted concentration-dependent phenylethylamine curve to the right, i.e. produced inhibition of phenylethylamine induced relaxation ($EC_{50} = 61.42 \pm 1.2 \times 10^{-6} \text{ mol/L}$, p < 0.001). Since phenylethylamine is described as a potent inhibitor of histamine N-methyl-transferase (HMT), it is possible that modulation of histamine catabolism may lead to H₂ mediated relaxation of precontracted rat uterus in oestrogenized animals.

Key words: histamine, phenylethylamine, uterus, estrogen

Introduction

Histamine effects on the smooth muscle of a rat uterus are already known. Namely, histamine produces relaxation of precontracted rat uterus (1). Those effects are mediated via H_1 and H_2 histamine receptors (2). However, ovarian steroid hormones have a significant influence on applied histamine effects on precontracted rat uterus, i.e. H_1 relaxant response to histamine was present only in progesterone dominant rat uteri (2).

Also, it has been reported that ovarian hormones treatment can increase the number of rat uterine mast cells (3). Nevertheless, there is still no reliable evidence of the influence of ovarian steroid hormones on histamine catabolism in the rat uterus. Special interest can be focused on histamine N-methyl-transferase (HMT), enzyme responsible for rapid inactivation of histamine by methylation of ring tele-nitrogen

Mirko Rosić, MD, PhD Medical Faculty, University of Kragujevac Serbia and Montenegro Svetozara Markovića 69 34 000 Kragujevac, Serbia and Monteneg

34 000 Kragujevac, Serbia and Montenegro e-mail: rosic@infosky.net

in histamine (4-6). Furthermore, there is strong evidence that at least a few of histamine methylation products show higher selectivity to different classes of histamine receptors in various tissues, but at the same time have rather week biological activity (7, 8).

The aim of this study was to evaluate possible effects of phenylethylamine, a potent inhibitor of HMT, on the isolated uterus motility in rats pretreated with estrogen.

Materials and Methods

Preparation of isolated rat uterus horns

Young female albino rats of the Wistar strain, weighing between 250 g and 300 g, pretreated with diethylstilbestrol (5 mg/kg i.p., 24 hours before sacrifice), were used in this study. Rats were killed by cervical dislocation (according to Schedule 1 of the Animals, Scientific procedures, Act 1986, UK) and exsanguinated. Each experiment was conducted on isolated preparations from five different animals. After laparatomy, both uterine horns were rapidly removed, cut along the longitudinal axis and placed in organ bath.

Address for correspondence:

Experimental design

Each isolated preparation was mounted in the 10 mL organ bath with constant flow (5 mL/min) of De Jalon's solution (NaCl 154 mmol/L, KCl 5.6 mmol/L, CaCl₂ 0.4 mmol/L, KH₂PO₄ 1.18 mmol/L, NaHCO₃ 5.95 and glucose 2.5 mmol/L) maintained at 31 ± 1 °C to avoid spontaneous contractions. The bath was aerated continuously with 95% O₂ and 5% CO₂. One end of the isolated uterine horn was fixed to the organ bath, and the other was fixed to a force-displacement transducer (IT-1 sensor, EMKA Technologies) coupled with tension amplifier and chart recorder.

All preparations were loaded with 1 g weight and allowed to equilibrate 30 minutes. At the end of the equilibration period a submaximal plateau contraction of preparation was obtained by adding KCl (60 mmol/L).

The first set of experiments consisted of recording the uterine horn preparations (precontracted with KCl) responses to histamine (1, 2.6, 26, 260 \times 10^{-6} and 2.6×10^{-3} mol/L, 2 minute each concentration) in animals pretreated with diethylstilbestrol, as well as uterine response to same doses of histamine in the presence of S(+)-chlorpheniramine (permanent perfusion for 5 minutes before agonist use and during agonist's action, with final concentration of 5 \times 10⁻⁷ mol/L) or ranitidine (permanent perfusion for 5 minutes before agonist use and during agonist's action, with final concentration of 1×10^{-6} mol/L). The second set of experiments consisted of recording the uterine horn preparations (precontracted with KCI) responses to phenylethylamine (13, 26, 52, 80 and 130 \times 10⁻⁶ mol/L, 2 minute each concentration) in animals pretreated with diethylstilbestrol, as well as uterine response to same doses of phenylethylamine in the presence of S(+)-chlorpheniramine (permanent perfusion for 5 minutes before agonist use and during agonist's action, with final concentration of 5 \times 10⁻⁷ mol/L) or ranitidine (permanent perfusion for 5 minutes before agonist use and during agonist's action, with final concentration of 1×10^{-6} mol/L). Next concentration of histamine or phenylethylamine on the same preparation was applied only after a period of 15 min. All drugs were applied to organ bath using micro infusion pump with constant flow of $125 \,\mu$ L/min.

Relaxant responses were measured as changes in isometric tension and converted into a percentage of the reference maximum relaxations for each group of experiments.

Chemicals

Drugs used in these experiments were histamine dihydrochloride, ranitidine hydrochloride, diethylstilbestrol, (Sigma Chemical Co, USA), S(+)-chlorpheniramine maleate (RBI, USA), phenylethylamine (Calbiochem, GB) and KCI (Zorka Šabac, Serbia). The drugs were prepared on the day of experiment in NaCl 154 mmol/L (Zorka Šabac, Serbia). Concentrations reported are expressed as final concentrations within the organ bath.

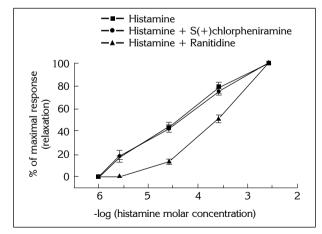
Statistical analysis

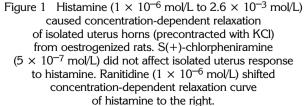
Each concentration was assayed on isolated preparatus from five different animals. Concentration-response curves were constructed using linear regression according to least-squares analysis (9, 10). Effective concentration of agonists that produced 50% of maximal response and response duration (EC₅₀) was calculated for each agonist together with its confidence limits (1.96 × standard error). The Student *t* test was used for the comparison of maximal responses (expressed as mean \pm SEM) for groups 1 and 2. The results were considered statistically significant when $p \le 0.05$.

Results

The effects of histamine on the isolated uterus horns from oestrogenised rats

Histamine (1 × 10⁻⁶ mol/L to 2.6 × 10⁻³ mol/L) produced concentration-dependent relaxation of isolated uterus horns (precontracted with KCl) from oestrogenized rats (EC₅₀ = 40.52 ± 1.1 × 10⁻⁶ mol/L, p < 0.001). S(+)-chlorpheniramine (5 × 10⁻⁷ mol/L), H₁ receptors antagonist, did not affect isolated uterus response to histamine (EC₅₀ = 39.41 ± 1.3 × 10⁻⁶ mol/L). Ranitidine (1 × 10⁻⁶ mol/L), H₂ receptors antagonist, shifted concentration-dependent relaxation curve of histamine to the right (EC₅₀ = 130.41 ± 1.5 × 10⁻⁶ mol/L, p < 0.001) (*Figure 1*).





The effects of phenylethylamine on the isolated uterus horns from oestrogenised rats

Phenylethylamine (13×10^{-6} mol/L to 130×10^{-6} mol/L) produced concentration-dependent relaxation of isolated uterus horns (precontracted with KCl) from oestrogenized rats (EC₅₀ = 40.98 ± 1.1×10^{-6} mol/L, p < 0.001). S(+)-chlorpheniramine (5×10^{-7} mol/L), H₁ receptors antagonist, did not affect isolated uterus response to phenylethylamine in oestrogenized rats (EC₅₀ = 38.89 ± 1.1×10^{-6} mol/L). On the other hand, ranitidine (1×10^{-6} mol/L), H₂ receptors antagonist, has shifted concentration-dependent relaxation curve of phenylethylamine to the right (EC₅₀ = $61.42 \pm 1.2 \times 10^{-6}$ mol/L, p < 0.001) (*Figure 2*).

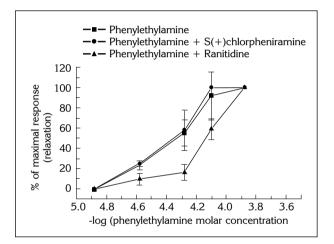


Figure 2 Phenylethylamine $(13 \times 10^{-6} \text{ mol/L} \text{ to } 130 \times 10^{-6} \text{ mol/L})$ produced concentrationdependent relaxation of isolated uterus horns (precontracted with KCl) from oestrogenized rats. S(+)-chlorpheniramine (5 × 10⁻⁷ mol/L) did not affect isolated uterus response to phenylethylamine in oestrogenized rats. Ranitidine (1 × 10⁻⁶ mol/L) shifted concentration-dependent relaxation curve of phenylethylamine to the right.

In the presence of the highest concentration of phenylethylamine, oestrogenised isolated rat uterus horns showed maximal relaxation (16.83 \pm 6.12% of KCl-induced contraction) (*Figure 3*).

The maximal relaxation ($60.57 \pm 3.55\%$ of KClinduced contraction) in this experimental group was reached with the highest concentration of histamine (*Figure 3*).

Discussion

Our results show that histamine induced relaxation of isolated precontracted rat uterus in animals treated with estrogen. The relaxation effect of histamine is concentration-dependent (EC₅₀ = 40.52 \pm 1.1 × 10⁻⁶ mol/L).

In animals treated with estrogen, histamine induced relaxation is mediated only via H₂ histamine receptors because H₁ antagonist failed to prevent this effect (EC₅₀ = 40.52 ± 1.1 × 10⁻⁶ mol/L vs. EC₅₀ = 39.41 ± 1.3 × 10⁻⁶ mol/L with H₁ blockade, p > 0.05). The histamine-induced relaxation of rat uterus in animals treated with estrogen is prevented by H₂ histamine antagonist (EC₅₀ = 40.52 ± 1.1 × 10⁻⁶ mol/L vs. EC₅₀ = 130.41 ± 1.5 × 10⁻⁶ mol/L with H₂ blockade, p < 0.001).

Effects of histamine on isolated rat uterus (precontracted with KCl) have already been described (1, 2). Our results indicate that histamine produces relaxation of KCl-precontracted isolated rat uterus via stimulation of H₂ histamine receptors. This is in accordance with similar published results (2, 8) supporting the conclusion that histamine produces H₂ relaxation of KCl-precontracted uterus in estrogen pretreated animals.

Phenylethylamine produced concentration-dependent relaxation of isolated rat uterus (precontracted with KCl) in oestrogenized rats (EC₅₀ = 40.98 ± 1.1 × 10⁻⁶ mol/L, *p* < 0.001). In addition, it is clear that H₂ blockade has shifted concentration-dependent phenyl ethylamine curve to the right, i.e. produced inhibition of phenylethylamine-induced relaxation (EC₅₀ = 61.42 ± 1.2×10^{-6} mol/L, *p* < 0.001).

The relaxation effect of phenylethylamine is weak comparing to the effects of exogenous histamine (*Figure 3*).

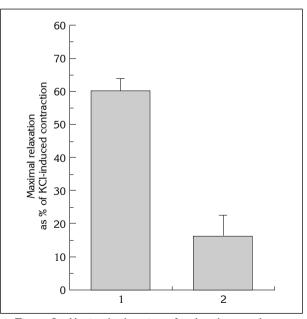


Figure 3 Maximal relaxation of isolated uterus horns (KCI-induced contraction). 1 – histamine in estrogen pretreated rats and 2 – phenylethylamine in estrogen pretreated rats.

Phenylethylamine-induced relaxation of oestrogenized rat's uterus could be the consequence of direct stimulation of H₂ histamine receptors. Phenylethylamine is described in literature as a potent inhibitor of HMT, i.e. it affects histamine metabolic turnover in various tissues by methylation of ring telenitrogen in histamine (4-6). It is also known that some of histamine methylation products show higher selectivity in different classes of histamine receptors in various tissues, but at the same time, they have rather weak biological activity (7, 8). Our results suggest that blockade of methylation (degradation) of endogenous histamine molecule in rat uterus could lead to H₂ mediated relaxation in rats treated with estrogen. It is also known that tissue histamine content and mast cell density are greater during diestrus and in mice treated with progesterone (3). Further investigations have to clarify whether the H2 relaxation of precontracted rat uterus is the consequence of direct phenylethylamine binding to these receptors, or it is a consequence of HMT inhibition, and consequent alteration of endogenous histamine catabolism. Since there is no evidence in literature for direct phenylethylamine binding to histamine receptors, though there is strong evidence for phenylethylamine inhibition of HMT in various tissues, it seems reasonable at the moment, to accept the second hypothesis. If it is true, then the modulation of rapid turnover pool of histamine in uterus leads to a variation of biological effects of this endogenous amine. In course of this idea, the alteration of histamine metabolism could lead to various mechanisms of its actions with different consequences. So, not only does the concentration of histamine itself affect the tissue, but also variation in histamine metabolic pathways could lead to different (sometimes opposite) final effects via stimulation of histamine receptors pool.

In the end, it is very important to emphasize that we performed our experimental protocol with continuous perfusion of preparation in organ bath. This allowed us to avoid accumulation of products of metabolic degradation, because the tissue was continuously washed out. Administration of drugs in the organ bath was performed by means of micro infusion pump (125 μ L/min) – small volume that cannot affect environmental conditions in the organ bath (10 mL volume, and perfusion rate of 5 mL/min). Previous experiments were predominantly performed in classic organ bath without constant washing-out of tissue, where the accumulation of degradation product could 'mask' some effects.

EFEKTI FENILETILAMINA NA ESTROGENOM TRETIRANI IZOLOVANI UTERUS PACOVA

Mirko Rosić, Gvozden Rosić, Zorica Lazić, Suzana Pantović, Rajko Lazarević

Medicinski fakultet, Univerzitet u Kragujevcu, Srbija i Crna Gora

Kratak sadržaj: Dobro je poznato da histamin dovodi do relaksacije prekontrahovanog uterusa u životinja tretiranih estrogenom. Ovaj efekat histamina nastaje kao posledica stimulacije H₂ histaminskih receptora. Naši rezultati potvrđuju ova saznanja (u prisustvu histamina efekat relaksacije je imao vrednost $EC_{50} = 40,52 \pm 1,1 \times 10^{-6}$ mol/L). Takođe, feniletilamin izaziva relaksaciju prekontrahovanog uterusa kod pacova koji su bili tretirani estrogenom ($EC_{50} = 40,98 \pm 1,1 \times 10^{-6}$ mol/L, p < 0,001). Ranitidin, selektivni antagonist H₂ histaminskog receptora, umanjuje ovaj efekat feniletilamina. Blokada H₂ histaminskih receptora ranitidinom je tako pomerila udesno koncentraciono-zavisnu krivu feniletilamina tj, dovela je do inhibicije feniletilaminom izazvane relaksacije ($EC_{50} = 61,42 \pm 1,2 \times 10^{-6}$ mol/L, p < 0,001). S tim u vezi, feniletilamin se opisuje kao potentni inhibitor histamin-metil-transferaze (HMT) tako da modulacija katabolizma histamina možda dovodi do relaksacije prekontrahovanog uterusa pacova, tretiranog estrogenom, preko histaminskih H₂ receptora.

Ključne reči: histamin, feniletilamin, uterus, estrogen

References

- Black JW, Duncan WAM, Durant CJ, Ganellin CR, Parsons ME. Definition and antagonism of histamine H₂ receptors. Nature 1972; 236: 385–90.
- Rubio E, Estan L, Morales-Olivas FJ, Martinez-Mir I. Influence of hormonal treatment on the response of the rat isolated uterus to histamine and histamine receptor agonists. Eur J Pharmacol 1992; 212: 31–6.
- Aydin Y, Tuncel N, Gurer F, Tuncel M, Kosar M, Oflaz G. Ovarian, uterine and brain mast cells in female rats: cyclic changes and contribution to tissue histamine. Comp Biochem Physiol A Mol Integr Physiol 1998; 120: 255–62.
- Ohrui T, Yamauchi K, Sekizawa K, Ohkawara Y, Maeyama K, Sasaki M, Takemura M, Wada H, Watanabe T,

Takishima T. Histamine N-methyl-transferase controls the contractile response of guinea pig trachea to histamine. J Pharmacol Exp Ther 1992; 261: 1268–72.

- Barth H, Lorenz W, Niemeyer I. Inhibition and activation of histamine methyltransferase by methylated histamines. Hoppe Seylers J Physiol Chem 1973; 354: 1024–5.
- Tachibana T, Taniguchi S, Fujiwara M, Imamura S. Regulation of the activity of histamine N-methyltransferase from guinea pig skin by biogenic amines. Exp Mol Path 1986; 45: 257–69.
- 7. Rosić M, Anđelković I, Collis C, Segal M, Stojadinović D, Rosić G. The effects of histamine and histamine

derivatives on the isolated guinea-pig heart. Yugoslav Physiol Pharmacol Acta 1992; 28 (3): 211–14.

- Rubio E, Navarro-Badenes J, Palop V, Morales-Olivas FJ, Martinez-Mir I. The effects of histamine on the isolated mouse uterus. J Auton Pharmacol 1999; 19 (5): 281–9.
- Tallarida JR, Murray RB. Manual of pharmacological calculations with computer programs. 2nd ed. New York: Springer-Verlag, 1987: 297.
- Kenakin RT. The classification of drugs and drug receptors in isolated tissues. Pharmacol Rev 1984; 36: 165–222.

Received: December 20, 2004 Accepted: June 18, 2005