## UC 577,1;61

Jugoslov Med Biohem 22: 207-211, 2003

## SALIVARY CARCINOEMBRYONIC ANTIGEN AS AN INFLAMMATORY MARKER

Snežana Golubović<sup>1</sup>, Ljiljana Janković<sup>2</sup>, Aris Movsesijan<sup>2</sup>, Žanka Bojić<sup>1</sup>, Miroslava Janković<sup>1</sup>

<sup>1</sup>Institute for the Application of Nuclear Energy–INEP, Banatska 31b, 11080, Zemun–Belgrade, Yugoslavia <sup>2</sup>Department of Paradontology and Oral Medicine, University School of Stomatology, Belgrade, Yugoslavia

Summary: In this study we examined concentration of carcinoembryonic antigen (CEA) in paired saliva and serum samples from healthy individuals and patients with periodontal disease. CEA concentration was determined immunoradiometrically using highly specific anti-CEA antibodies. The salivas from periodontally healthy subjects revealed CEA concentrations with median value of 62 mg/L. Distribution of salivary CEA concentrations in patients with periodontal diseases were very broad with median values:  $74 \mu g/L$  (stage I), 84 mg/L (stage II),  $240 \mu g/L$  (stage III) and  $412 \mu g/L$  (necrotizing ulcerative periodontitis-NUP). Analysis of the obtained values indicated statistically significant increase in salivary CEA, in subjects with periodontal diseases. Metronisadole treatment in patients with NUP leads to statistically significant decrease in salivary CEA. The results obtained suggested salivary CEA as a potential marker of the alterations of periodontium.

Key words: CEA, saliva, serum, periodontal disease, protein, immunoradiometric assay.

#### Introduction

Carcinoembryonic antigen (CEA) was first described in 1965 by Gold and Freedman as an oncofoetal antigen present in colonic tumours and foetal gut (1). So far, a number of CEA-related genes as well as splice variants of individual genes have been identified (2 4). CEA gene family is divided into two main subgroups: CEACAM genes, coding for CEA-related cell adhesion molecules and PSG genes, coding for pregnancy-specific glycoproteins (5). The CEA subgroup members are cell surface associated glycoproteins, showing a complex expression pattern in normal and neoplastic tissues (6, 7). Thus, CEA being a wellknown colorectal tumour marker was also detected in different normal adult tissues (8).

Immunochemical techniques and RNA blot analyses revealed the presence of CEA and CEA-related molecules in human submandibular gland, gingival tissue and saliva (9, 10). Saliva is a mucosal fluid pro-

Institute for the Application of Nuclear Energy-INEP, Banatska 31b,

11080 Zemun-Belgrade, Yugoslavia

e-mail: sneza@inep.co.yu

duced by secretion of different glands (11). It protects oral cavity against harmful external agents (12). Periodontal disease affects the periodontium and includes both gingivitis and periodontitis. Pathogenesis of chronic gingival inflammation and periodontitis is complex and still not fully recognized (13, 14). The most accepted idea is that putative pathogenic bacteria induce periodontal disease by releasing various proteolytic enzymes and by provoking an immune response that triggers host cells for the expression of degrading enzymes (15). Since CEA possesses homoand hetero-cell adhesion properties, including bacterial recognition, we suppose that it could play a role in the maintenance of the structure and function of oral epithelium as well as in the related pathological processes (16 18).

The aim of this pilot study was to examine wheather locally produced *i.e.* salivary CEA could be a potential marker of the alterations of periodontium. In this study we report on the results of estimation of CEA concentration in the saliva of healthy individuals and patients with various stages of inflammation of oral tissues. This topic have not been extensively studied so far, and simple and non-invasive collection of saliva favored its choice as a clinical specimen in comparison to other traditional diagnostic body fluids.

207

Address for correspondence

Snežana Golubović

## **Materials and Methods**

#### Materials

Monoclonal anti-CEA antibodies, capture ( $IgG_1$ ,  $K_a = 3 \times 10^{10}$  mol <sup>1</sup>×L) and tracer (IgG1,  $K_a = 2 \times 10^{10}$  mol <sup>1</sup>×L), were purchased from Medix Biochimica (Kauniainen, Finland). Radioiodine <sup>125</sup>I, was from Radioisotope Centre Polatom (Otwock-Swierk, Poland). Carcinoembryonic antigen was from Medix Biotech. Inc. (Foster City, USA). Bovine serum albumine (BSA) was from Sigma (St. Louis, USA). All other chemicals were reagent grade.

#### Subjects

This study was carried out on a group of consenting patients and healthy volunteers (control group) seen at Department of Paradontology and Oral Medicine, University School of Stomatology, Belgrade, Yugoslavia. The status of periodontal tissue was assessed using the following indices: plaque index (Silnes and L e), calculus surface index (Green), gingival index (L e and Silnes), papillary bleeding index (Cowell) and tooth mobility index as described (19). According to the values of periodontal indices 19 patients with periodontal diseases were divided in three groups (stage I III). In addition, the separate group of 18 patients with diagnosis of necrotizing ulcerative periodontitis was examined before and after treatment (250 mg four times per day; five days) with metronidasole (Orvagil, Galenika).

The control group included 11 subjects without any signs of periodontal disease.

#### Collection of saliva and serum

Both saliva and blood were taken from control and patients groups. Whole saliva was collected by spitting, without any stimulation. All samples were taken till 11 a.m. Saliva was cleared by centrifugation (11000×g; 20 min) followed by dialysis against physiological saline. Peripheral venous blood was drawn and serum is separated by centrifugation after 30 60 min. The corresponding samples were used immediately or stored at 20 ° C until processed.

#### Analytical procedures

Carcinoembryonic antigen (CEA) concentration was determined using immunoradiometric assay, IRMA CEA (INEP, Yugoslavia), standardized against the 1st International Reference Preparation of CEA 73/601. All probes were done in triplicate. Samples were tested undiluted and diluted (1/10 and 1/100). The diluent was 0.05 mol/L phosphate buffer saline (pH 7.4) containing 10 g/L BSA. Radioactivity was measured using ISOMEDIC 4/600 gamma counter (ICN, USA). Protein concentration was determined according to Lowry (20) with bovine serum albumin as a standard. Optical density was measured using double beam spectrophotometer CE 594 (CECIL, England).

#### Statistical analysis

Kruskal-Wallis one way analysis of variance and Mann-Whitney (I test were used to analyze the obtained results. The data for each group were averaged and mean, median and standard deviations were calculated. Differences between groups were considered as statistically significant at p < 0.05.

#### Results

CEA concentrations of the examined saliva samples are presented in *Table I*. The salivas from periodontally healthy subjects revealed CEA concentrations with a median value of  $62 \mu g/L$  in the range from 23 102  $\mu g/L$ . Generally, salivary CEA concentrations did not exceed 102  $\mu g/L$ . On the contrary, distribution of salivary CEA concentrations in patients with periodontal diseases were very broad with median values: 74  $\mu g/L$  (stage I), 84  $\mu g/L$  (stage II), 240  $\mu g/L$  (stage III) and 412  $\mu g/L$  (NUP group). Marked individual variations were noticed in each group tested, especially in the group of patients with necrotizing ulcerative periodontitis (NUP). In spite of this, the scatter diagram indicates that most of the patients (29 of 37) have no overlap with the control range. Analysis of the

Table I Salivary CEA and protein concentrations in healthy subjects and periodontal patients

	Salivary CEA			Protein		
Subjects	(µg/L)*		concentrations (g/L)**			
	Median		Range	Median		Range
Healthy	62	23	102	1.07	0.70	1.47
(n=11)						
Periodontal	74	13	160	0.79	0.60	1.37
l stage						
(n=6)						
Periodontal	84	30	2600	0.90	0.66	1.37
ll stage						
(n=8)						
Periodontal	240	150	1200	0.93	0.70	1.65
III stage						
(n=5)						
NUP	412	110	4000	0.93	0.60	2.50
(n=18)						
				1		

 Difference between groups is statistically significant, p<0.05 (Kruskal-Wallis analysis of variance; H=25.9648; γ>c<sup>2</sup>);

\*\* Difference between groups is not statistically significant, p > 0.05 (Kruskal-Wallis analysis of variance; H=1.133;  $\chi < c^2$ ); (n, number of samples; p, level of significance).

	Control	Patients					
	Healthy	Periodontal	Periodontal	Periodontal	NUP		
Saliva*	(n=11)	l stage	II stage	III stage	(n=18)		
		(n=6)	(n=8)	(n=5)			
	1.06 ± 0.258	0.84 ± 0.228	0.97 ± 0.240	1.04 ± 0.354	1.13 ± 0.480		
Serum	N.D.	N.D.	N.D.	N.D.	N.D.		

Table II Salivary CEA and serum CEA concentrations in healthy subjects and periodontal patients

Values are means  $\pm$  S.D. Salivary CEA concentrations are expressed as mg CEA/mg total proteins. \*Difference between groups is statistically significant, p<0.05. N.D. non-detected. CEA concentrations in serum were below assay lowest standard (< 2  $\mu$ g/L).

Table III Salivary CEA and protein concentrations in NUP patients before and after treatment with metronidasole

NUP patients	Salivary CEA (µg/L)		P**	Protein (mg/mL)		p***	
	Median	Range		Median	Range		
I before treatment	410	130 4000	< 0.05	0.48	0.60 2.50	>0.05	
II after treatment*	200	67 1100		0.21	0.60 1.45		
* Patients were treated with metronidasole as described in Methods (n, number of samples; p, level of significance). ** Difference in salivary CEA concentrations is statistically significant (Man-Whitney U test; $U = 39 U_{0.05} = 45 U < U_{0.05}$ ). *** Difference in salivary protein concentrations is not statistically significant (Man-Whitney U test; $U = 53$ , $U > U_{0.05}$ ).							

obtained values by Kruskal-Wallis analysis of variance as well as by Mann-Whitney (I test indicated statistical-

ly significant (p<0.05) increase in salivary CEA concentrations in subjects with periodontal diseases.

In addition to CEA, total salivary proteins were also determined (*Table I*). There were no statistically significant (p>0.05) differences in protein concentrations between the examined groups. When CEA concentrations, expressed as mg CEA/mg total protein, were compared, statistically significant increase in patients groups were also observed (*Table II*). In relation to this, it is important to notice that CEA concentrations in sera of all examined subjects were below 2 µg/L, excluding the contribution of serum exudate to the salivary CEA concentration (*Table II*).

The salivary CEA concentrations, before and after the metronisadole treatment, in patients with NUP are presented in *Table III*. Mann-Whitney analysis of variance have not shown statistically significant difference in protein concentration (p > 0.05) after the treatment. However, it leads to statistically significant (p < 0.05) decrease in salivary CEA concentrations. The decrease was observed in all examined saliva samples from NUP patients.

#### Discussion

The results presented in this paper demonstrated statistically significant increase in salivary CEA concentrations in patients with inflammatory periodontal disease in comparison to healthy control group. For accurate determination of CEA and for proper validation of the results obtained, two major issues were carefully addressed. First, the selected capture antibody has no cross-reactivity with human CEA-related molecules, ensuring specific recognition of CEA molecule (10, 21). Second, the control group, used as a reference, included age-matched subjects screened on almost complete absence of gingival inflammation and periodontal breakdown. This was very important in relation to reported large variation in saliva composition between healthy individuals, in general (15).

Thus, salivary CEA concentration correlated with stage of periodontal disease, in contrast to total salivary protein concentration which remains in almost the same range. The examined groups were constituted on the basis of well defined criteria for severity of tissue damage (19). The lowest mean salivary CEA concentration ( $62 \mu g/L$ ) was detected in periodontally healthy individuals and the highest mean salivary CEA concentration ( $904 \mu g/L$ ) in patients with NUP. Comparison of paired saliva and serum samples indicated that CEA response in patients is locally produced, since in all cases serum CEA concentrations were below cut-off value.

The periodontal tissues are among the most biologically active in the body and the balance between effector-molecule induced tissue breakdown and tissue formation is the essence of periodontal health (23). Pignatelli and co-workers, demonstrated that CEA can function as an accessory cell adhesion molecule, mediating cell-matrix interaction, and they proposed that the detection of high levels of CEA in colorectal cancers may be only an epiphenomenon resulting from the disruption of cell-matrix interaction (17). Therefore, one of the hypotheses to explain high salivary CEA level in patients with NUP could be enhanced tissue damage. This might be also related to the results obtained in the group of patients before and after metronidasole treatment. Thus, the administration of metronidasole which acts as anti-inflammatory agent, leads to statistically significant decrease in salivary CEA concentration in all examined subjects.

Finally, the results obtained indicated salivary CEA as potentially useful adjunct for diagnosis and follow-up of patients with periodontal diseases. CEA, such as most salivary molecules is a multifunctional molecule and it may have both protective and detrimental properties (14). The proposed function of CEA, based on *in vitro* studies, such as bacterial ligand (18), suppressive factor to the host immunocompetent cells (23), and innate immunity factor (8), suggested that salivary CEA deserves further studies not only in clinical and diagnostic purposes but also in fundamental terms.

Acknowledgments. This work was supported by the Ministry of Science, Technologies and Development of the Republic of Serbia, project code 1504: Glycobiological aspects of physiological and pathophysiological processes.

# SALIVARNI KARCINOEMBRIONALNI ANTIGEN KAO INFLAMATORNI MARKER

Snežana Golubović<sup>1</sup>, Ljiljana Janković<sup>2</sup>, Aris Movsesijan<sup>2</sup>, Žanka Bojić<sup>1</sup>, Miroslava Janković<sup>1</sup>

<sup>1</sup>Institut za primenu nuklearne energije–INEP, Banatska 31b, 11080, Beograd, Jugoslavija <sup>2</sup>Klinika za paradontologiju i oralnu medicinu, Stomatološki fakultet, Univerzitet u Beogradu, Jugoslavija

*Kratak sadržaj:* U ovom radu je uporedno ispitana koncentracija karcinoembrionalnog antigena (CEA) u uzorcima salive i seruma zdravih osoba i osoba sa periodontalnim oboljenjem. Koncentracija CEA je određena imunoradiometrijskom metodom zasnovanoj na upotrebi specifičnih anti-CEA antitela. Medijalna vrednost salivarnog CEA u zdravih osoba bila je 62  $\mu$ g/L. Medijalne vrednosti salivarnog CEA u pacijenata sa periodontalnim oboljenjem, distribuirane u širokom opsegu koncentracija, bile su: 74  $\mu$ g/L (I stadijum), 84  $\mu$ g/L (II stadijum), 240  $\mu$ g/L (III stadijum) i 412  $\mu$ g/L (nekrotični ulcerativni periodontitis NUP). Analiza dobijenih vrednosti ukazala je na statistički značajno uvećanje salivarnog CEA u pacijenata sa NUP dobijeno je po tretmanu sa metronidazolom. Dobijeni rezultati ukazali su na salivarni CEA kao potencijalni marker periodontalnih promena.

Ključne reči: CEA, saliva, serum, periodontalno oboljenje, protein, imunoradiometrijski test.

## References

- Gold P, Freedman SO. Specific carcinoembryonic antigens of the human digestive system. J Exp Med 1965; 122: 467 81.
- Thompson JA. Molecular cloning and expression of carcinoembryonic antigen gene family members. Tumor Biol 1995; 16: 10 6.
- Beauchemin N, Benchimol S, Cournoyer D, Fuks A, Stanners CP. Isolation and characterization of full-length functional cDNA clones of human carcinoembryonic antigen. Mol Cell Biol 1987; 7: 3221 30.
- 4. Brandriff BF. Order and genomic distances among members of the carcinoembryonic antigen (CEA) gene family determined by fluorescence in sity hybridization. Genomics 1992; 12: 773 79.
- 5. Beauchemin N, Draber P, Dveksler G, Gold P, Gray-Owen S, Grunert F et al. Nomenclature announcement.

Redefined nomenclature for members of the carcinoembryonic antigen family. Exp Cell Res 1999; 252: 243 49.

- Von Kleist S, Chavanel G, Burtin P. Identification of an antigen from normal human tissue that crossreact with the carcinoembryonic antigen. Proc Natl Acad Sci USA 1972; 69: 2492 94.
- 7. Svenberg T. Carcinoembryonic antigen-like substances of human bile. Isolation and partial characterization. Int J Cancer 1976; 17: 588 596.
- Hammarstr m S, Baranov B. Is there a role for CEA in inate immunity in the colon? Trends Microbiol 2001; 9: 119 125.
- Zoubir F, Khan WN, Hammarstr m S. Carcinoembryonic antigen gene family members in submandibular salivary gland: demonstration of pregnancy-specific glycoproteins by cDNA cloning. Biochem Biophys Res Commun 1990; 169: 203 216.

- Golubović S, Marović-Stojković V, Janković Lj. Immunohistochemical detection of carcinoembryonic antigen (CEA) in the epithelia of inflamed gingiva. Archive of Oncology 1997; 5: 61 4.
- Schenkels LCPM, Veerman ECI, Nieuw Amerongen AV. Biochemical composition of human saliva in relation to other mucosal fluids. Crit Rev Oral Biol Med 1995; 6: 161 75.
- Tabak LA. In defense of the oral cavity: Structure, biosyntesis and function of salivary mucins. Annu Rev Physiol 1995; 57: 547 64.
- Henskens YMC, Van der Velden U, Veerman ECI, Nieuw Amerongen AV. Protein, albumin and cystatin concentrations in saliva of healthy subjects and of patients with gingivitis or periodontitis. J Periodont Res 1993; 28: 43 8.
- Levine MJ. Salivary macromolecules: A structure/function synopsis. In: Malamud D, Tabak L, ed. Saliva as a diagnostic fluid. vol 694: Annals of the New York Academy of Sciences, 1993; 11 6.
- Henskens Y.M.C. Dissertation. Salivary cystatins and their relation to periodontal inflammation. Vrije Universitet, Amsterdam, Netherlands 1994.
- Benchimol S, Fuks A, Jothy S, Beauchemin N, Shirota K, Stanners CP. Carcinoembryonic antigen, a human tumor marker, function as an intercellular adhesion molecule. Cell 1989; 57: 327 34.

- Pignatelli M, Durbin H, Bodmer WF. Carcinoembryonic antigen functions as an accesory adhesion molecule mediating colon epithelial cell-colagen interactions. Proc Natl Acad Sci 1990; 87: 1541 45.
- Leusch HG, Hefta SA, Drzeniek Z, Humel K, Markos-Pusztai Z, Wagener C. Escherichia coli of human origin binds to carcinoembryonic antigen (CEA) and non-specific crossreacting antigen (NCA). FEBS Letters 1990; 261: 405 9.
- Janković Lj, Jelić S, Filipović-Lješković I, Ristović Z. Salivary immunoglobulins in cancer patients with chemotherapy-related oral damage. Oral Oncol Eur J Cancer 1995; 31B: 160 5.
- Lowry OH, Rosebrough NJ, Farr AL, and Randall RJ. Protein measurement with the Folin phenol reagent J Biol Chem 1951; 193: 265 75.
- Golubović S. Immunoradiometric and lectin assays for quantitative determination and characterization of the tumour markers chorionic gonadotropin and carcinoembryonic antigen. M.A. Thesis, Faculty of Pharmacy, University of Belgrade, 1996.
- Hefti AF. Aspects of cell biology of the normal periodontium. Periodontology 2000 1993; 3: 64 75.
- Prado IB, Laudanna AA, Carneiro CRW. Susceptibility of colorectal-carcinoma cells to natural-killer-mediated lysis: Relatioship to CEA expression and degree of differentiation. Int J Cancer 1995; 61: 854 60.

Received: October 29, 2002 Aceepted: January 23, 2003